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Woolf et al.

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(54) **HUMAN DRG11-RESPONSIVE AXONAL GUIDANCE AND OUTGROWTH OF NEURITE (DRAGON) PROTEINS AND VARIANTS THEREOF**

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patent is extended or adjusted under 35
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Related U.S. Application Data

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18, 2002.

(51) **Int. Cl.**
C07K 14/475 (2006.01)
A61K 38/18 (2006.01)

(52) **U.S. Cl.** **530/350; 514/2; 514/12**

(58) **Field of Classification Search** None
See application file for complete search history.

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(57) **ABSTRACT**

This invention features methods and compositions useful for treating and diagnosing diseases of the nervous system, retina, skin, muscle, joint, and cartilage using a Dragon family protein. Protein and nucleic acid sequences of human, murine, zebrafish, and *C. elegans* Dragon family members are also disclosed.

9 Claims, 20 Drawing Sheets

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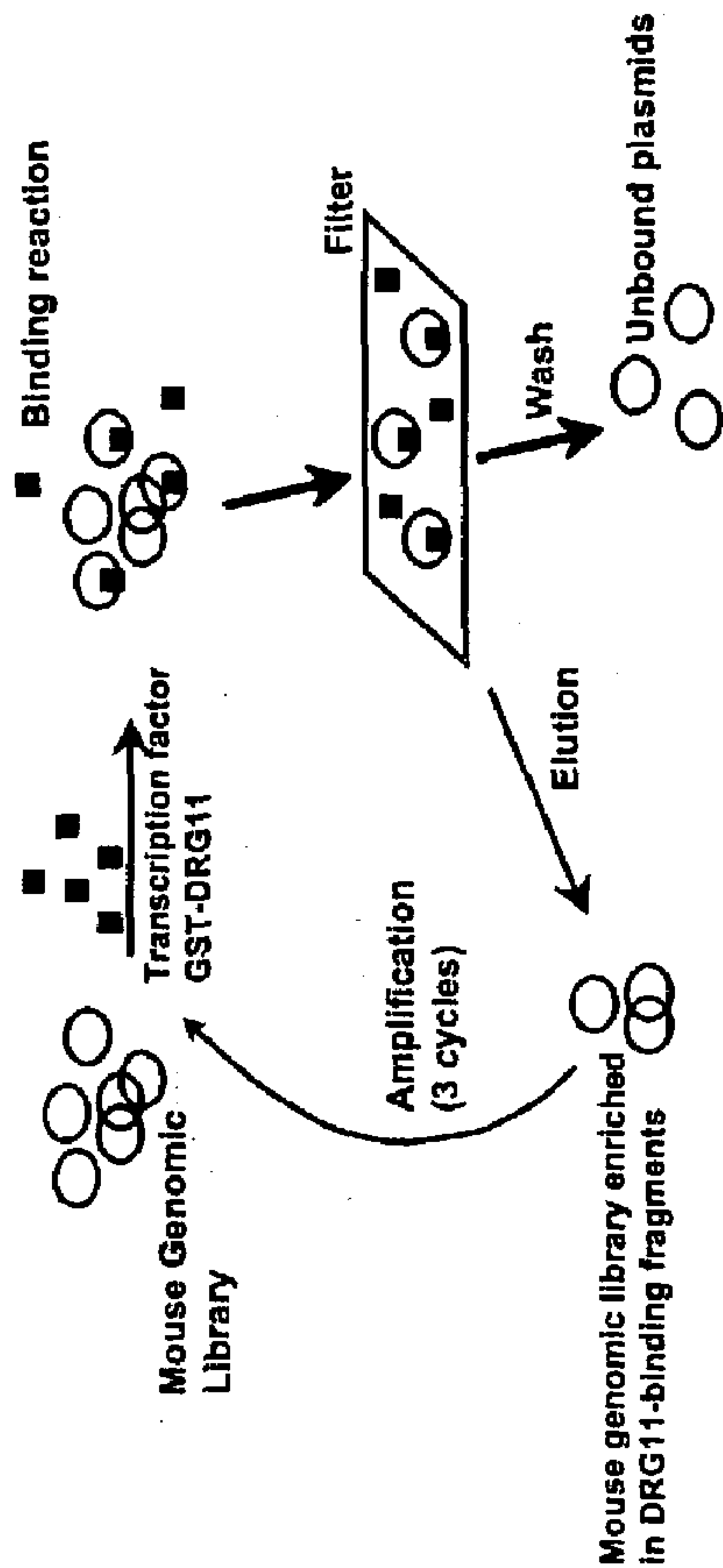
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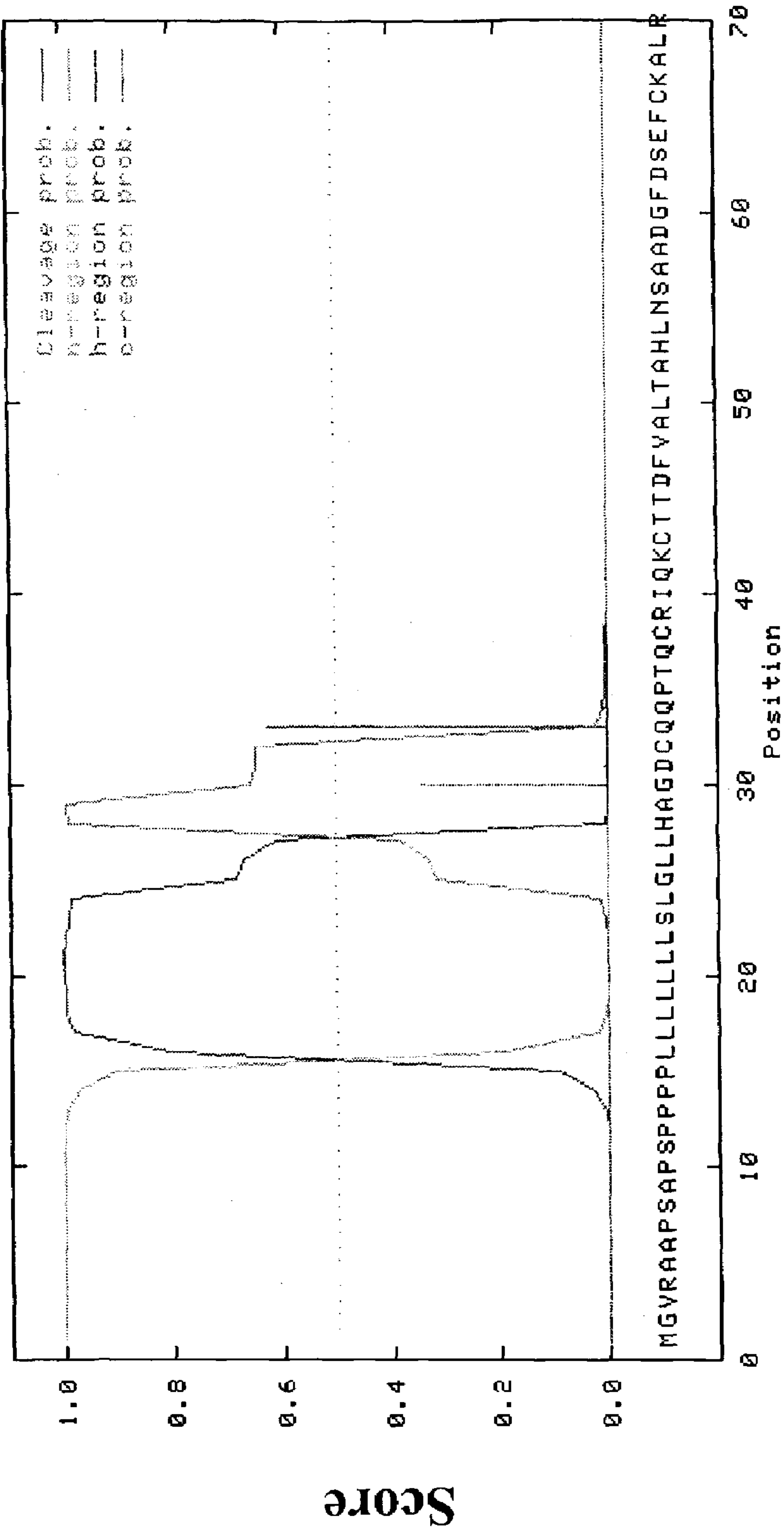
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[illegible]

DRAGON protein structure:

GPI-anchor

Figure 2



>Sequence
Prediction: Signal peptide
Signal peptide probability: 1.000
Signal anchor probability: 0.000
Max cleavage site probability: 0.628 at 33

Figure 3

mDRAGON	PSPPPPLLLLLLSLGLLHAGD-----CQQPTQCRIQKCTTDFVALT AHL
	P P P P P L L L L L L L A C T R + C V A A +
IGFBP	PLPPPPLLPLPLLLLLLGASGGGGARA E V L F R C P P C T P E R L A A C G P P V A P P A A V
	NSAADGF D S E F C K A L R -----A Y A G C T Q R T S K A C -----R G N L V Y --H S A V L G I S D L
	+ A G + + R + C + + A C + G Y + L + L
	AAVAGGARMPCAELVREPGCGCCSVCARLEGEACGVYT P R C G Q G L R C Y P H P G S E L P L Q A L
	-MSQRNCSKDGPTS-STNPEVTHDPCNYHSHGGVREH 134
	M + C K + P E D + H S G G + E +
	VMGEGTCEKRRDAEY GAS P E Q V A D N G D D H S E G G L V E N 164

Figure 4

mDRAGON	PPSPPP---PLL-LLLLSLGLLHAGDCQQPTQCRIQ-KCTTDFVALTAHLNSAAD---G
	PPSPPP PLL LLL L LL AG C+ + + K T +A T+H S + G
EPHB3	PPSPPPGLLPLLPLLPLLPLLPAG-CRALEETLMDTKWVTSELAWTSHPESGWEEVSG
	FDSEFCKALRAYAGCTQRTS 102
	+D E +R Y C R S
	YD-EAMNPIRTYQVCNVRES 86

Figure 5

mDRAGON PAAAGAEQSRRLPGLWPPSPPPPL-----LLLLLSLGLHAGDCQQPTQCRIQ-KCTTDSV
P A G + RRP + PP PPPP+ LLLLL G A C + C +CT
hNotch 3 PGARGRRRRRP-MSPPPPPPVRALPLLLLLAGPGA-APPCLDGSPCANGGRCTQLPS

ALTAHLNSAADGFDFSECKALRAYAGCTQRTSKACRGNLVYHSAVLGISDLMNQNCCKD
A L G+ E C+ C S C G V S+V+ + S R C +
REAACL--CPPGWVGERCQLEDP---CH---SGPCAGRGVCQSSVAGTAREFSCR-CPRG

--GPTSSTNPEVTHDPCNYHSHGGVREHGGDQR-----PPNY 157
GP S PC +HG R G D R PP Y
FRGPDCLPDCPLSSPC---AHGA-RCSVGPDGFRFLCSCPPGY 150

mDRAGON KSLHIVEKESGRYVEMHARYIGTTVFVRQLGRYLTLAIRMPEDLAMSYEESQDLQLCVNG
+ LH+ E+ ++E R + ++ + L + I D+ + +E +++
mPIP5K QKLHVGEESKKNFLEKLKRDVEFLAQLKIMDYSLLVGIH---DVDRAEQEEMEVE---ER

CPMSECIDDGQGQVSAILGHSPLPHTT--SVQAWPGY-----TLETASTQCHEKMPVK
EC +DG G S + + P + ++ ++P + +++ + + HE P K
AEEEECENDGVGG-SLLCSYGTPPDSPGNLLSFPRFFGPGFEFDPSPVDVYAMKSHESAPKK

DIYFQSCVFDLLTTGDANFTAA-----AHSALEDVEALHPRK--ERWHIFPSS 395
++YF + + D+LT DA AA H A ++ ++P + +R++ F S+
EVYFMA-IIDILTPYDAKKKAAHAAKTVKHGAGAEISTVNPEQYSKRFEFMSN 389

Figure 6

mDRAGON QKCTTDSVALTAHLNSAADGFDSEFCKALRAYAGCTQRTSKACRGNLVYHSAVLGSD--
+ C T AA G +E C + A GC + +G Y G D
THR1 ENCLTTYGCCPDGKGAAGHHNEGCGCVYAQYGCCPDGKTSAGAGFY-----GCPDSC

LMSQRNCSKDGTSSSTNPEVTHDPCNYHSHG-----GVREHGGDQRPNNYLF
SQ C DG T + PC Y +G G R G D R Y C
AQSQFGCCPDGKTPARGSHKEGCPQCQYTRYGCCPDGETTALGPRNDGCDDCRYAKYGCC

mDRAGON 60 QKCTTDFVALTAHLNSAADGFDSEFCKALRAYAGCTQRTSKACRGNLVYHSAVLGSDLM 119
+ C A S+ GF E C+ C QRT+ C GN H L I+ +
mSlit2 830 EPCHKKVCAGMCPSSQSGFTCE-CEEGWMGPLCDQRTNDPCLGNKCVHGTCLPINAES 888

120 SQRNCSK-DGPTSSSTNPEVTHDPCNYHS--HGGVREHGGDQRPNNYLFCCGLFGD 171
C + G E +PC HG R G G +P G+ GD
889 YSCKCLEGHGGVLCDEEEDLFNPCQMIKCKHGKRLSGVG--QPYCECNSGFTGD 941

Figure 7

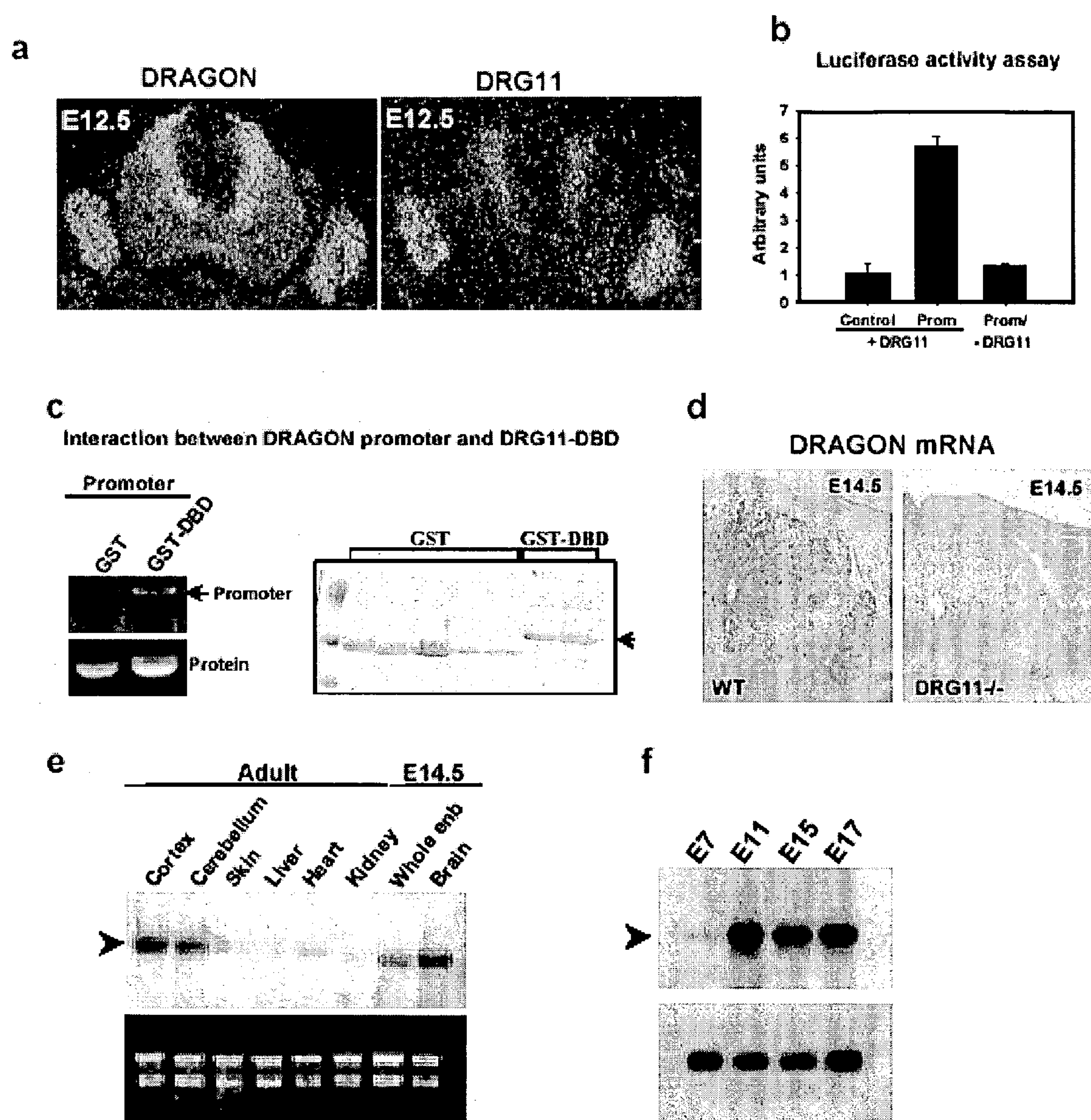


Figure 8

a		b	
DRAGON	MGVRAAPSCAAAPAAAGAEQSPFGLMFPSPPEPLLLLLLSGLHAGDQOQPTQCRIO	Human	MGLEAAPSSARAAARAEVEQERRRPGLCPP--PHEHLLLPSPSLGLHAG--DCQQAQCRI
DL-2	-----MQPERRLVVTGAGWGMGRCAGRSALGHMPTLAFGLCSFPALISPCRIIL	Mouse	MGVPAASCAAPAAAGAEQERRRPGLCPPSPPEHLLLPSPSLGLHAG--DCQQAQCRI
DL-1	-----MGCSPSPSPHGSPTTSTLLTLLLLCGGAHSONKIL	Zebrafish	-----NGMFRAGSYYPGAEERLISPLHLVLCCTLSSPTIESQVYTPQCRI
DRAGON	KCTTDFVALTAHL-----NSAADPEES--EECKALRAVAGCTORTSKARGNLVH	Human	OKCTTDFVSE--TSHLNSAVDGFSEFCCKALRAVAGCTORTSKARGNLVHSAVLGISDLA
DL-2	SHKNSSEFATSSG-----SHAPASDVPPEFALRNYAGCTRPTATPCRGDIAYH	Mouse	OKCTTDFVALTAHLNSAADGDPDSEFCCKALRAVAGCTORTSKARGNLVHSAVLGISDLA
DL-1	ECNAEIVSSELLHRRGGSPDTPRGGGRGSLAEGGLRALPSVALTTRTASTCRGDIAYH	Zebrafish	OKCTTDFVSE--TSHLNSPSLDGFDCEFCCKALRAVAGCTORTSKARGNLVHSAVLGISDLA
DRAGON	SAVLSESLDMSQRUCSKDGPTSTNPEVTHDECNVISHGG-----VREHGGDQRPPT	Human	SQRNCSKDGPTSTSTNPEVTHDPCNYHSHAGAKREHPRGDPAPPS-----LFCGLFGDPHLE
DL-2	SAVHGSEDLMSCHNCSKDGPTSQRRVETLPPAGDSQSRSDSPETCHYKSTKHSAANY	Mouse	SQRNCSKDGPTSTSTNPEVTHDPCNYHSHAGAKREHPRGDPAPPS-----LFCGLFGDPHLE
DL-1	SAVHSEEDLMTTHNCSROGPTAPAPAGPALGAGAPLTPDPCDYEARFRLHGRAGE	Zebrafish	SQRNCSKDGPTSTSTNPEVTHDPCNYHSHAGAKREHPRGDPAPPS-----LFCGLFGDPHLE
DRAGON	LFEGFLGDPHLRTFKEHFOCTCKVEGAWPLLDNNYLSQMTNVPVVPSSATATNKVTILF	Human	TFKDNFOTCKVEGAWPLLDNNYLSQVNTNVPVVPSSATATNKVTILFKEHHECTDQKVY
DL-2	THGLFGDPHLRTFKEHFOCTCKVQGAWPLLDNNYLSQMTNVPVVPSSATATNKVTILF	Mouse	TFKDNFOTCKVEGAWPLLDNNYLSQVNTNVPVVPSSATATNKVTILFKEHHECTDQKVY
DL-1	LHVASPEDPHVRSERHOEHCHQVQGAWPLLDNNYLSQMTNVPVVPSSATATNKVTILF	Zebrafish	TFKDNFOTCKVEGAWPLLDNNYLSQVNTNVPVVPSSATATNKVTILFKEHHECTDQKVY
DRAGON	FAOHECTQKVKYQVTDLPAAFDVGTSGGJD-VKSLHVEHESRYVENHARYIGTT	Human	QAVTDDLPAAFVDTGTSGGSDAKSPIVERESCHYVENHARYIGTAVEMRGVRYLTDA
DL-2	HFQCEQVQKVKYQVTDLPAAFDVGTSGGJDVKSDFIVEPESRYVENHARYIGTAVEMRGVRYLTDA	Mouse	QAVTDDLPAAFVDTGTSGGSDAKSPIVERESCHYVENHARYIGTAVEMRGVRYLTDA
DL-1	HFQCEQVQKVKYQVTDLPAAFDVGTSGGJDVKSDFIVEPESRYVENHARYIGTAVEMRGVRYLTDA	Zebrafish	QAVTDDLPAAFVDTGTSGGSDAKSPIVERESCHYVENHARYIGTAVEMRGVRYLTDA
DRAGON	FEYRQMGRYLTALRNPEDLMSSEB--SODLQLCVNGCPMSFCDDGGOVSAIILCHSL	Human	IRMPEDLMSSEB--SODLQLCVNGCPMSFCDDGGOVSAIILCHSL-----RSLVQAW
DL-2	EVYRQMGRYLTFAVRHPEEIVNAVEDRDSGLYLCURSCPLNQDFQAFRANRESRPP	Mouse	IRMPEDLMSSEB--SODLQLCVNGCPMSFCDDGGOVSAIILCHSL-----RSLVQAW
DL-1	EIRTAGQLSESENAVEDRANESA--EEDLQLCVNGCPMSFCDDGGOVSAIILCHSL	Zebrafish	IRMPEDLMSSEB--SODLQLCVNGCPMSFCDDGGOVSAIILCHSL-----RSLVQAW
DRAGON	PHTTSVQAWPS-VTETASTQHEKMPVKDIYFQSCVFDLLTTGDANFTAAHSALEVE	Human	PBYTLETANTQCHERMPVKDIYFQSCVFDLLTTGDANFTAAHSALEVEDVACHPRKERH
DL-2	AAASPSVVPETPYETAVAKKEKLPVEDHYVQAVFEDLITGQVVFETAAVYALBCK	Mouse	PBYTLETASTQCHERMPVKDIYFQSCVFDLLTTGDANFTAAHSALEVEDVACHPRKERH
DL-1	-----RATAEDTARRLSKEGLPVEDAYFQSCVFDLSVSCDPHETVAOTATDAR	Zebrafish	GVETTESASRRRDQEMAKDIYFQSCVFDLLTTGDANFTAAHSALEVEDVACHPRKERH
DRAGON	AHPKPRKPRMIFP-----SSCGGCRDIPVGLGLTCELIIMEL-----	Human	IFPSCNGTTPRAGSDSVSGLTCELIIMEL
DL-2	MHSNVDKLHLRPTRELPGAVAAAATTFBAPQ--LLGTPLLVLPVFM--	Mouse	IFPSCCG-----CRDLPVGLGLTCELIIMEL
DL-1	IFLTEDENMLFE-----SDAGPPDSPA-CLVPEHSALEFLWCFKSK	Zebrafish	IFPSCSRLSPSLLTALSSFLP-----

Figure 9

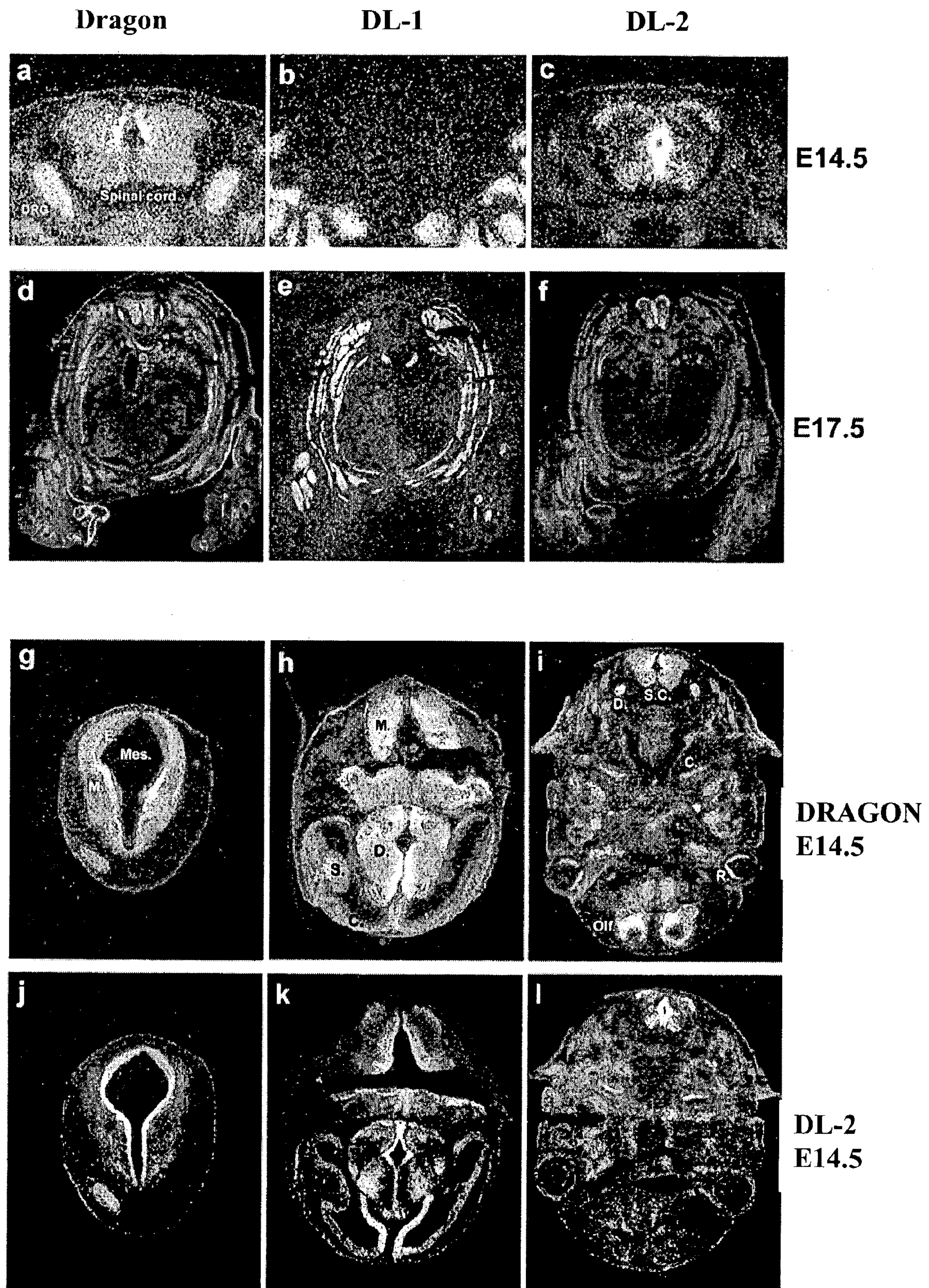


Figure 10

**DRAGON
(mRNA)
Adult rat
DRG**

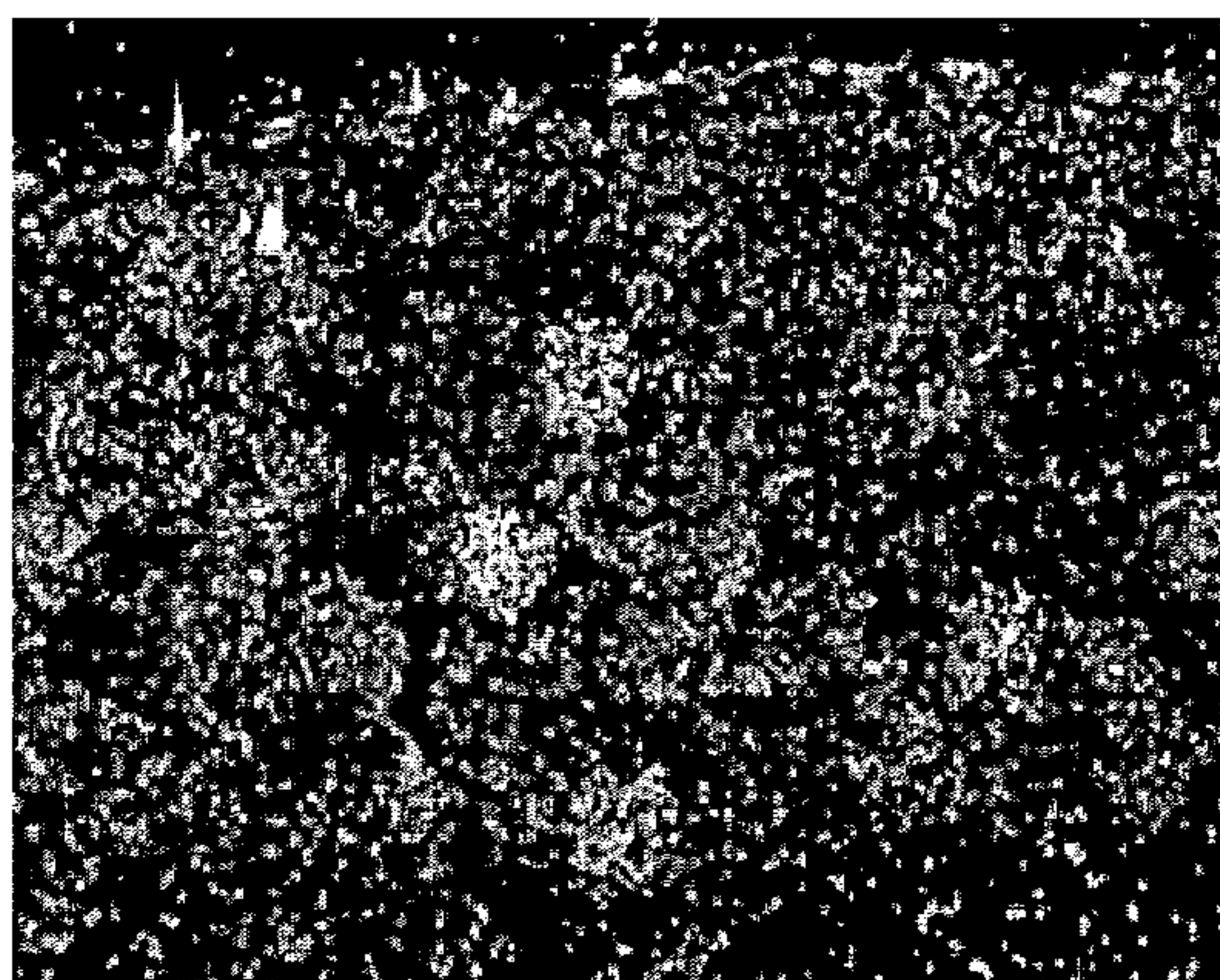
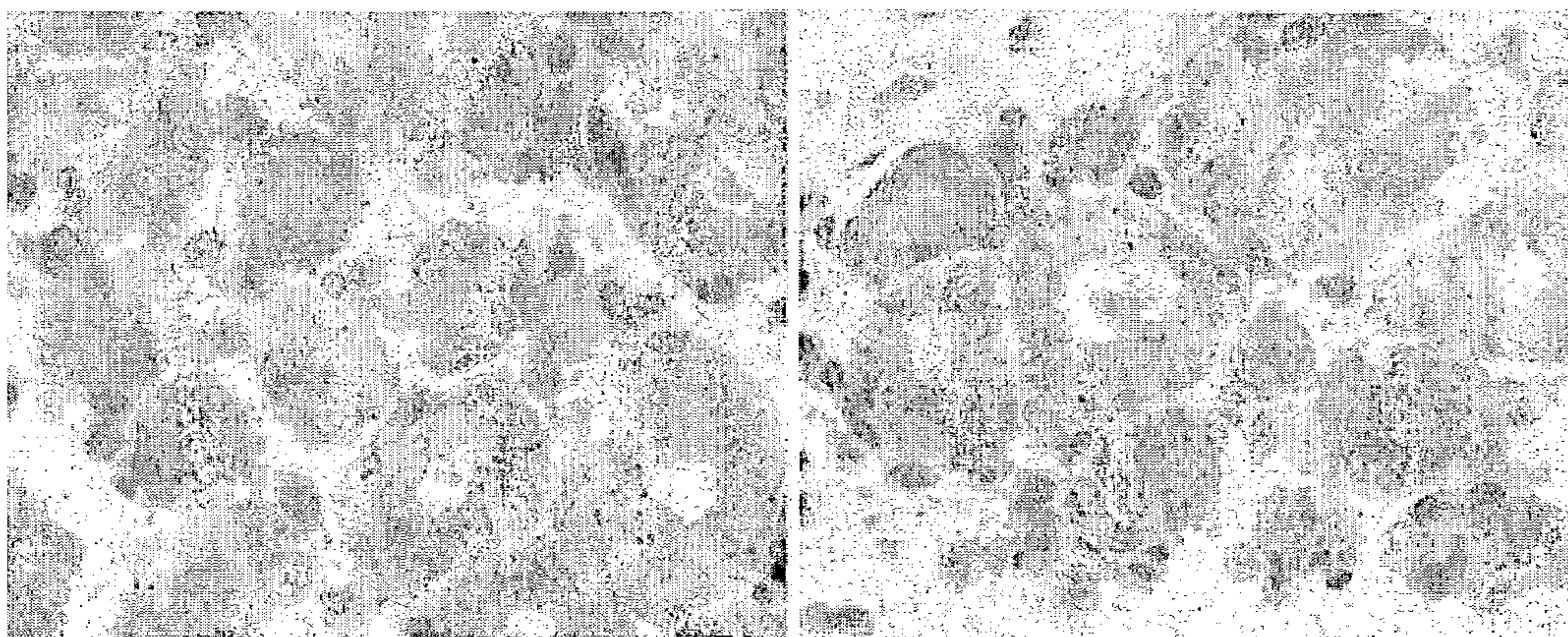
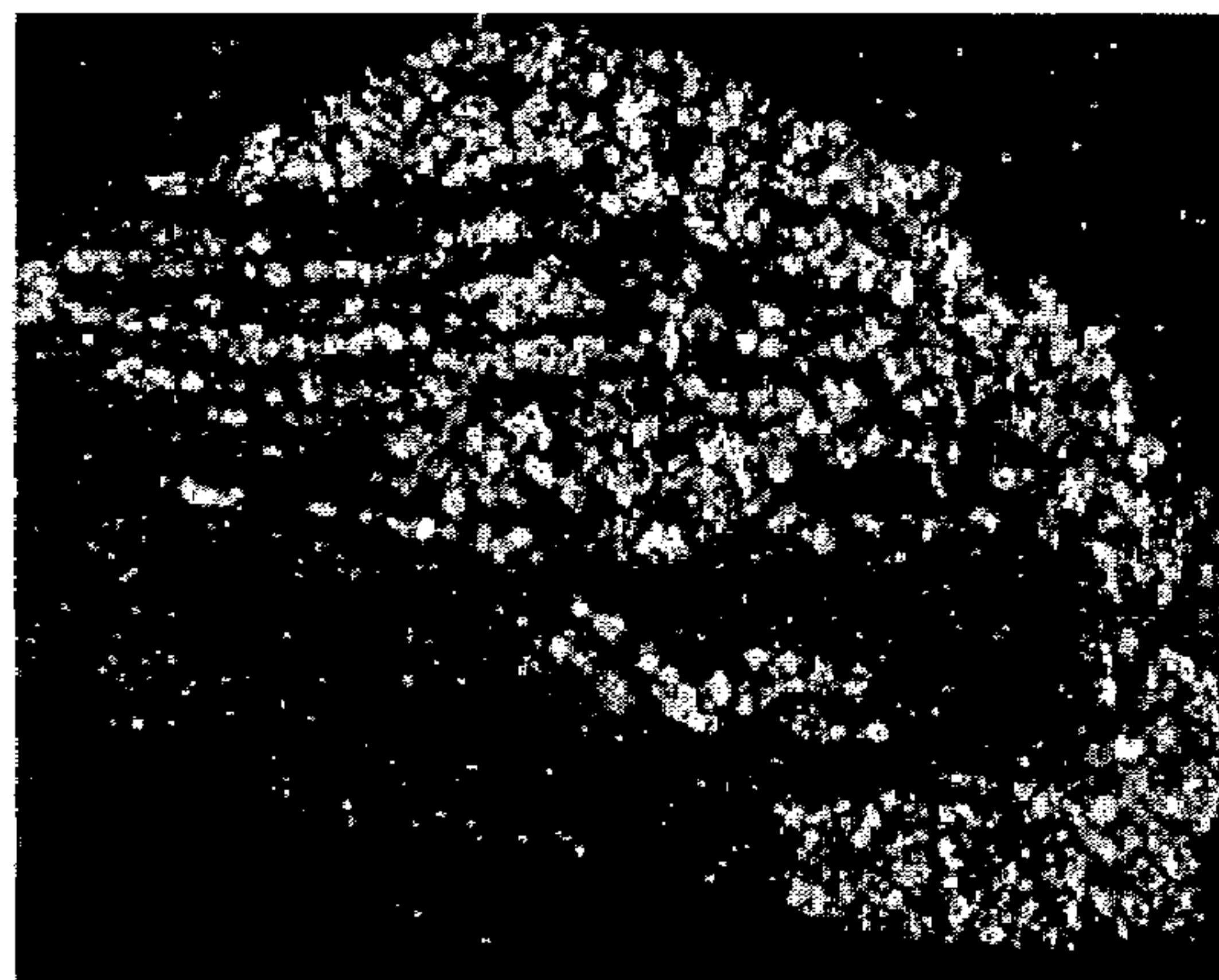


Figure 11



Figure 12



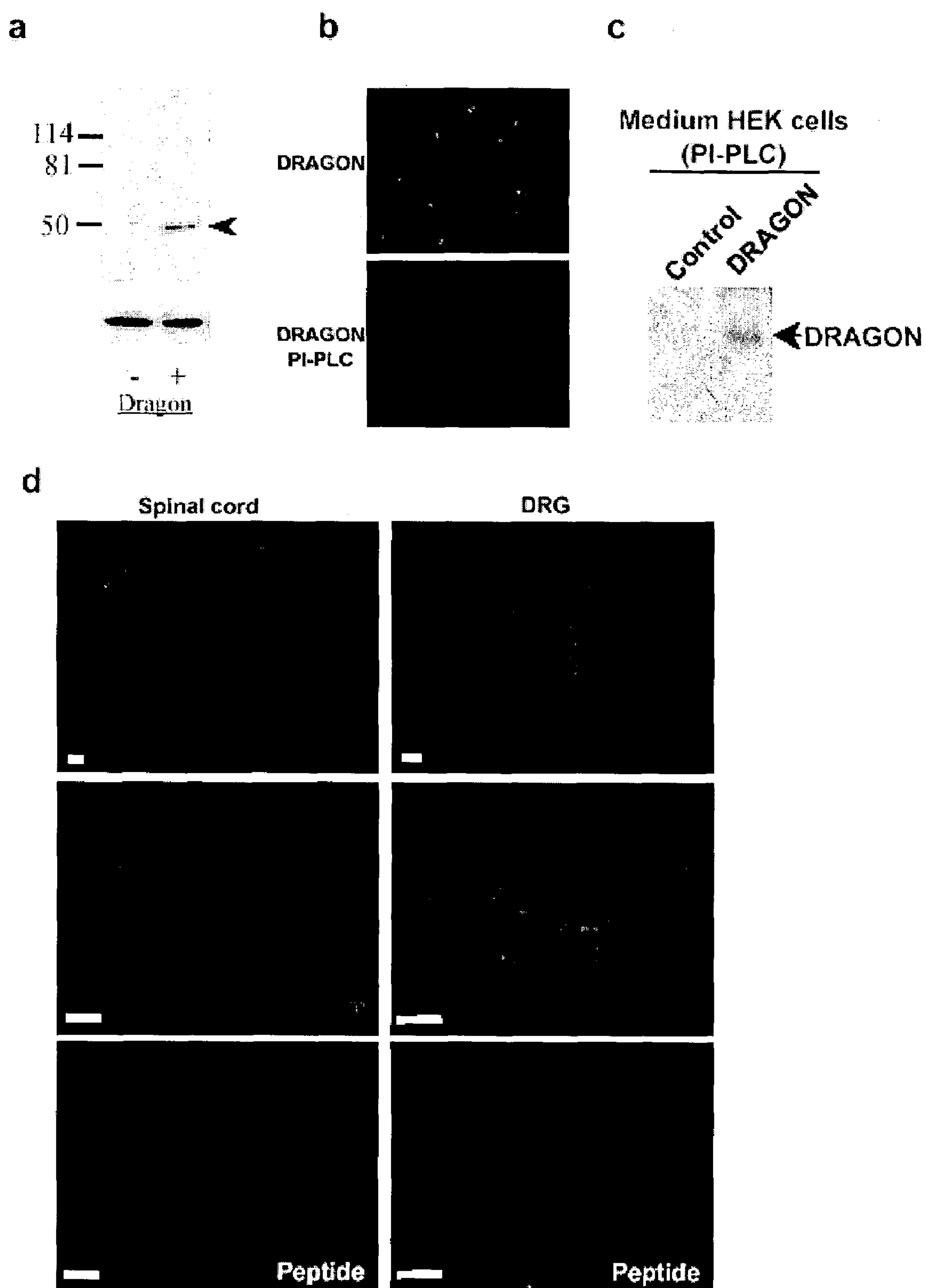
Figure 13

Figure 14

Cell adhesion assay

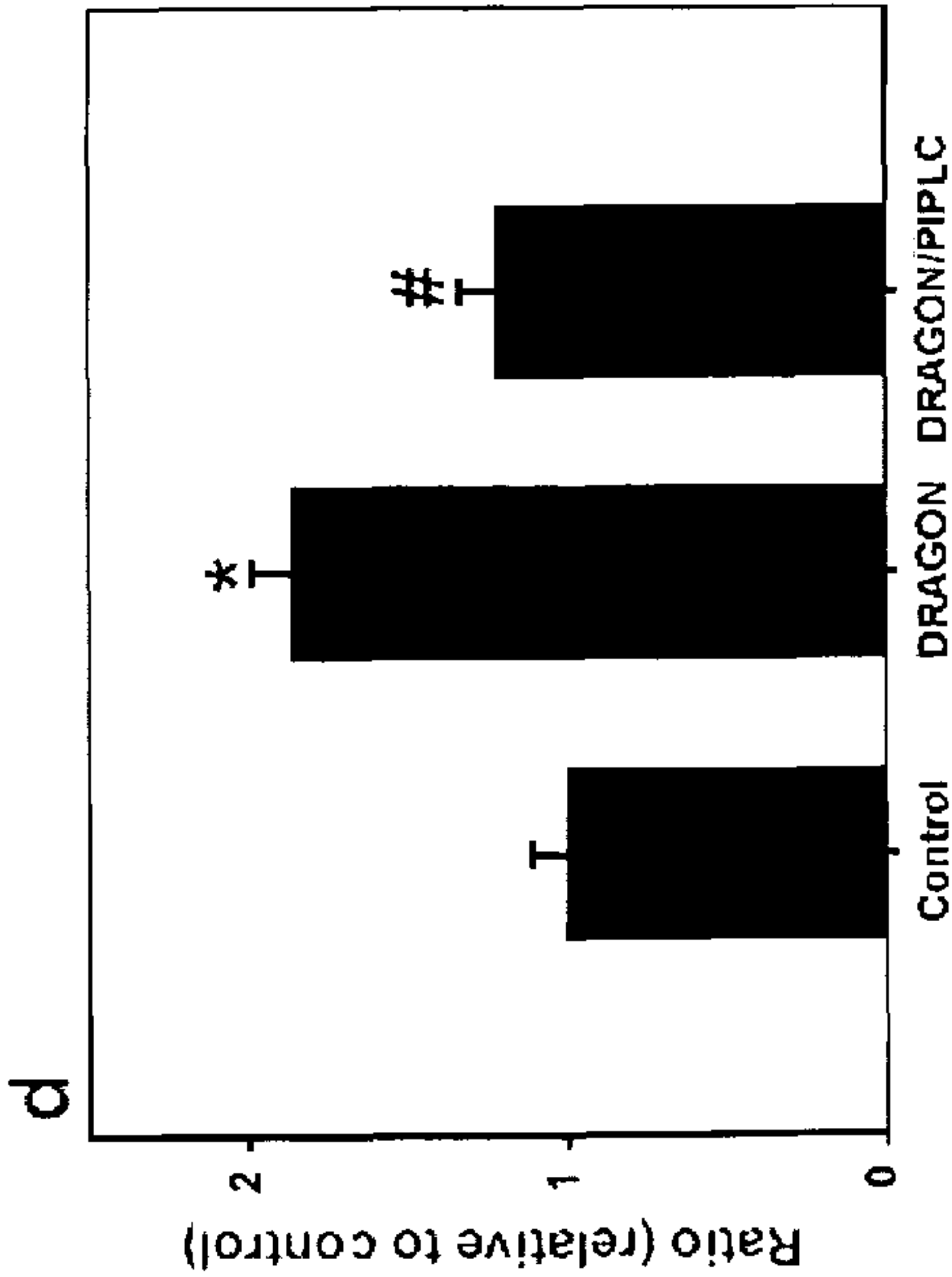
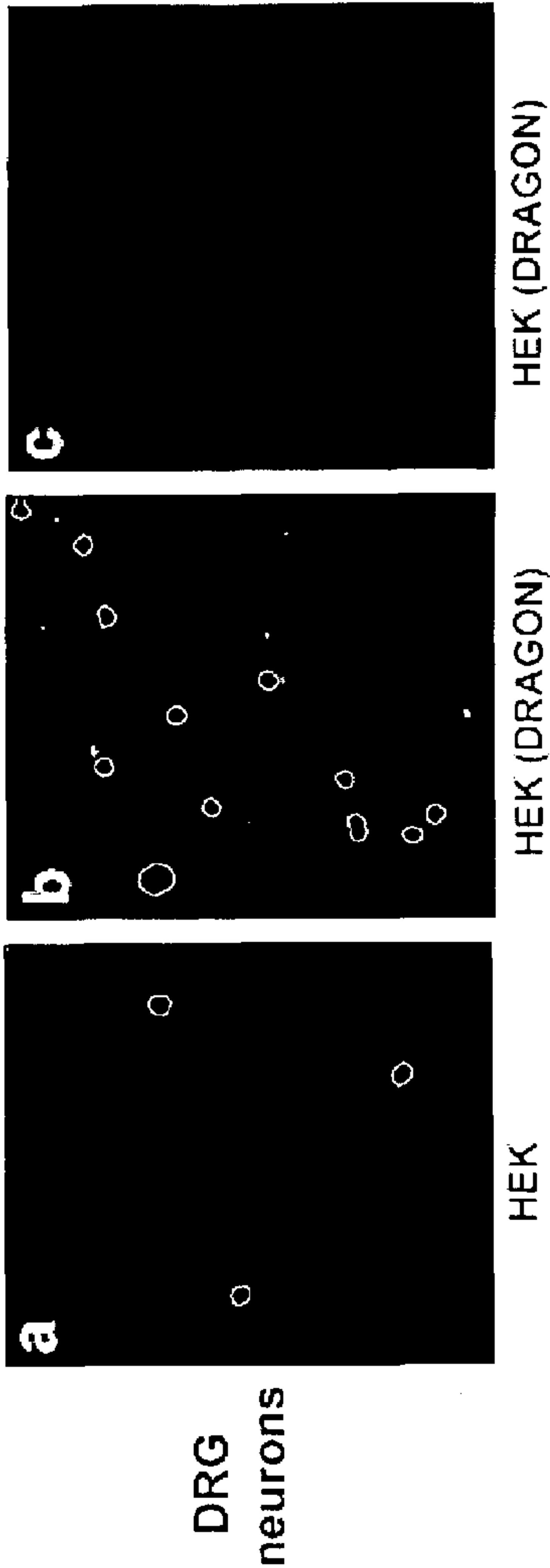
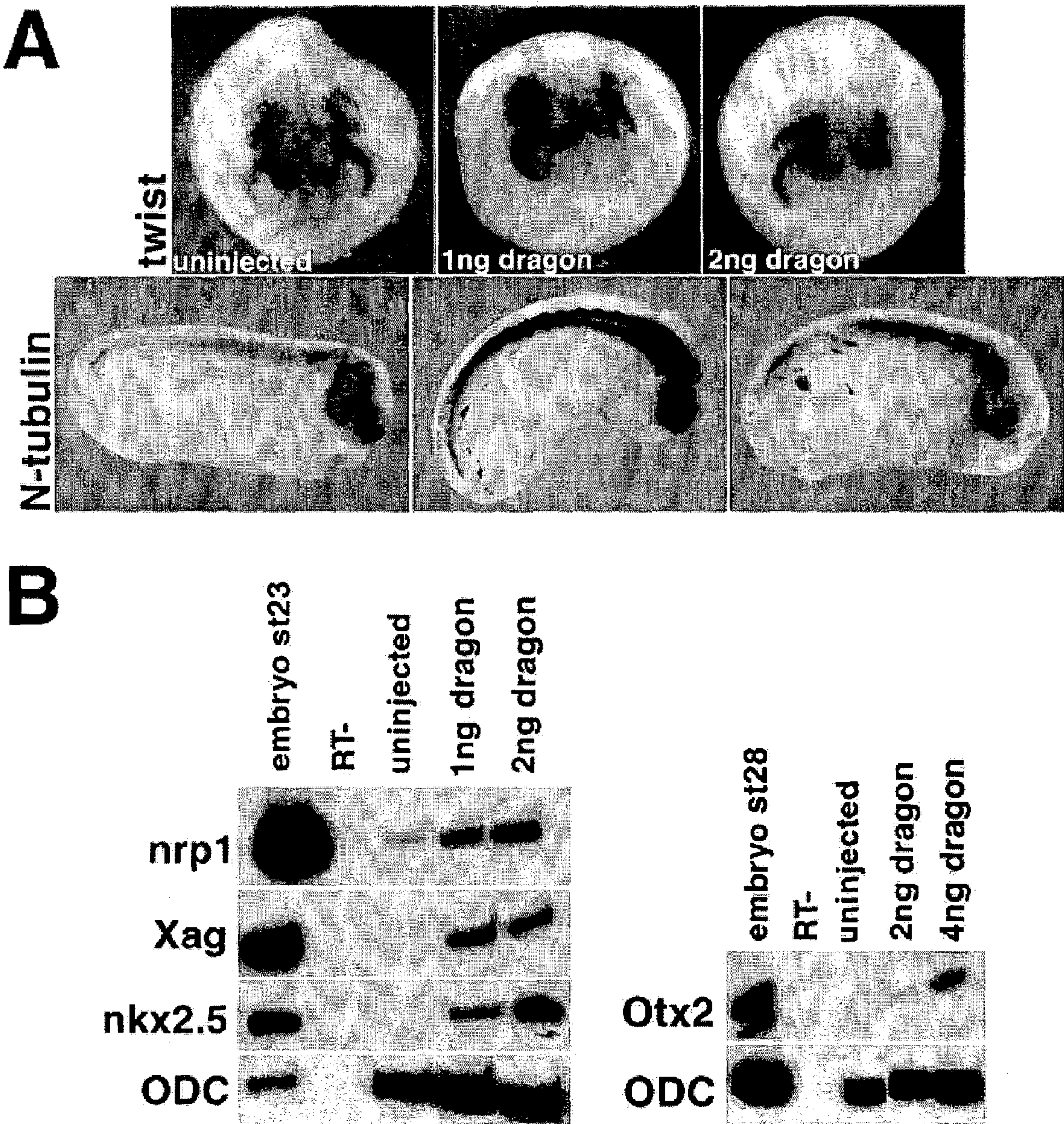


Figure 15



Northern blot (*dragon*)

FIGURE 16A

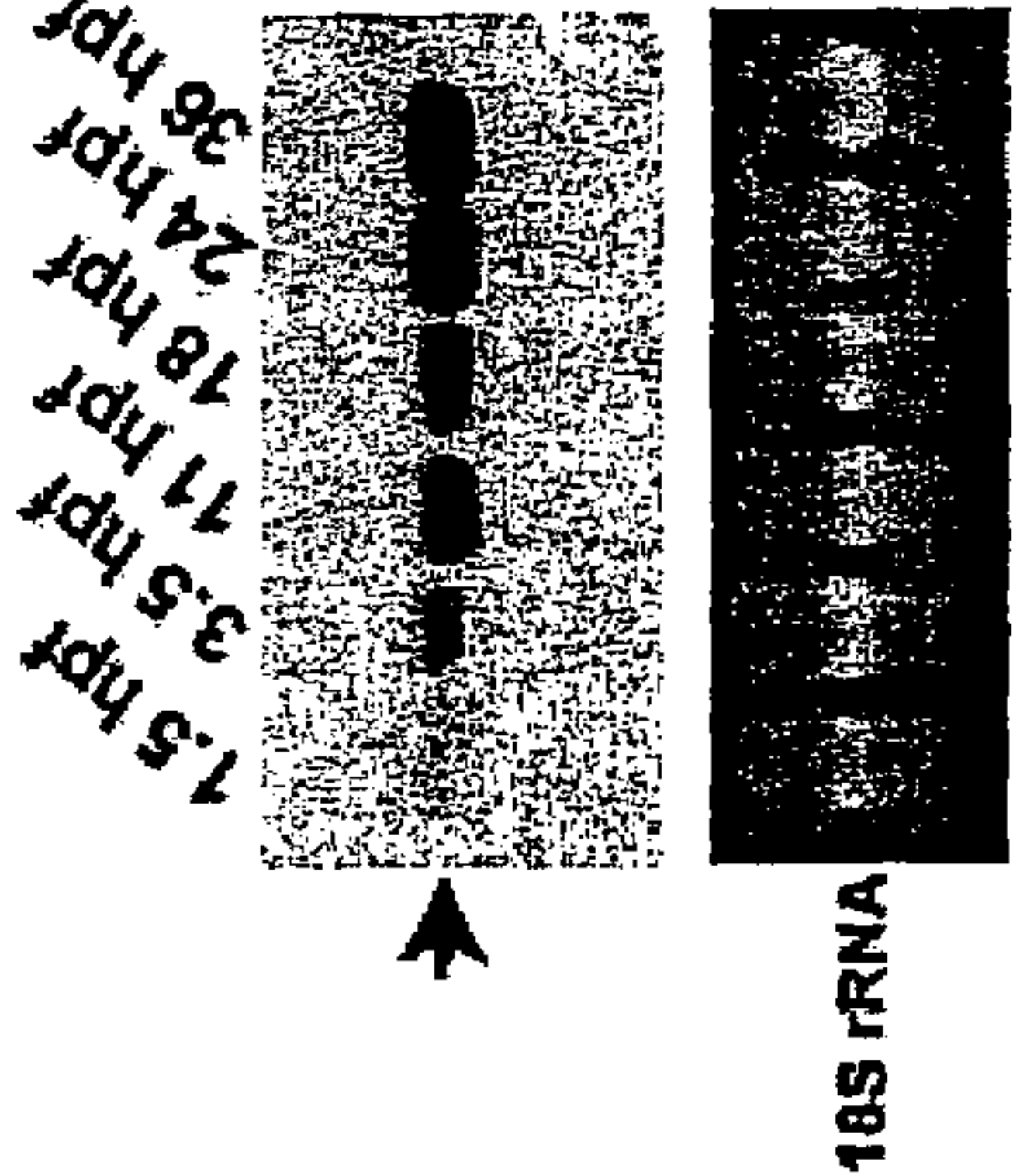


FIGURE 16B

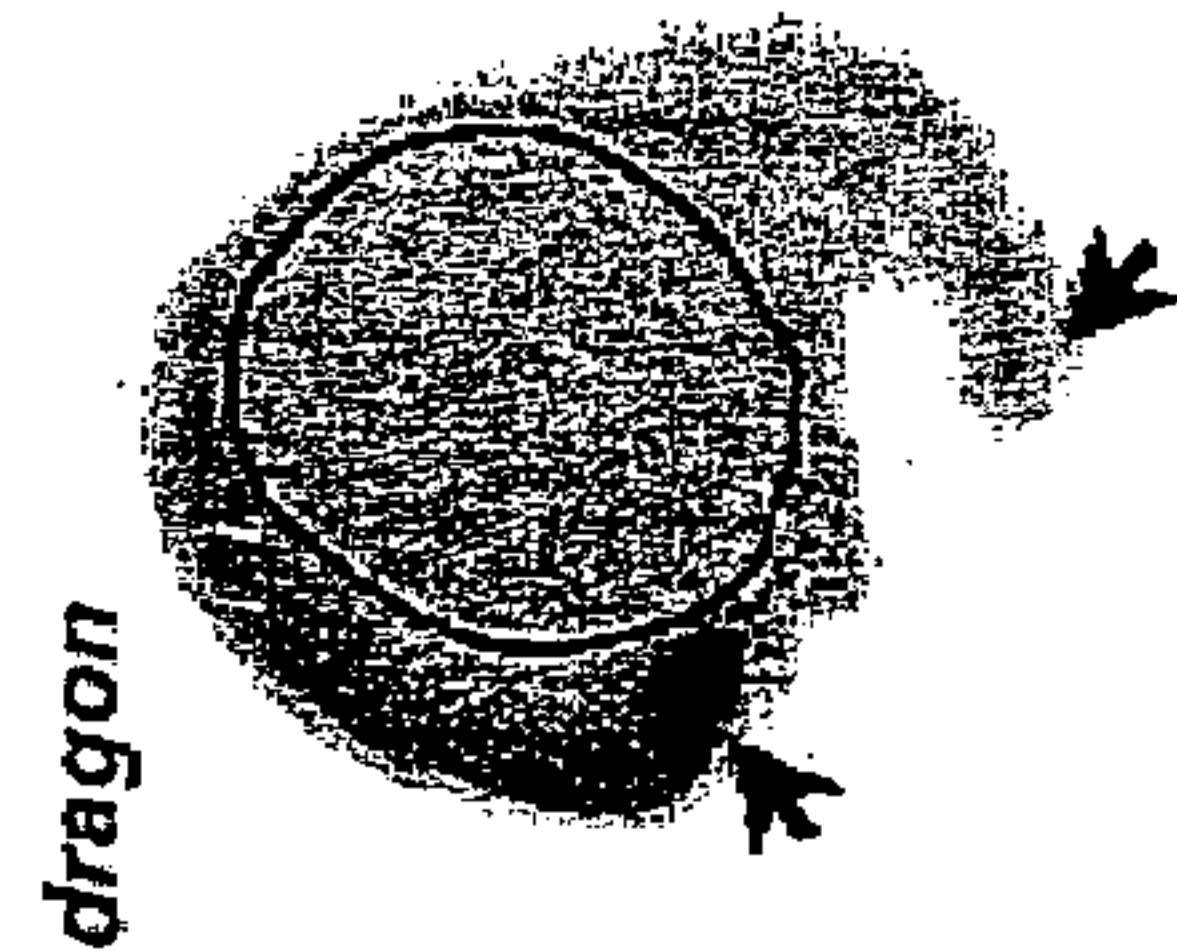


FIGURE 16C

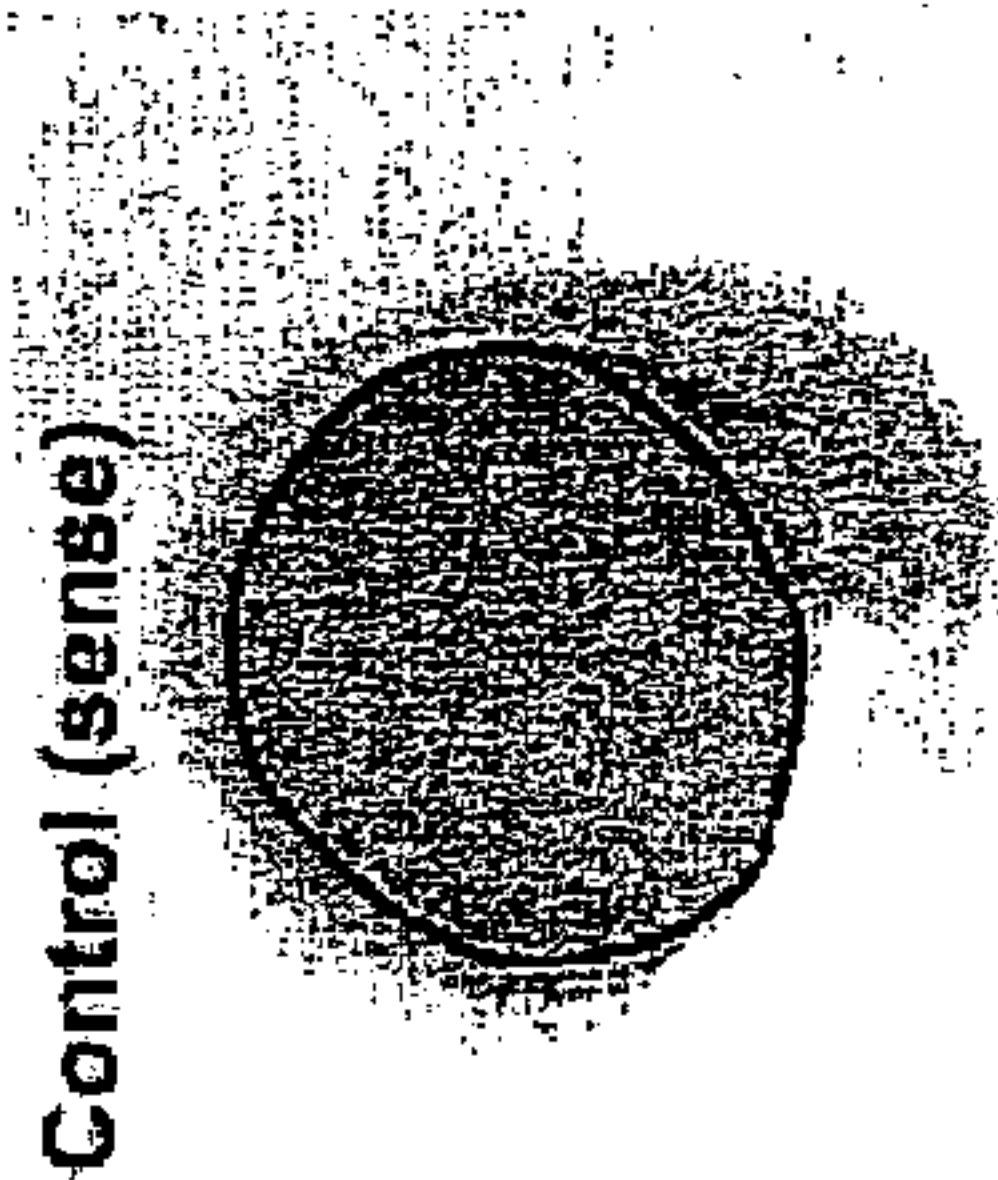
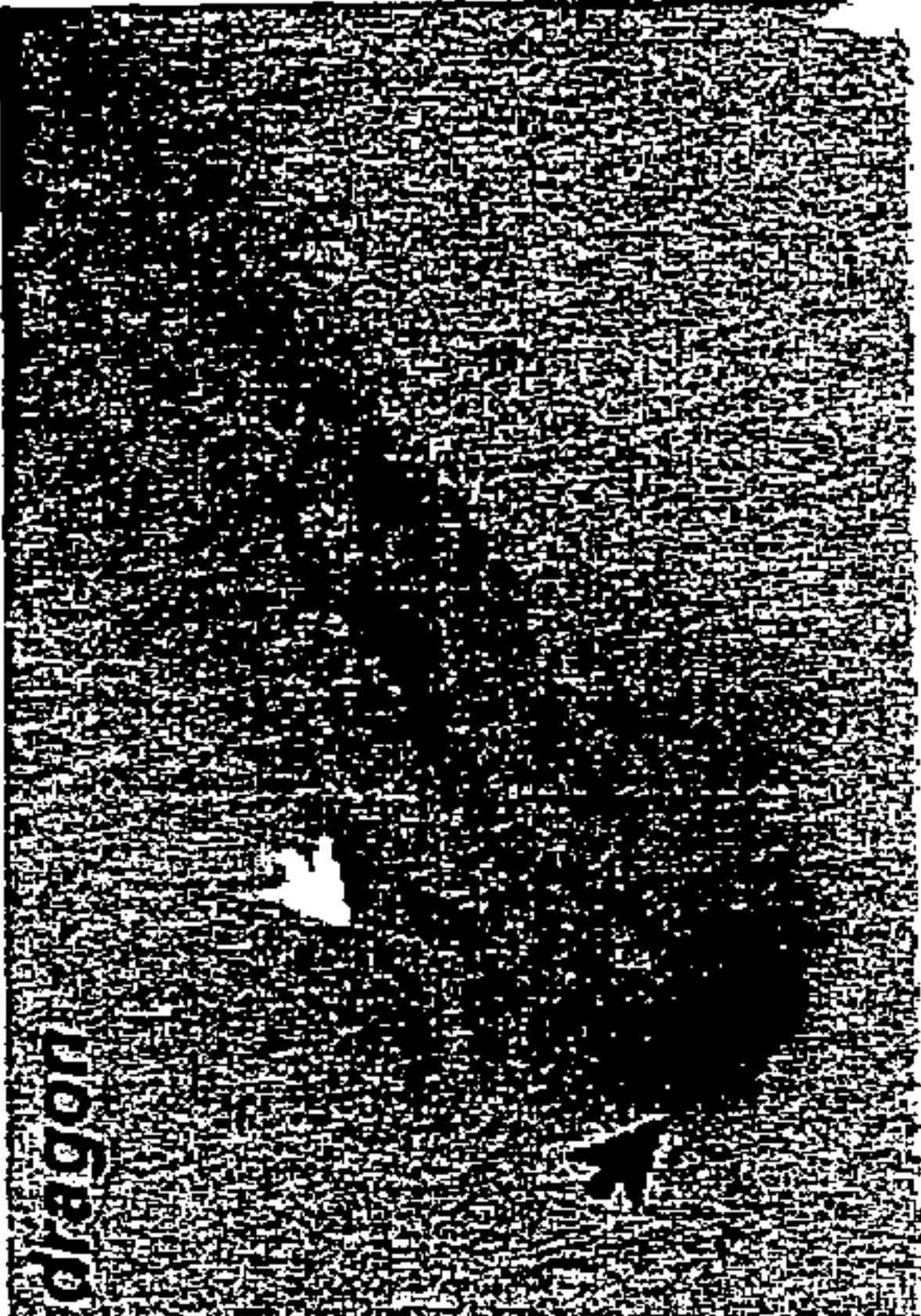


FIGURE 16D



dragon over-expression (25 hpf)

FIGURE 16E

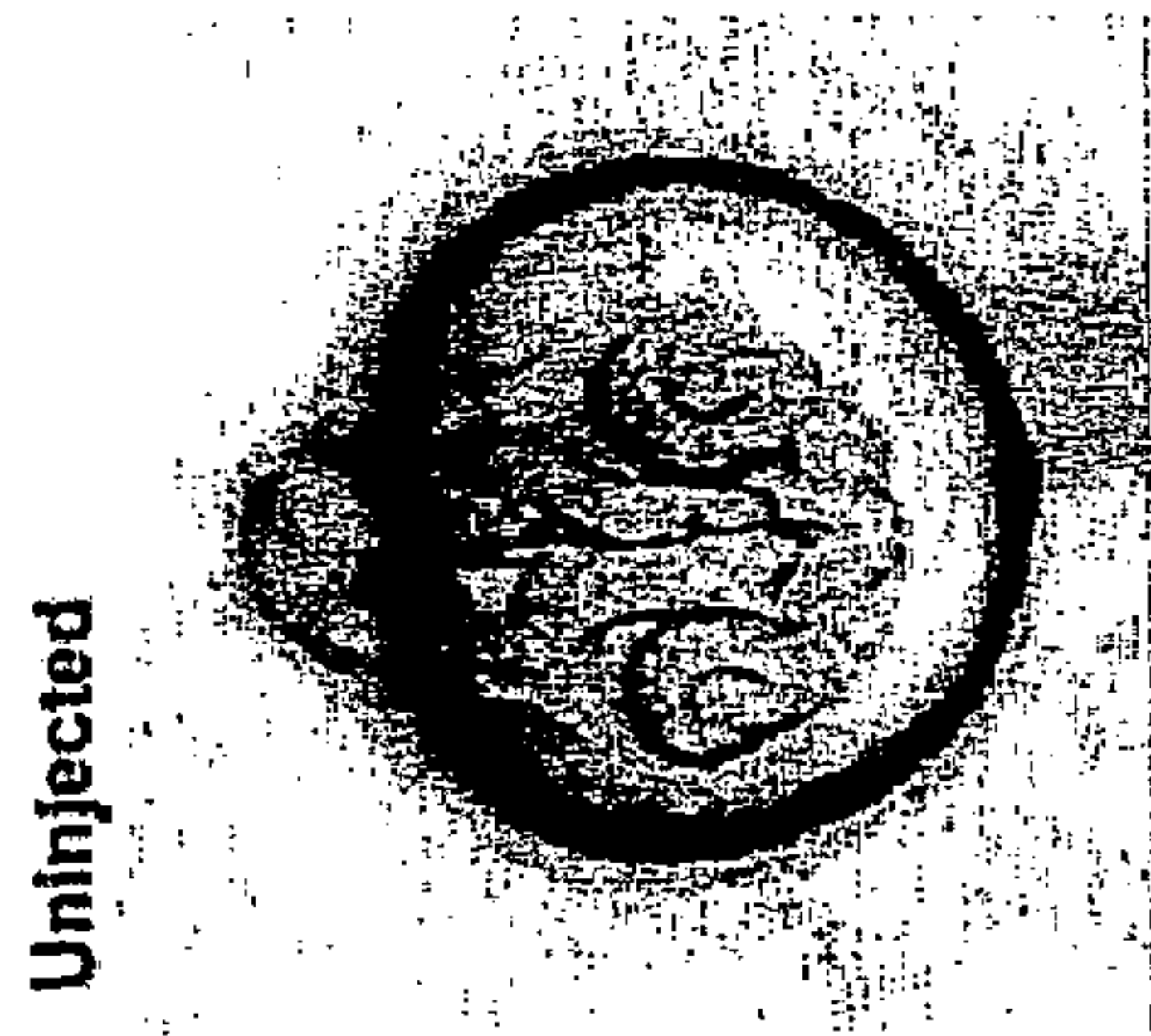


FIGURE 16F

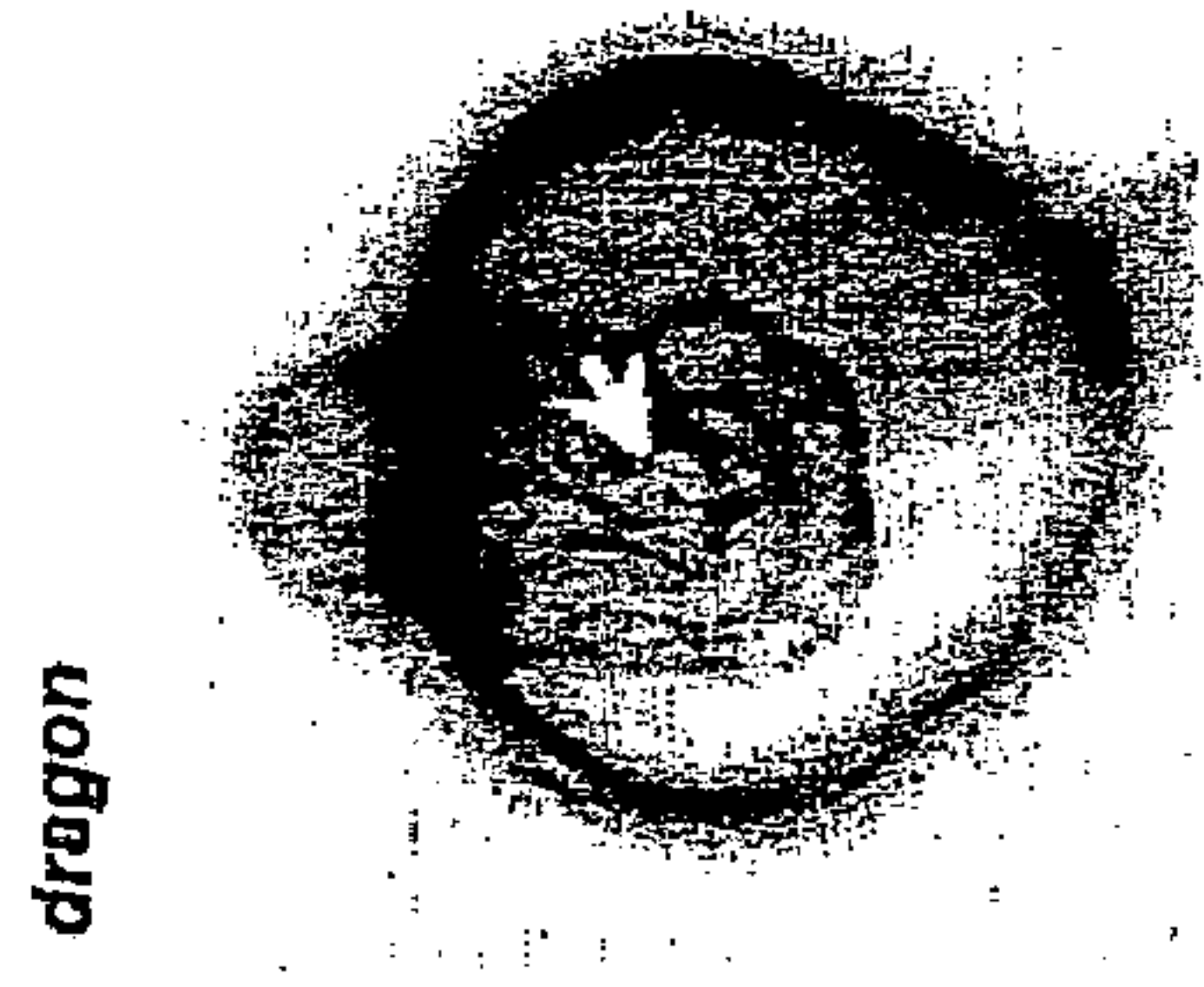
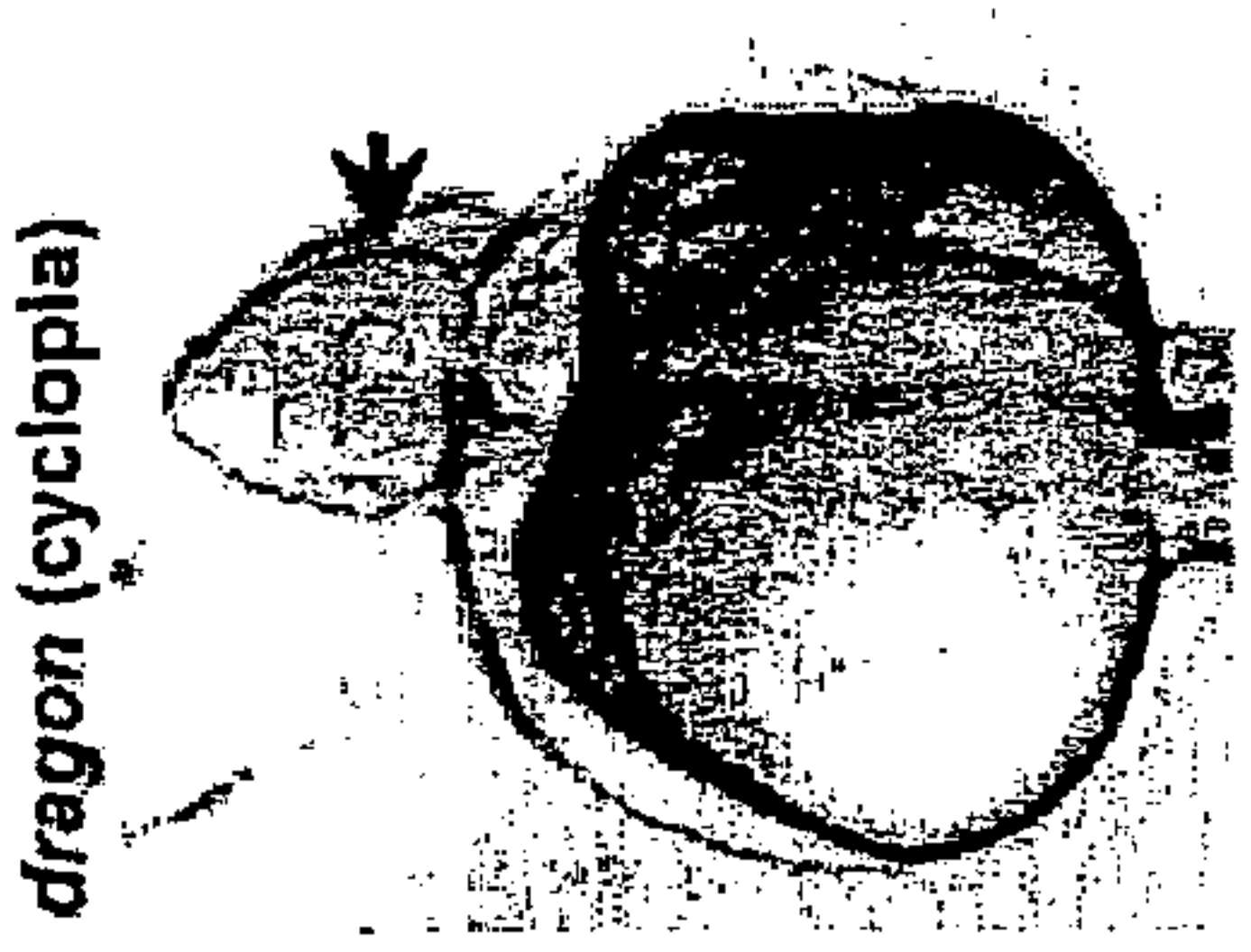


FIGURE 16G



dragon knock-down (Morpholino)

FIGURE 16H

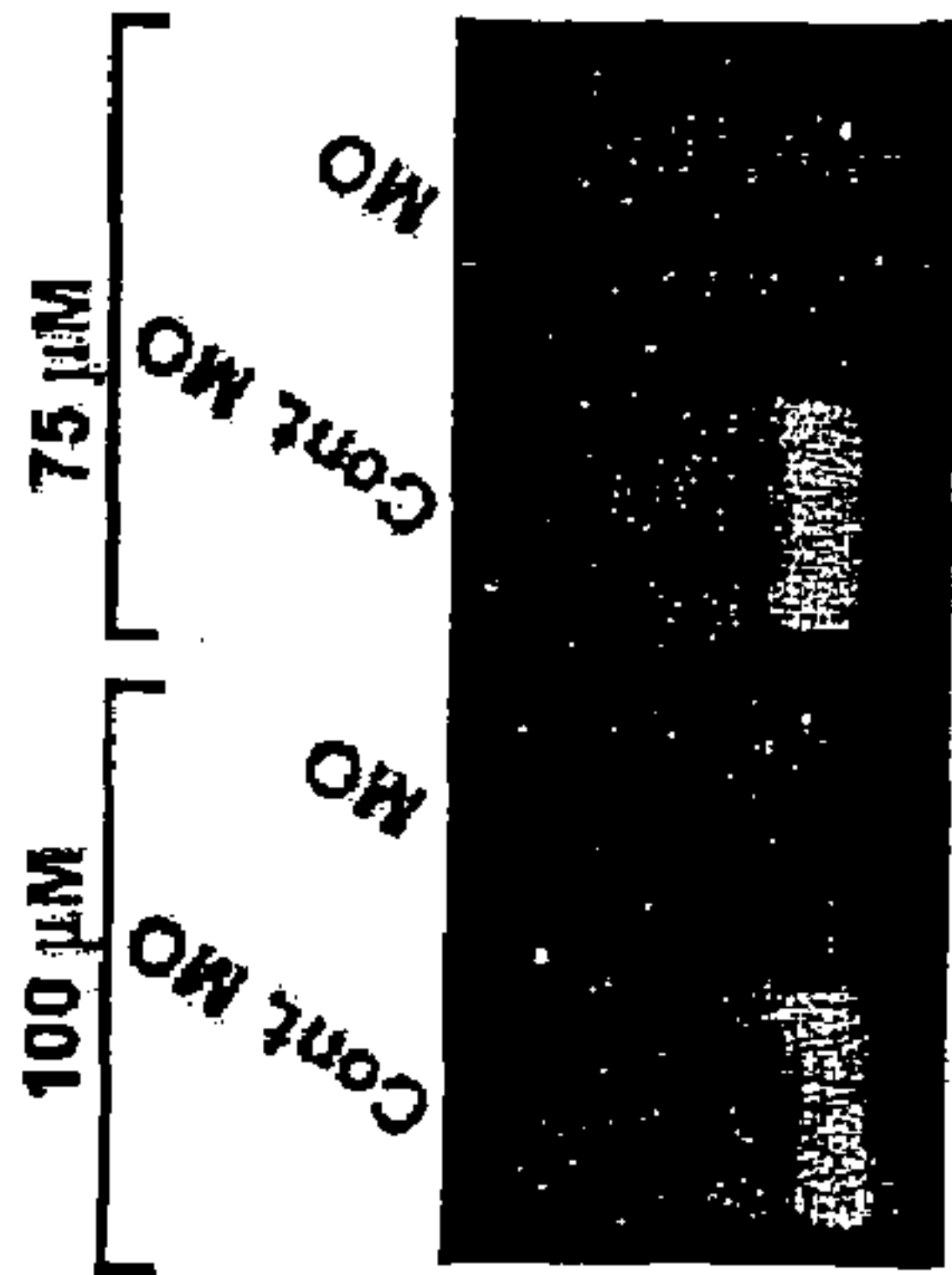


FIGURE 16I

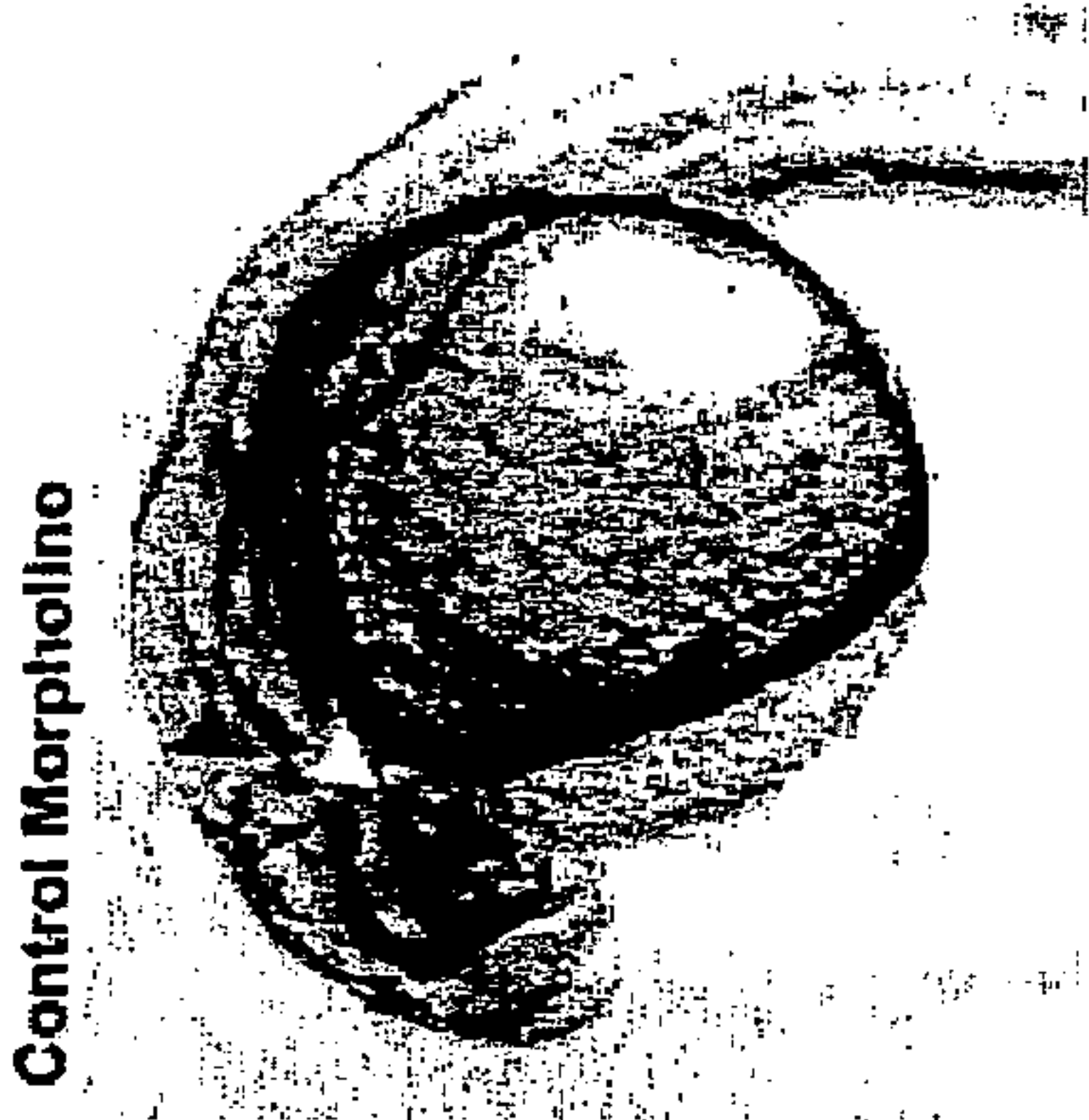


FIGURE 16J

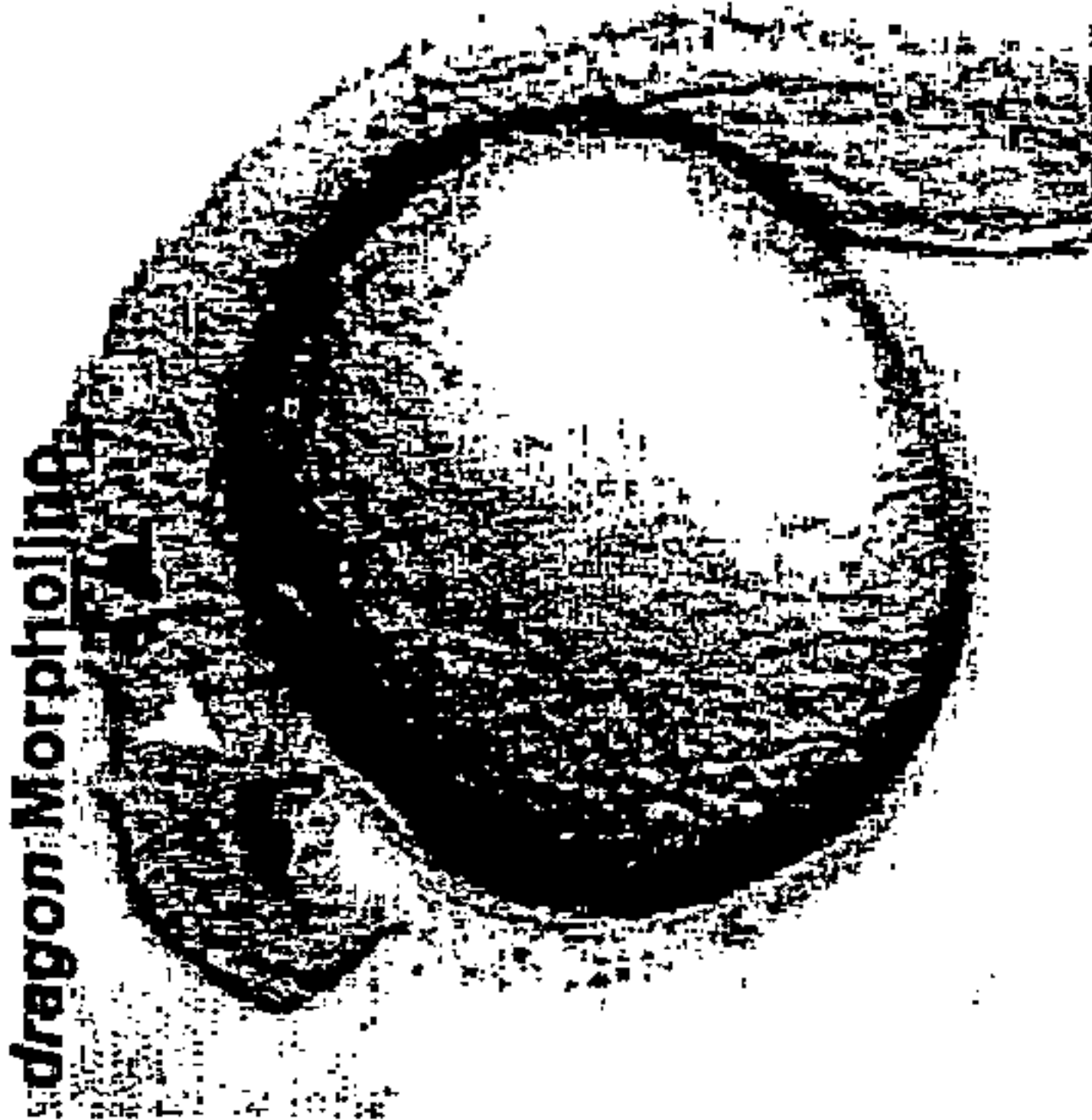


FIGURE 16K

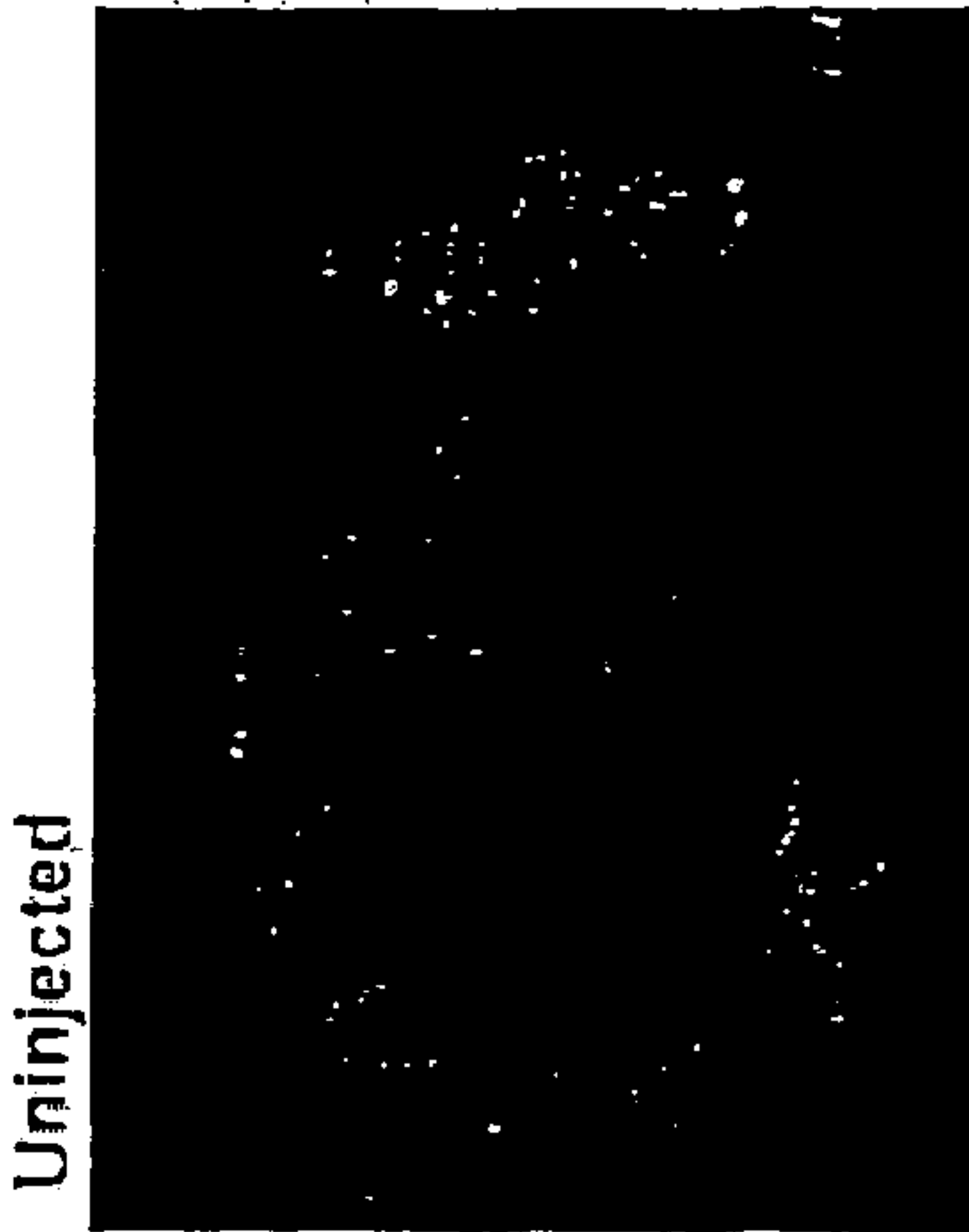


FIGURE 16L

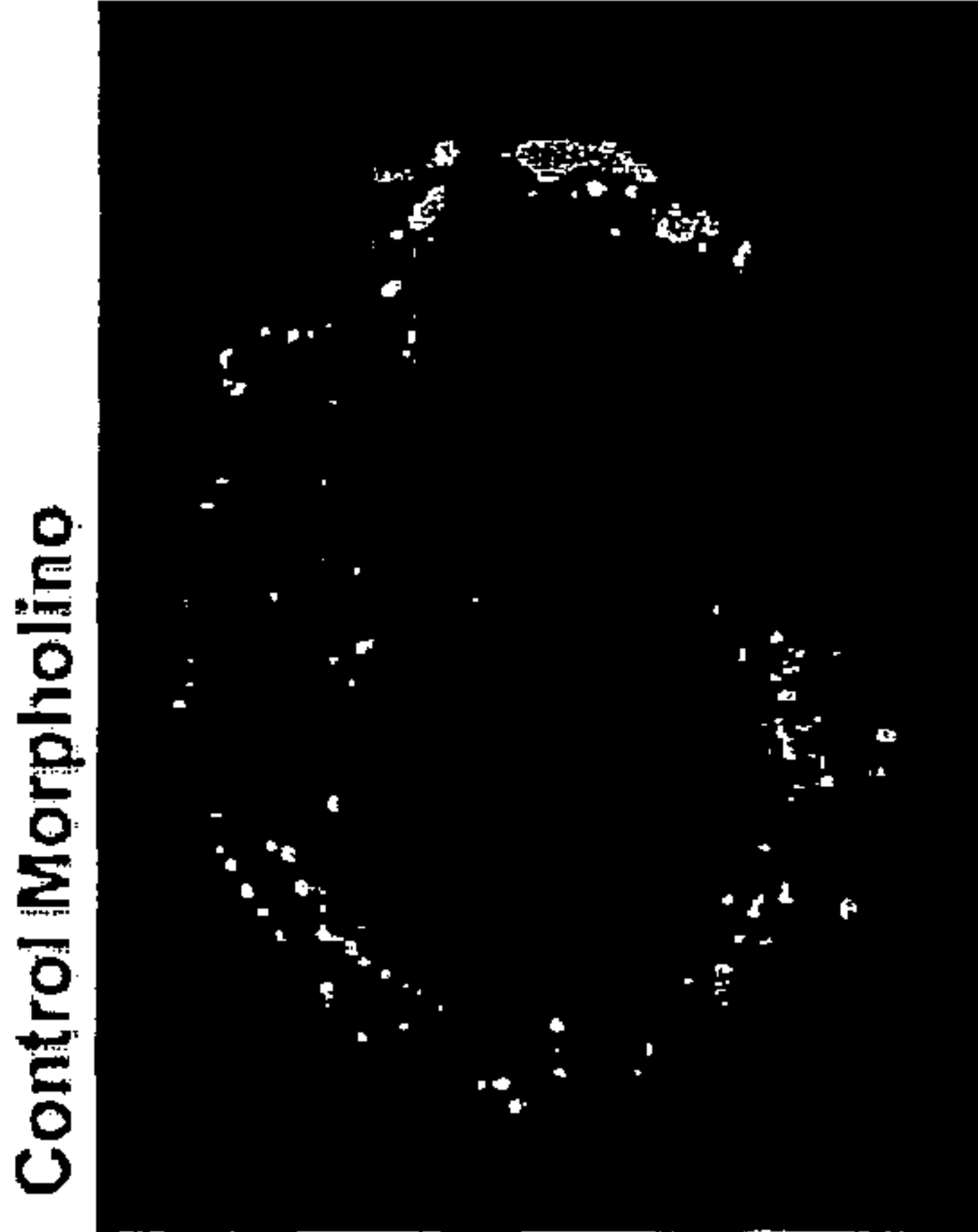


FIGURE 16M



Figure 17

mDRAGON FCGLFGDPHLRTFKDHFQCKVEGAWPLIDNNYLSVQVTNVPVPGSSATATNKVTIIIEK
+C LFGDPHL F QTC EGA PL+DN Y VQVTN V + T KVT++ +
YCSLFGDPHLIMFNGSVQTCSEEGARPLVDNRYFLVQVTNRNVRGEALT TTTVTKVTVLVR

C. elegans AQHECTDQKVYQAVTDD--LPAAFVDGTTSGGDGDKSLHIVEK--ESGRYVEMHARYIG
+H CT Y+A +D+ LP FVDGTT + S H VE + YVE+ +I
-KHNCTASLRYEASSDEEGLPRGFVDGTF----QMTSKHSVEVLWQDDNYVEIALHFIH

TTVFVRQLGRYLTLAIRMPEDLAMS YEESQDL--QLCVNGCPMSECI
+++ +R+ G YL++++R P + E D+ +LC +GC S I
SSIHRRQGPYLSVSVRAP---TIVLETGGDVARELCWSGCRKSSRI

C. elegans DRAGON

MRRHWKEFECEKWESCNDNSHVKRKHVNTGHICGGKFELSEKNLAAKFKYSGDTVWRGRPNFLKSLCYFN
PPPSNRKLIKYSLFGDPHLIMFNGSVQTCSEEGARPLVDNRYFLVQVTNRNVRGEALT TTTVTKVTVLVRK
HNCTASLRYEASSDEEGLPRGFVDGTTFQMTSKHSVEVLWQDDNYVEIALHFIHSSIHRRQGPYLSVSV
RAP TIVLETGGDVARELCWSGCRKSSRI PAELAVEMTKKFAECYRRRVHVPKKVAEETTFLSEQKVLPIY
DRCKDIGNIGVFFDASARKILNFRVSGSQVTS LQNCARRGLRRGQAIILERYFSAPKPKFHLCTATGG
QVTAIQSFEARRGLRRGQATTVERCISAPRDP TDLKIFALTDNCEETKKYWNFFRYDILCDTHSQNFLLP

Figure 18

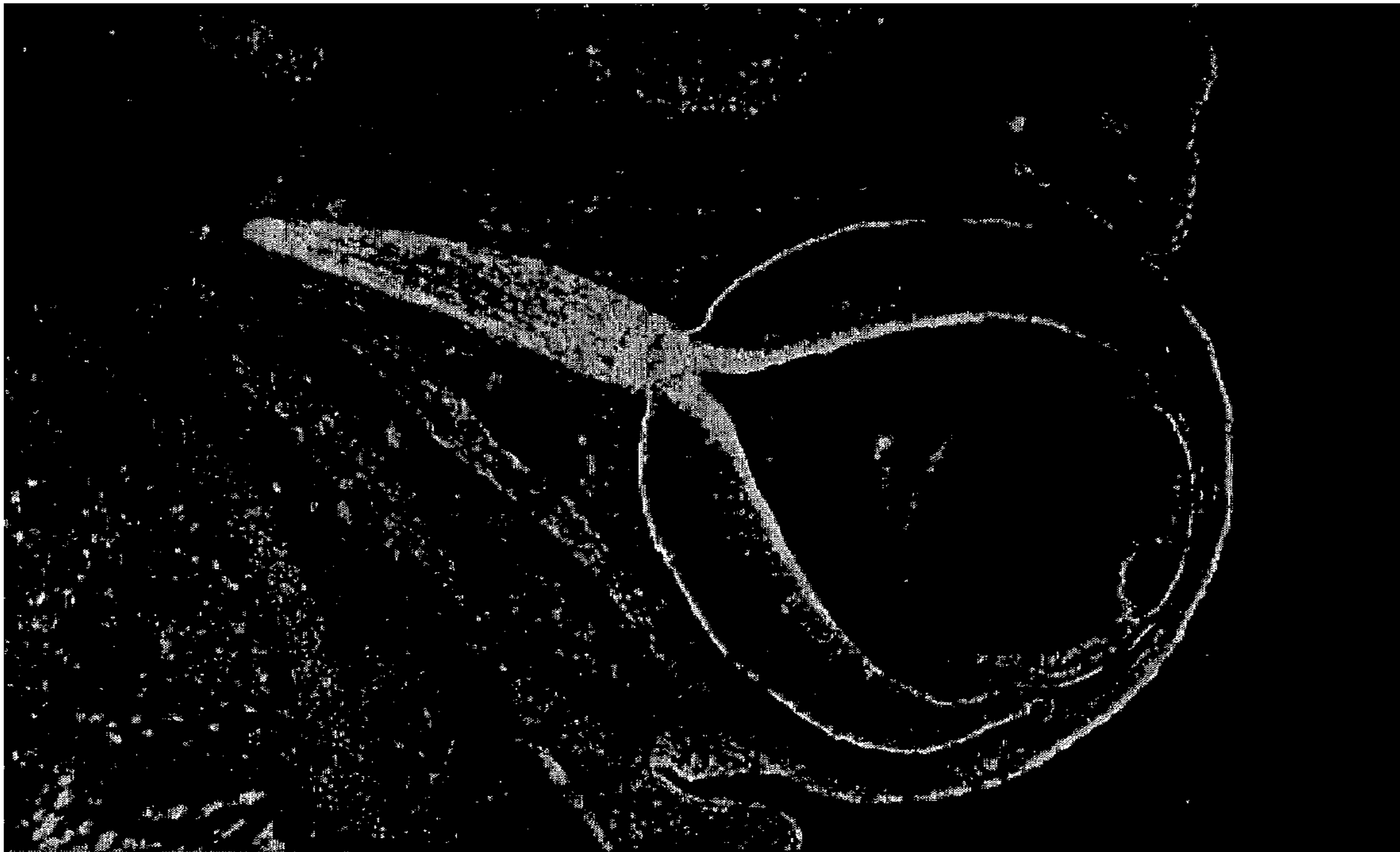


Figure 19



1

**HUMAN DRG11-RESPONSIVE AXONAL
GUIDANCE AND OUTGROWTH OF
NEURITE (DRAGON) PROTEINS AND
VARIANTS THEREOF**

This application claims benefit of U.S. Provisional Application No. 60/373,519, filed Apr. 18, 2002, hereby incorporated by reference.

**FEDERALLY SPONSORED RESEARCH OR
DEVELOPMENT**

This invention was made with Government support under Grant No. 5R01-NS038253 awarded by the National Institutes of Health. The Government has certain rights in this invention.

FIELD OF THE INVENTION

This invention relates to a DRG11-responsive gene and its homologs useful for treating and diagnosing diseases, developmental defects, and injuries of the nervous system, retina, skin, muscle, bone, and joint tissue.

BACKGROUND OF THE INVENTION

Developmentally regulated transcription factors drive developmental gene programs that result in embryo formation and the birth, proliferation, growth, migration, and differentiation of the cells that eventually make up the different tissues of the body. This involves the expression and repression of many genes including those whose protein products act as regulators of this process as signal molecules. When the signal proteins are secreted, they may act both as paracrine signals between different cells, including on stem cells, and as autocrine signals on the same cells that produce the signal molecule. When the protein is not secreted, but rather inserted into the cell membrane, it may contribute to cell-cell interactions.

In the case of the developing nervous system, multiple secreted and non-secreted signal molecules expressed at different times and in different spatial locations are involved in: (i) determining the induction of the neural plate; (ii) regionalization of the neural tube along dorsoventral and anteroposterior axes; (iii) generation of neurons and glia from multipotent precursors (neuronal determination); (iv) determination of survival or apoptotic cell death; (v) migration of neurons; (vi) differentiation and regional patterning of neurons; (vii) neurite outgrowth and axon guidance; (viii) formation of specific synaptic connections between neurons, and (ix) determining neuronal-glia interactions.

Some of these signal molecules may be re-expressed in the adult after injury, or the failure of such re-expression may relate to the failure of mature neurons to survive, grow, or regenerate after injury. Some of the signal molecules may act in pathological situations to either promote or suppress abnormal growth or function. These signal molecules, acting on specific transmembrane receptors, may serve as neuronal determinants, survival factors, growth factors, guidance cues, or differentiation factors, and many may have potential therapeutic roles as biological agents beyond their specific involvement in development. Such factors can have biological activity both in vivo and for maintaining cultured cells in vitro, or for converting pluripotent stem cells into specific neuronal or non-neuronal subtypes. Similarly, mimicking the action of these signal molecules by activating their

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membrane bound receptors or the intracellular signal transduction pathways coupled to their receptors, may also have therapeutic potential.

SUMMARY OF THE INVENTION

We have discovered a novel gene family, designated "Dragon" (DRG11-Responsive Axonal Guidance and Outgrowth of Neurite), expressed in the nervous system, retina, skin, muscle, bone, and joint tissue. Three homologous proteins have been identified in each of the mouse, zebrafish, and human. A partial sequence of an ortholog has also been identified in *C. elegans*.

The invention features substantially pure DRAGON, Dragon-like 1 (DL-1), and Dragon-like 2 (DL-2) proteins, fragments, homologs, and orthologs, as well as non-naturally occurring but substantially identical proteins. Preferably, the proteins are mammalian and/or are substantially identical to murine DRAGON, Dragon-like 1 (mDL-1), or Dragon-like 2 (mDL-2) (SEQ ID NO: 5-7, respectively), or human DRAGON, Dragon-like 1 (hDL-1), or Dragon-like 2 (hDL-2) (SEQ ID NO: 8-10, respectively). Also included in this invention are the zebrafish homologs of DRAGON, DL-1, and DL2 (SEQ ID NO: 28-30, respectively) and the *C. elegans* homolog containing the polypeptide sequence of SEQ ID NO: 18.

Also featured are substantially pure nucleic acids which encode DRAGON and the Dragon-like proteins, for example, from mammals. Preferably, the nucleic acids are substantially identical to the murine DRAGON, DL-1, or DL2 (SEQ ID NO: 1-3, respectively), human DRAGON, DL-1, or DL-2 (SEQ ID NO: 4, 31, and 32, respectively), or zebrafish DRAGON, DL-1, or DL-2 (SEQ ID NO: 25-27, respectively). Other embodiments include nucleic acids which, but for the degeneracy of the genetic code, would be substantially identical to the identified murine, human, and zebrafish Dragon family members, as well as nucleic acids which hybridize under high stringency or, less preferably, low stringency conditions to any of those nucleic acids.

Monoclonal and polyclonal antibodies that selectively bind the Dragon family proteins, for example, of SEQ ID NO: 5-10, 18, or 28-30, can also be prepared and are included in the invention. Preferably, the antibodies specific for murine DRAGON bind a protein sequence encoded by residues 38-56, 261-278, or 369-386 of SEQ ID NO: 5, and the antibodies specific for hDRAGON bind a protein sequence encoded by residues 54-72, 277-294, or 385-408 of SEQ ID NO: 8. Other immunogenic portions of the proteins of the Dragon family also can be used for antibody production.

Also provided are expression vectors containing a coding sequence operably linked to an expression control element, such as a promoter or enhancer element. Preferred coding sequences include those that encode any Dragon family protein or fragment thereof, or express an antisense nucleic acid which is complementary to and capable of hybridizing to a nucleic acid that encodes a member of the Dragon family, or its promoter. Preferably, these antisense nucleic acids include at least 12 and more preferably at least 25 contiguous nucleotides. These vectors can be used to transfect cells, resulting in the production of Dragon family proteins and/or sense or antisense nucleic acids. Suitable cells include, for example, bacteria, yeast, and mammalian, for example, human cells. Transfection may result in stable or transient Dragon expression.

Transgenic non-human organisms with altered Dragon expression levels are also included in the invention. A

transgenic organism of this invention can either have a homologous Dragon-coding sequence (for example, a human DRAGON-coding sequence) inserted into its genome such that Dragon expression is increased, or the endogenous Dragon gene(s) can be disrupted rendering the organism Dragon-deficient. Any non-human organism can be used for transgenic Dragon expression or can be rendered Dragon-deficient. Preferably, the transgenic or Dragon-deficient organisms are *C. elegans* or mammals, such as mice.

The invention also provides a method for treating a patient with a neurological disorder, a developmental deficit, or a congenital disorder of the nervous system by administering a therapeutically effective amount of a Dragon family protein. Preferably, the Dragon protein is a mammalian protein, for example, hDRAGON or hDL-2. Neurological disorders that can be treated according to the methods of this invention include neurodegenerative diseases such as Parkinson's disease, Huntington's disease, Alzheimer's disease, motor neuron diseases and other spinal muscular atrophies, and neuropathies including diabetic neuropathy and inherited demyelinating neuropathies. Other nervous system injuries or functional disorders that can be treated, for example, by DRAGON or DL-2, include those caused by trauma, (e.g., peripheral nerve, dorsal root, spinal cord, and brain injury) cerebrovascular disease (e.g., ischemia, thrombosis, or hemorrhage), chemical-induced neurotoxicity, metabolic diseases, infection, primary and secondary neoplasms of the nervous system, congenital abnormalities of nervous system (e.g., neurofibromatosis, phakomatosis, cerebral palsy, mental retardation), sensory and motor abnormalities, (e.g., pain, nociceptive inflammatory, peripheral and central neuropathic pain), cognitive and mood disorders, psychoses, epilepsy, coordination disorders. Nervous system disorders caused by non-neuronal cells are also amenable to treatment using mammalian DRAGON or DL-2. Disorders of this type include, for example, demyelinating disorders, axonal conduction deficits, and abnormal growth of glial cells (e.g., gliomas, Schwannomas, and neurofibromatosis), degenerative diseases of the retina, cochlea and olfactory mucosa. Treatment may be by any method and is preferably by oral, parenteral, intrathecal, or intracerebrovascular administration.

DRAGON may also be used to treat disorders of the skin. Preferably, the DRAGON protein is mammalian, most preferably human. Administration of a DRAGON protein will preferably be topical in a cream, gel, ointment, or irrigation solution. Alternatively, the DRAGON protein can be administered by subcutaneous injection at or near the lesion site. Disorders amenable to treatment include trauma (i.e., accidental or surgical), burns (i.e., chemical, thermal, or radiation), allergic reactions such as eczema, psoriasis, or contact dermatitis, pressure ulcers, and acne.

The DRAGON protein may also be used to treat disorders of the retina and optic nerve. Administration of a DRAGON protein, preferably a mammalian (i.e., human) protein, may be in the form of eye drops or an irrigation solution. Alternatively, intraocular or intraorbital injection may be used. Retinal disorders amenable to treatment using a DRAGON protein include, for example, traumatic injuries (i.e., detached retina), macular degeneration, and sarcoidosis. Optic nerve diseases amenable to treatment with a DRAGON protein include, for example, ischemic optic neuropathy, primary glaucomatous optic nerve disease (GOND), toxic optic nerve disease, and Leber's Hereditary Optic Neuropathy (LHON).

Dragon-like 1 (DL-1) can be used for treatment of disease conditions of the bone, muscle, joint, or cartilage including

muscle wasting, congenital myopathies, muscular dystrophy, and the innervation of muscle by motor axons (e.g., following peripheral nerve injury), bone fracture, metabolic disorders of bone, disorders of bone formation or resorption, neoplasms of bone, congenital abnormalities of bone (bone dysplasias, achondroplasia, or endochondromatosis), inflammatory or degenerative joint diseases (e.g. arthritis, osteoarthritis, and rheumatoid arthritis), muscle paralysis and other diseases resulting in the failure of the neuromuscular system (e.g., myasthenia gravis).

DL-2 can also be used for treatment of cardiovascular diseases and disorders including, for example, developmental heart abnormalities, congenital cardiac malformations, and blood vessel malformations (e.g., aneurisms).

Embryonic or adult pluripotent cells can be induced to differentiate into neuronal, retinal, epidermal (DRAGON or DL-2), or myogenic (DL-1) phenotypes by contacting the cells with a Dragon protein in a manner sufficient to increase the Dragon biological activity. The Dragon protein that contacts the cells to induce a particular phenotype may result from the overexpressing a Dragon nucleic acid by the cells or the cells can be cultured in the presence of an exogenously applied Dragon protein. Preferably, the cells are human embryonic stem cells or bone marrow-derived stem cells. Optionally, a TGF- β family member or TGF- β receptor can be inhibited to aid in inducing a neuronal phenotype. Cells that have been induced or regulated by DRAGON or DL-2 treatment may be used for subsequent administration to patients to replace lost or abnormally functioning cells.

The invention also provides a method for diagnosing a Dragon-related condition in a patient. Typically, the condition is diagnosed by assessing a Dragon family nucleic acid (e.g., gene) for one or more mutations that reduce Dragon biological activity. The Dragon family nucleic acid may be, for example, DRAGON, DL-1, or DL-2. These mutations may be in an untranslated region such that gene expression is impaired. Alternatively, the mutation may be in a coding region, causing a reduction in protein function. Common techniques for assessing Dragon nucleic acids include, for example, Northern and Southern analysis, including the polymerase chain reaction (PCR), and restriction fragment length polymorphism (RFLP) analysis. Any appropriate patient sample can be used in the diagnostic screening provided; however, particularly useful sample sources include, for example, blood samples and tissue biopsies.

The Dragon family proteins and nucleic acids can also be used to identify candidate compounds which modulate (increase or decrease) Dragon expression, or mimic or inhibit Dragon biological activity. Compounds identified using these screening techniques are useful for treating Dragon-related diseases and conditions described herein. A method for identifying candidate compounds that modulate Dragon activity includes the steps of: (a) exposing a sample to a test compound, wherein the sample contains a Dragon nucleic acid, a Dragon promoter operably linked to a reporter gene (for example, a detectable label such as alkaline phosphatase), or a Dragon protein; and (b) identifying a useful candidate compound by assaying for a change in the level of Dragon expression or biological activity in the sample, relative to a sample not exposed to the test compound.

The invention also features a method for identifying endogenous and synthetic Dragon family binding partners such as Dragon receptors and Dragon ligands. The method includes the steps of: (i) providing a Dragon fusion protein which consists of a Dragon protein linked to a tag molecule; (ii) contacting a sample containing a putative Dragon binding partner with a Dragon fusion protein under conditions

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which allow for a Dragon-Dragon binding partner complex to form, (iii) detecting the Dragon fusion protein by detecting the tag molecule, and (iv) interpreting the Dragon fusion protein detection to determine whether the Dragon fusion protein is complexed to a Dragon binding partner from the sample. In one embodiment, a further step of (v) isolating the Dragon-Dragon binding partner complex using a method directed against the tag molecule. In one preferred embodiment, the step (iii) detecting is done using a detectably labeled antibody. Preferred techniques in step (iii) include, for example, Western blotting and ELISA assays. In another preferred embodiment, the step (v) isolating is done using affinity chromatography.

By "Dragon protein" or "Dragon-family protein" is meant any polypeptide that is substantially identical to the human, murine, or zebrafish DRAGON, Dragon-like 1 (DL-1), or Dragon-like 2 (DL-2) proteins. Dragon proteins also include substantially identical fragments of DRAGON, DL-1, DL-2, or any other Dragon-family protein. Dragon fragments are typically at least 50, 100, 150, 200, 250, 300, 350, or 400 amino acids in length.

By "Dragon nucleic acid" or "Dragon-family nucleic acid" is meant any polynucleotide that is substantially identical to the human or murine DRAGON, DL-1, or DL-2 cDNA sequences, any polynucleotide having a degenerate sequence that encodes a DRAGON, DL-1, or DL-2 protein, or any polynucleotide whose complement hybridizes to a human or murine DRAGON, DL-1, or DL-2 sequence under high stringency conditions. Alternatively, a Dragon nucleic acid encodes a protein which is substantially identical to the human or murine DRAGON, DL-1, or DL-2 proteins.

By "DRAGON protein" is meant a polypeptide having a sequence substantially identical to SEQ ID NO: 5, 8, 18, or 28.

By "DL-1 protein" is meant a polypeptide having a sequence substantially identical to either SEQ ID NO: 6, 9, or 29.

By "DL-2 protein" is meant a polypeptide having a sequence substantially identical to either SEQ ID NO: 7, 10, or 30.

By "DRAGON nucleic acid" is meant a polynucleotide having a sequence which encodes a DRAGON protein. Preferably, a DRAGON nucleic acid is substantially identical or hybridizes under high stringency conditions to SEQ ID NO: 1, 4, or 25.

By "DL-1 nucleic acid" is meant a polynucleotide having a sequence which encodes a DL-1 protein. Preferably, a DL-1 nucleic acid is substantially identical or hybridizes under high stringency conditions to SEQ ID NO: 2, 26, or 31.

By "DL-2 nucleic acid" is meant a polynucleotide having a sequence which encodes a DL-2 protein. Preferably, a DL-2 nucleic acid is substantially identical or hybridizes under high stringency conditions to SEQ ID NO: 3, 27, or 32.

By "substantially identical" is meant a polypeptide or nucleic acid exhibiting at least 50%, 75%, 85%, 90%, 95%, or even 99% identity to a reference amino acid or nucleic acid sequence. For polypeptides, the length of comparison sequences will generally be at least 20 amino acids, preferably at least 30 amino acids, more preferably at least 40 amino acids, and most preferably 50 amino acids, or full-length. For nucleic acids, the length of comparison sequences will generally be at least 60 nucleotides, preferably at least 90 nucleotides, and more preferably at least 120 nucleotides, or full length.

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By "high stringency conditions" is meant any set of conditions that are characterized by high temperature and low ionic strength and allow hybridization comparable with those resulting from the use of a DNA probe of at least 40 nucleotides in length, in a buffer containing 0.5 M NaHPO₄, pH 7.2, 7% SDS, 1mM EDTA, and 1% BSA (Fraction V), at a temperature of 65° C., or a buffer containing 48% formamide, 4.8×SSC, 0.2 M Tris-C1, pH 7.6, 1×Denhardt's solution, 10% dextran sulfate, and 0.1% SDS, at a temperature of 42° C. Other conditions for high stringency hybridization, such as for PCR, Northern, Southern, or in situ hybridization, DNA sequencing, etc., are well-known by those skilled in the art of molecular biology. See, e.g., Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, N.Y., 2000, hereby incorporated by reference.

By "Dragon antisense nucleic acid" is meant a nucleic acid complementary to a Dragon coding, regulatory, or promoter sequence, including human and murine DRAGON, Dragon-like 1 (DL-1) and Dragon-like 2 (DL-2). Preferably, the antisense nucleic acid decreases expression (e.g., transcription and/or translation) of the Dragon by at least 5%, 10%, 25%, 50%, 75%, 90%, 95%, or even 99%. Preferably, the Dragon antisense nucleic acid comprises from about 8 to 30 nucleotides. A Dragon antisense nucleic acid may also contain at least 40, 60, 85, 120, or more consecutive nucleotides that are complementary to a Dragon mRNA or DNA, and may be as long as a full-length Dragon gene or mRNA. The antisense nucleic acid may contain a modified backbone, for example, phosphorothioate, phosphorodithioate, or other modified backbones known in the art, or may contain non-natural internucleoside linkages.

A Dragon antisense nucleic acid may also be encoded by a vector where the vector is capable of directing expression of the antisense nucleic acid. This vector may be inserted into a cell using methods known to those skilled in the art. For example, a full length Dragon nucleic acid sequence, or portions thereof, can be cloned into a retroviral vector and driven from its endogenous promoter or from the retroviral long terminal repeat or from a promoter specific for the target cell type of interest. Other viral vectors which can be used include adenovirus, adeno-associated virus, vaccinia virus, bovine papilloma virus, or a herpes virus, such as Epstein-Barr Virus.

By "vector" is meant a DNA molecule, usually derived from a plasmid or bacteriophage, into which fragments of DNA may be inserted or cloned. A vector will contain one or more unique restriction sites, and may be capable of autonomous replication in a defined host or vehicle organism such that the cloned sequence is reproducible. A vector contains a promoter operably linked to a gene or coding region such that, upon transfection into a recipient cell, an RNA is expressed.

By "substantially pure" is meant a nucleic acid, polypeptide, or other molecule that has been separated from the components that naturally accompany it. Typically, the polypeptide is substantially pure when it is at least 60%, 70%, 80%, 90%, 95%, or even 99%, by weight, free from the proteins and naturally-occurring organic molecules with which it is naturally associated. For example, a substantially pure polypeptide may be obtained by extraction from a natural source, by expression of a recombinant nucleic acid in a cell that does not normally express that protein, or by chemical synthesis.

By a "promoter" is meant a nucleic acid sequence sufficient to direct transcription of a gene. Also included in the invention are those promoter elements which are sufficient

to render promoter-dependent gene expression controllable for cell type-specific, tissue-specific or inducible by external signals or agents (e.g. enhancers or repressors); such elements may be located in the 5' or 3' regions of the native gene, or within an intron.

By "operably linked" is meant that a nucleic acid molecule and one or more regulatory sequences (e.g., a promoter) are connected in such a way as to permit expression and/or secretion of the product (e.g., a protein) of the nucleic acid molecule when the appropriate molecules (e.g., transcriptional activator proteins) are bound to the regulatory sequences.

By "signal sequence" is meant a nucleic acid sequence which, when operably linked to a nucleic acid molecule, facilitates secretion of the product of the nucleic acid molecule. The signal sequence is preferably located 5' to the nucleic acid molecule.

By "transgene" is meant any piece of nucleic acid that is inserted by artifice into a cell, or an ancestor thereof, and becomes part of the genome of the animal which develops from that cell. Such a transgene may include a gene which is partly or entirely heterologous to the transgenic animal, or may represent a gene homologous to an endogenous gene of the animal.

By "transgenic" is meant any cell which includes a nucleic acid sequence that has been inserted by artifice into a cell, or an ancestor thereof, and becomes part of the genome of the organism which develops from that cell. Preferably, the transgenic organisms are transgenic mammals (e.g., rodents or ruminants), or *C. elegans*, Zebra fish, or *Drosophila*. Preferably the nucleic acid (transgene) is inserted by artifice into the nuclear genome.

By "antibody that selectively binds" is meant an antibody capable of a high affinity interaction with a specific target molecule, having a dissociation constant of <1 μ M, <100 nM, <10 nM, <1 nM, or even <100 pM. Preferably, the antibody has at least 10-fold, 100-fold, 1,000-fold, or even 10,000-fold lower affinity for other, non-target molecules.

By a "neurological disorder" is meant any disease or condition that causes injury to any component of the peripheral or central nervous system, including the retina. Neurological disorders include acute and chronic conditions. Acute conditions include, for example, trauma, stroke, and chemical-induced neurotoxicity. Chronic conditions include, for example, neurodegenerative diseases and cancers of the nervous system including gliomas, schwannomas, and astrocytomas. Neurological disorders can also arise from developmental defects, including inherited or congenital defects (e.g. cerebral palsy), and autoimmune diseases (e.g., multiple sclerosis). Neurological disorders also include functional disorders such as paralysis, and epilepsy, as well as sensory, mood, and psychomotor disorders (e.g., fibromyalgia, dyesthesia).

By a "neurodegenerative disease" is meant any disease of the central, peripheral, or autonomic nervous system that is characterized by progressive neuronal loss or dysfunction, including but not limited to Alzheimer's Disease, dementia pugilistica, Parkinson's Disease, Huntington's Disease, Niemann-Pick disease, multiple sclerosis, neuropathies (e.g., central, peripheral, compression type, and diabetic) and ischemic conditions such as stroke and cerebral artery infarction. Defects in myelin repair are also considered neurological diseases. The defects may arise during the process of demyelination, the removal of myelin debris following injury, or the remyelination process.

By a "bone disorder" is meant any condition of the bone which is characterized by altered bone remodeling. Bone

disorders include physical traumas such as bone fractures, metabolic bone diseases such as Paget's disease and hyperostosis, and bone neoplasms (e.g., osteochondromas, osteogenic sarcoma).

By a "joint disorder" is meant any trauma or disease process which causes inflammation in or around the cartilage or joint capsule. Joint disorders include, for example, inflammatory arthritis, rheumatoid arthritis, and osteoarthritis.

By a "muscle disorder" is meant any dysfunction of muscle tissue regardless of cause. Muscle disorder may arise for congenital abnormalities, trauma, metabolic disease, or autoimmune disease. Muscle disorders include, for example, muscular dystrophy, myasthenia gravis, transient and periodic muscle paralysis, muscle wasting diseases, muscular dystrophy, myotonia congenital, myotonic dystrophy, and loss of innervation of motor endplates.

By a "Dragon-related condition" is meant any disease or disorder which is associated with the dysfunction or altered (increased or decreased) activity or expression of any one or more of the Dragon protein family. Alternatively, Dragon-related conditions can also refer to any disease or disorder which, although not associated with Dragon dysfunction, is amenable to treatment by modulating (increasing or decreasing) the activity or expression of any one or more Dragon proteins or nucleic acids or by mimicking their actions. Dragon-related conditions include, for example, neurological, retinal, and skin disorders, neurodegenerative diseases, and muscle, bone, or joint disorders.

By a "therapeutically effective amount" is meant a quantity of compound (e.g., a Dragon family protein) delivered with sufficient frequency to provide a medical benefit to the patient. Thus, a therapeutically effective amount of a Dragon family protein is an amount sufficient to treat or ameliorate a Dragon-related condition or symptoms.

By "treating" is meant administering a pharmaceutical composition for the purpose of improving the condition of a patient by reducing, alleviating, or reversing at least one adverse effect or symptom.

BRIEF DESCRIPTION OF DRAWINGS

FIG. 1A is a schematic diagram illustrating a strategy for genomic screening with a CpG island library. The plasmid DNA was bound with GST-DRG11-DBD and passed through a nitrocellulose filter. DRG11-bound plasmids were eluted and amplified in bacterial cultures. The DRG11-bound plasmids were concentrated by repeating the cycle a total of five times. FIG. 1B is the nucleic acid (SEQ ID NO: 1) and deduced polypeptide sequence of murine DRAGON (mDRAGON; SEQ ID NO: 5). The DRAGON protein contains an N-terminal signal peptide and a C-terminal glycosylphosphatidylinositol (GPI) anchor.

FIG. 2 is a graph illustrating the result of a computational structure-function analysis of mDRAGON (SEQ ID NO: 5), demonstrating the presence of a signal sequence which results in protein secretion.

FIG. 3 is a sequence alignment of hDRAGON (SEQ ID NO: 8) and a portion of the insulin-like growth factor binding protein 2 (IGFBP2; SEQ ID NO: 12).

FIG. 4 is a sequence alignment of hDRAGON (SEQ ID NO: 8) and a portion of the ephrin type-B receptor 3 precursor (EPHB3; SEQ ID NO: 13).

FIG. 5 is a sequence alignment showing domain homology between mDRAGON (SEQ ID NO: 5) and portions of human Notch 3 (SEQ ID NO: 14) and murine phosphatidylinositol-4-kinase type II beta (SEQ ID NO: 15).

FIG. 6 is a sequence alignment showing the domain homology between mDRAGON (SEQ ID NO: 5) and a portion of thrombospondin-1 (SEQ ID NO: 16; THR-1) and Slit2 (SEQ ID NO: 17).

FIG. 7A is a photomicrograph of an in situ hybridization study showing that DRAGON and DRG11 mRNAs are both expressed in the dorsal root ganglion (DRG) and the spinal cord at E12.5. FIG. 7B is a bar graph showing the DRG11-dependent enhancer activity of the DRAGON promoter fragment. FIG. 7C shows the results of a pull-down experiment using either GST or GST-DBD (DBD=DRG11 DNA Binding Domain). The purified proteins (right panel) were incubated with the DRAGON promoter fragment, and "pulled down" using glutathione sepharose. Only GST-DBD fusion protein pulled down the promoter fragment as assessed by agarose gel electrophoresis. FIG. 7D is a photomicrograph of an in situ hybridization study demonstrating a decrease in DRAGON mRNA expression in the DRG and the spinal cord of DRG11^{-/-} mouse at E14.5, compared to wildtype. FIG. 7E shows the result of a Northern blot analysis of DRAGON mRNA expression in adult and embryonic E14.5 tissue. FIG. 7F shows the result of a Northern blot analysis of DRAGON mRNA expression in whole mouse embryos during development. β -actin mRNA levels were used as a loading control.

FIG. 8A is an amino acid sequence alignment of mDRAGON (SEQ ID NO: 5), mDL-2 (SEQ ID NO: 7), and mDL-1 (SEQ ID NO: 6). FIG. 8B is an amino acid sequence alignment of mDRAGON (SEQ ID NO: 5), hDRAGON (SEQ ID NO: 8), and zDRAGON (SEQ ID NO: 26).

FIGS. 9A–9L are a series of photomicrographs showing, by in situ hybridization, the developmental distribution of DRAGON family members in the mouse embryo. FIG. 9A–9C demonstrate that DRAGON and DL-2, but not DL-1, mRNA is expressed in mouse embryonic E14.5 spinal cord. DRAGON is the only family member expressed in the DRG. FIGS. 9D–9F are transverse sections of whole mouse E17.5 embryo demonstrating DRAGON, DL-1, and DL-2 mRNA expression. FIGS. 9G–9I demonstrate DRAGON mRNA expression in transverse sections of mouse E17.5 embryo head. (FIG. 9G: Mes.: mesencephalic vesicle; E.: ependymal layer; M.: mantle layer. FIG. 9H: M.: myelencephalon; D: diencephalon; S: striatum; C: cortex. FIG. 9I: D: DRG; S.C.: spinal cord; C.: cochlea; R.: retina; Olf.: future olfactory lobe) FIGS. 9J–9L demonstrate DL-2 mRNA expression in transverse section of mouse E17.5 embryo head.

FIG. 10 is a series of photomicrographs showing the distribution of DRAGON mRNA in the adult rat DRG by in situ hybridization.

FIG. 11 is a series of photomicrographs showing the distribution of DRAGON mRNA in the brain of an E18 mouse by in situ hybridization.

FIG. 12 is a series of photomicrographs showing the distribution of DL-2 mRNA in the brain of an E18 mouse by in situ hybridization.

FIGS. 13A–13D provide experimental results using a novel anti-DRAGON rabbit polyclonal antibody. FIG. 13A is a Western blot analysis of protein extract from untransfected HEK293 cells (–), or those transfected (+) with DRAGON expression vector. A distinct band having a molecular weight of about 50 KDa is recognized by the anti-DRAGON antibody in transfected, but not control, cells. ERK protein level was used as a loading control. FIG. 13B is a photomicrograph of an immunocytochemical study showing significant staining of DRAGON-expressing HEK cells (top). Pretreatment of DRAGON-expressing HEK cells with PI-PLC causes a significant reduction of anti-

DRAGON staining (bottom). Non-transfected HEK cells show no anti-DRAGON staining (not shown). FIG. 13C is a photomicrograph of a Western blot analysis of samples of DRAGON-expressing HEK cell culture medium, with or without pretreatment using PI-PLC. A band corresponding to DRAGON is detected in PI-PLC treated medium samples. FIG. 13D is a series of photomicrographs from an anti-DRAGON immunohistochemical study of adult spinal cord and DRG at low (top) and high (middle) magnification. As a control, the anti-DRAGON antibody was pretreated with the immunogenic DRAGON fragment prior to immunohistochemical staining (bottom). Scale, 100 μ M.

FIGS. 14A–14C are a series of photomicrographs that demonstrate the adhesion of DRG neurons to DRAGON-expressing HEK 293 cells. P14 neonatal DRG neurons were plated on a monolayer of confluent HEK cells (FIG. 14A) and DRAGON transfected HEK cells (FIG. 14B). The culture slides were washed, fixed, and immunostained for DRG neurons using anti-NeuN (neuronal marker). Double immuno-labeling using anti-NeuN and anti-DRAGON indicates a direct interaction between DRAGON expressing HEK cells and DRG neurons (FIG. 14C). FIG. 14D is a bar graph quantifying the adhesion experiment results. A 1.9-fold increase in the number of adherent DRG neurons when plated on DRAGON-expressing HEK 293 cells, compared to control HEK 293 cells. Pretreatment of the DRAGON-expressing HEK cells with PI-PLC significantly reduced the adherence of DRG neurons.

FIG. 15A is a series of photomicrographs demonstrating the effect of DRAGON overexpression in *Xenopus laevis*. Embryos were injected in the animal pole of 1 out of 2 cells at the 2-cell stage with DRAGON RNA and analyzed at late neurula (st23) for changes in neural crest patterning and early tadpole stages (st28) for ectopic induction of neural tissue. DRAGON overexpression inhibits twist RNA expression. However, DRAGON induces ectopic N-tubulin RNA expression. FIG. 15B is a Northern blot from animal cap explants demonstrating that DRAGON induces anterior neural markers, cement gland markers, and nkx2.5 (a cardiac marker).

FIG. 16A is a Northern blot showing the developmental expression of DRAGON in Zebrafish embryos over the first 36 hours post fertilization (hpf). FIGS. 16B–16C are lateral views of an 18–20 somite stage zebrafish embryo following in situ hybridization using a DRAGON antisense probe (FIG. 16B) or the sense control (FIG. 16C). DRAGON expression is strongest at the anterior pole and in the tail-bud region (arrows). More diffuse and lower levels of expression are seen in other parts of the brain. FIG. 16D is a dorsal view of a flat-mounted embryo showing DRAGON staining in the CNS. DRAGON expression is particularly strong in the region surrounding the olfactory placodes (black arrow) and in the retina (white arrow). FIGS. 16E–16G are photomicrographs demonstrating that DRAGON overexpression causes abnormalities in brain morphology and, at a lower frequency (7–15%), cylopia. FIG. 16H is a Northern blot of zebrafish embryo RNA demonstrating that a morpholino oligonucleotide (MO) targeted against the splice donor site of DRAGON exon1 blocks RNA splicing and protein expression. An inverted morpholino oligonucleotide (cont. MO), which preserves the base composition, was used as the control. Primers flanking the intron used for RT-PCR produce the predicted bands from the end products of splicing in the control but not the experimental morpholinos. FIGS. 16I–16J are photomicrographs of 24 hour zebrafish embryos following MO injection. Morphologically, the eyes are affected and extensive cell death in the brain obscures the

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clear definition of the midbrain-hindbrain boundary. FIGS. 16K–16M show TUNEL staining of MO injected embryos at the 21 somite stage revealing a pattern of cell death correlating with the pattern of DRAGON expression.

FIG. 17 is a sequence alignment of mDRAGON (SEQ ID NO: 5) and a region of *C. elegans* DRAGON. The full length *C. elegans* DRAGON is also provided (SEQ ID NO: 18).

FIG. 18 is a photomicrograph showing the distribution of DRAGON expression in the retina and optic nerve of a mouse embryo (E14.5) using immunohistochemistry.

FIG. 19 is a photomicrograph showing the distribution of DRAGON expression in rat glabrous skin (base of the epidermis of the hindpaw) using immunohistochemistry. DRAGON expression is highest in the Merkel cells.

DETAILED DESCRIPTION

DRG11 is a paired homeodomain transcription factor that is expressed both by dorsal root ganglion (DRG) sensory neurons and by dorsal horn neurons early in development (Saito et al., Mol. Cell. Neurosci. 6:280–92, 1995). Its absence, following a null mutation of its gene, leads to abnormalities in the spatio-temporal distribution of sensory neuron projections to the dorsal horn, as well as defects in dorsal horn morphogenesis (Chen et al., Neuron 31:59–73, 2001). These developmental abnormalities may account for a significantly attenuated sensitivity to noxious stimuli in the DRG11 deficient mice (Chen et al., supra).

DRG11—Responsive Gene Identification

A Genomic Binding Site (GBS) strategy was used in a mouse CpG island library to isolate genes responsive to the transcription factor DRG11 and to identify proteins that are involved in the development of sensory pathways (primary sensory neurons and spinal cord neurons) and other neurons (FIG. 1A). The general strategy isolates DRG11-binding fragments from mouse genomic DNA using a fusion protein (DRG-GST) consisting of a recombinant DRG11 DNA binding domain (amino acids 31–90 of mDRG-11) and GST. DRG11-responsive genes are located and isolated from the genomic region adjacent to the DRG-11 binding site. Mouse CpG islands are selected by the methyl-CpG binding domain of MeCP2, which binds DNA methylated at CpG and allows fractionation of DNA according to its degree of CpG methylation. The CpG library consists of short stretches of DNA containing a high density of nonmethylated CpG dinucleotides. About 60% of human genes are associated with CpG islands. These regions often include the promoter region and one or more exons of associated genes, allowing the isolation of full length cDNAs and genomic mapping.

The GBS cloning using the CpG island library was performed according to the method of Watanabe et al. (Mol. Cell. Biol. 18:442–449, 1998). Briefly, ten micrograms of the mouse library plasmid DNA was incubated with the DRG-GST fusion protein. The resulting solution was then passed slowly through a presoaked nitrocellulose filter and washed. The trapped plasmid DNA was eluted from the filter and transformed into DH5 α (*E. coli*) competent cells. The cells were cultured in Luria broth-ampicillin (LB) medium and plasmid DNA was prepared. The cycle was repeated for a total of three times. After the third cycle, the plasmid DNA library, enriched in genomic fragments that bind to the DRG11 DNA binding domain, was plated on LB-ampicillin agar plates, and individual clones were amplified, sequenced, and characterized.

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Identification and Characterization of Murine DRAGON

Among the most abundant clones obtained, was a 363 base pair (bp) DNA fragment located 750 bp upstream of an open reading frame of a novel gene. Sequence analysis studies indicated that the genomic fragment is located in the promoter region of the new gene. Genomic database analysis, combined with RT-PCR and RACE (Rapid Amplification of cDNA Ends) of mouse DRG and spinal cord cDNA libraries found that the open reading frame encoded a novel cDNA (SEQ ID NO: 1) that we have called DRAGON. The nucleotide and predicted 436 amino acid sequence of DRAGON (SEQ ID NO: 5) are shown in (FIG. 1B).

Sequence analysis of the mDRAGON coding region identified conserved domains with homology to notch-3 (FIG. 5), phosphatidylinositol-4-phosphate-5-kinase type II beta (FIG. 5), insulin-like growth factor binding protein-2 (IGFBP2; FIG. 3), thrombospondin (FIG. 6), ephrin type-B receptor 3 precursor (EPHB3; FIG. 4), and Slit-2 (FIG. 6), all of which are known to influence axonal guidance, neurite outgrowth, and other neuronal developmental functions. The C-terminus of mDRAGON is also predicted to contain a hydrophobic domain indicative of a 21 amino acid extracellular GPI anchoring. A computational structure-function analysis of mDRAGON reveals the presence of a putative signal peptide sequence (FIG. 2), indicating that the gene product is a secreted protein, and further supporting an extracellular localization.

Identification of DRAGON Homologs

Sequence homology analysis using a mouse genome database identified two murine genes homologous to DRAGON. The cDNA sequences of these homologs (mDL-1 and mDL-2) are provided in SEQ ID NO: 2 and SEQ ID NO: 3, respectively. The deduced polypeptide sequences are also provided (SEQ ID NO: 6 and 7). Sequence alignments indicating areas of homology between mDRAGON, mDL-1, and mDL-2 are shown in FIG. 8A. The GPI anchor sequence is predicated to be at the C-terminal 27 and 36 amino acids of mDL-1 and mDL-2, respectively.

DRG11 Induces DRAGON Expression

Following the initial identification of mDRAGON using GBS cloning, the mDRAGON promoter (363 bp fragment) was confirmed to be DRG11-responsive using the reporter gene assay generally described by Ogura et al. (Proc. Natl. Acad. Sci. USA, 92:392–396, 1995). The 363 bp fragment was subcloned into the PGL3-Promoter reporter vector containing an SV40 promoter upstream of the luciferase gene. DRG11 triggered a 6-fold increase in luciferase activity as compared to control (FIG. 7B), revealing the presence of one or several DRG11 response elements in the 363 base pair promoter fragment. No induction in luciferase activity was detected in the absence of DRG11, indicating that the enhancer activity of the DRAGON promoter fragment was DRG11 dependent.

Tissue Localization of Dragon Gene Expression

In situ hybridization was used to demonstrate that at E12.5 DRG11 and DRAGON expression overlaps (FIG. 7A). In the DRG most neurons express both DRG11 and DRAGON; in the spinal cord DRG11 and DRAGON are expressed in the same medial region adjacent to the ventricular zone (FIG. 7A). A pull down assay was carried out to confirm interaction of DRG11 with the 363 bp promoter fragment of DRAGON obtained with the GBS screening. The promoter fragment was pulled down by a GST-DRG11-DBD fusion protein but not GST (FIG. 7C). Finally,

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DRAGON mRNA expression in DRG11 null mutant embryonic mice was examined. DRAGON expression in the spinal cord and DRG were significantly reduced in DRG11^{-/-} mice compared to wildtype littermates (FIG. 7D).

DRAGON mRNA is expressed in embryonic and adult mouse DRGs, spinal cord and brain, with little or no expression in the liver and kidney, and low levels in the heart (FIGS. 7E, 9A, 9D, 9G, 9J, and 10). DRAGON expression begins early in development (at least E7) (FIG. 7F), much earlier than DRG11 (E12). Its expression is dynamically regulated in the PNS and CNS during development.

The relative tissue distribution pattern of DRAGON, DL-2 and DL-1 mRNA in mouse embryos (E14.5) indicate that DRAGON and DL-2, but not DL-1, are primarily expressed in the nervous system, and that DRAGON and DL-2 expression in the nervous system is largely non-overlapping (FIGS. 9G–9L). DRAGON is heavily expressed in DRG neurons and in the dorso-medial mantle layer of the spinal cord, with lower expression laterally and ventrally. DL-2 shows no expression in the DRG but strong expression in the spinal cord and brain. In the spinal cord, DL-2 is expressed in the midline, extending from the roof to the floor plate around the central canal in the ependymal layer, medial and ventral to DRAGON (FIGS. 9C and 9F). DL-2-expressing neurons are also present in the marginal layer and ventral horn. A complementary DRAGON and DL-2 expression pattern is also present in embryonic brain. DRAGON is expressed in the alar plate of the myelencephalon, in the marginal layer of the mesencephalon, and with lower intensity laterally, in the basal plate of the pons, and in the cerebellar primordia. DL-2 is expressed in the ependymal layers of the myelencephalon, mesencephalon and pons. DL-2 but not DRAGON is expressed in the telencephalic cortex, most intensely medially. DRAGON is heavily expressed in the diencephalon, except in the ependymal layer where DL-2 is heavily expressed. DRAGON is homogeneously expressed in the striatum whereas DL-2 is only expressed on its medial surface (FIGS. 9G–9L). DRAGON, but not DL-2, is expressed in the cortex of the future olfactory lobe, retina and olfactory epithelium (FIGS. 11 and 12). Both DRAGON and DL-2 are expressed in the cochlea.

The mDL-1 gene has a very specific expression pattern in the developing mouse embryo. Expression was restricted to muscle and cartilage tissues distributed along the whole organism, indicating a role in muscle and bone development (FIGS. 9B and 9E). A structure-function analysis of the mDL-1 protein sequence indicated the presence of a signal peptide suggesting that mDL-1, like mDRAGON, is a secreted factor.

DRAGON Protein Expression

A rabbit polyclonal antibody was raised against the peptide sequence TAAAHSALEDVEALHPRK (SEQ ID NO: 11; residues 388–405 of MDRAGON), present in the C-terminus of DRAGON, upstream of its hydrophobic tail. The antibody binds with high affinity to recombinant DRAGON expressed in HEK293T transfected cells, recognizing a band of 50 KDa in Western blots (FIG. 13A). Antibody specificity was confirmed by immunocytochemistry of DRAGON-expressing HED293T cells (FIG. 13B). Western blots of protein extracts from neonatal and adult DRG and DRG primary cultures show a similar band with an additional lower band of 40 KDa, indicating possible proteolytic cleavage of endogenous DRAGON. Treatment of HEK293T cells expressing DRAGON with PI-PLC results in the decrease of

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DRAGON detection on HEK cells and its release into the culture medium (FIG. 13C), indicating that DRAGON is GPI-anchored.

Immunohistochemistry confirms expression of DRAGON in the DRG, spinal cord and brain in the areas where DRAGON mRNA is found (FIG. 13D). In the adult DRG, DRAGON is more abundantly expressed in small neurons with unmyelinated axons than in medium and large myelinated neurons (A δ and A β -fibers) (FIG. 13D). In the adult spinal cord, DRAGON expression is most prominent in the superficial laminae of the dorsal horn (FIG. 13D). Immunohistochemical studies also demonstrated that the DRAGON protein is expressed in the E14.5 mouse retina and optic nerve (FIG. 18) and skin (FIG. 19).

DRAGON Promotes Cellular Adhesion

Cell surface GPI-anchored proteins, including the ephrins and tenascin, act as neuronal and non-neuronal cell adhesion molecules, binding to molecules expressed on neighboring cells or in the extracellular matrix. To examine whether DRAGON has a cell adhesion role, we measured the amount of adhesion between DRG neurons and HEK293 cells expressing recombinant DRAGON. DRAGON expression caused nearly a two-fold increase in the number of cultured DRG neurons that adhered to a monolayer of DRAGON-expressing HEK cells, compared to control HEK cells (FIGS. 14A–14D). Moreover, pretreatment of DRAGON-expressing HEK cells with PI-PLC resulted in only basal levels of DRG adhesion (FIGS. 14A–14D). These results may reflect homophilic or heterophilic DRAGON interactions with the endogenous DRAGON protein expressed on the surface of DRG neurons.

DRAGON Promotes Neuronal Survival

The anti-DRAGON polyclonal antibody was added to neonatal rat DRG neuronal cultures to investigate the contribution of DRAGON to neuronal survival. Neuronal cultures were treated with 0.25% anti-DRAGON serum, 0.25% pre-immune serum (negative control), or vehicle. A statistically significant 20–25% increase in neuronal cell death was measured following anti-DRAGON treatment compared to controls.

	0.25% anti-DRAGON serum	0.25% pre-immune serum	Vehicle Control (no serum)
% viable neurons (mean)	41.8%	55.3%	51.8%
Standard Error (S.E.)	1.7%	2.3%	2.5%
Number of isolated DRG cultures (n)	12	12	11

Neural Induction in *Xenopus* Embryos

In order to determine whether DRAGON affects cell differentiation and early embryonic development, DRAGON was injected into one cell at the animal pole of *Xenopus* embryos at the 2-cell stage. Embryos were allowed to develop until early tadpole stages. By injecting one out of two cells, a control side and an experimental side are present in the same embryo. A variety of markers were measured, including twist (expressed in anterior neural crest cells) and N-tubulin (a general neuronal differentiation marker), to determine whether DRAGON affects early neural patterning. Ectopic DRAGON caused a decrease in neural crest

derivatives, as shown by loss of twist expression (FIG. 15A, top panels) and an increase in neuronal markers (FIG. 15A, bottom panels).

In ectodermal explant assays, DRAGON induced anterior neural markers (FIG. 15B). *Nrp1* is a pan-neural marker, *Otx2* is expressed within the forebrain and midbrain regions, and *XAG* is expressed in the cement gland (the most anterior structure in the tadpole). In addition, DRAGON induced *nkx2.5*, an early marker of cardiac development.

Identification of Dragon Homologs in Other Species

Zebrafish Dragon Genes

The cDNA and polypeptide sequences of zebrafish homologs of DRAGON (SEQ ID NO: 25 and 28), DL-1 (SEQ ID NO: 26 and 29), and DL-2 (SEQ ID NO: 27 and 30) are provided. The sequence and domain structure of the three zebrafish genes are highly conserved with the mouse genes (70–75% homology) and Northern blot analysis shows a single transcript in each case. (FIG. 16A). DRAGON mRNA is present at the 2–4 cell stage of zebrafish embryogenesis, which is prior to initiation of zygotic transcription, suggesting a maternal or early developmental role for the protein. After the mid-blastula transition, the levels of DRAGON mRNA increase and are then maintained at a high level for up to 72 hours post fertilization, the latest stage examined (FIG. 16A).

In-situ hybridization reveals widespread and strong DRAGON expression in the zebrafish embryo. At the 18-somite stage, DRAGON is expressed along the midline in the telencephalon, diencephalon, and mesencephalon (FIGS. 16B and 16C). DRAGON is also expressed in the developing retina (FIG. 16D).

Overexpression of DRAGON in zebrafish embryos following sense injection into the fertilized egg, leads to abnormalities in the morphology of the brain and eye in 75–85% of treated embryos. The most common features include abnormal ventricle development, inappropriate cell death, particularly in the hindbrain, and neural tube twisting. DRAGON overexpression results in cyclopedia in 10–20% of embryos. The single eye is in an abnormally ventral location, with the anterior portion of the brain being dorsal to the eye (FIGS. 16E–16G).

Embryos injected with a morpholino antisense oligonucleotide directed against the splice-donor site of the first exon of DRAGON show extensive CNS degeneration with a failure of development of the forebrain, hindbrain, and spinal cord (FIGS. 16I–16M). The knockdown of DRAGON splicing and expression was confirmed by RT-PCR and compared-to controls (FIG. 16H). Injected embryos had extensive apoptotic cell death in the brain, the brainstem and along the entire rostro-caudal extent of the spinal cord, as assessed by TUNEL assay and acridine orange staining. An inverted control oligonucleotide had no effect.

During early development, DL-1 shows high expression in the notochord and the adjacent adaxial cells (the earliest cells to develop into muscle fibers). Subsequently, DL-1 is expressed exclusively in the somites.

DL-2 mRNA first appears during zebrafish development at the three somite stage (approximately 10 hours postfertilization). DL-2 expression peaks at 18 hours, followed by a decrease over the next 72 hours.

Human Dragon Genes

The human homologs of all three murine Dragon gene family have been identified using the human genomic Celera

homologs (SEQ ID NO: 8–10) are about 90% identical to the murine Dragon proteins (SEQ ID NO: 5–7).

C. elegans DRAGON

Strong conservation of many domains present in members of the Dragon family among different species has also enabled us to identify the *C. elegans* ortholog (FIG. 17; SEQ ID NO: 18). The strong domain conservation pattern suggests a crucial role in development for the different members of this family.

Identification of Dragon Genes in Other Species

Homologs from other species can easily be identified based on sequence identity with the Dragon proteins and nucleic acids disclosed herein. Sequence identity may be measured using sequence analysis software on the default setting (e.g., Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wis. 53705). Such software may match similar sequences by assigning degrees of homology to various substitutions, deletions, and other modifications. Conservative substitutions typically include substitutions within the following groups: glycine, alanine, valine, isoleucine, leucine; aspartic acid, glutamic acid, asparagine, glutamine; serine, threonine; lysine, arginine; and phenylalanine, tyrosine.

Multiple sequences may also be aligned using the Clustal W(1.4) program (produced by Julie D. Thompson and Toby Gibson of the European Molecular Biology Laboratory, Germany and Desmond Higgins of European Bioinformatics Institute, Cambridge, UK) by setting the pairwise alignment mode to “slow,” the pairwise alignment parameters to include an open gap penalty of 10.0 and an extend gap penalty of 0.1, as well as setting the similarity matrix to “blosum.” In addition, the multiple alignment parameters may include an open gap penalty of 10.0, an extend gap penalty of 0.1, as well as setting the similarity matrix to “blosum,” the delay divergent to 40%, and the gap distance to 8.

In Situ Hybridization

The in situ hybridization methods used herein have been described previously (Karchewski et al., J. Comp. Neurol. 413:327, 1999). Hybridization was performed on fresh frozen, mounted tissue sections from mouse embryo and adult rat dorsal root ganglia (DRG) using terminally-labeled oligonucleotide probes. Probes had approximately 50% G-C content and were complementary and selective for mDRAGON mRNAs. Probes were 3'-end labeled with ³⁵S-dATP using a terminal transferase reaction and purified through a spin column (Qiagen). Hybridization was done under very high stringency conditions such that probe annealing required at least 90% sequence identity (Dagerlind et al., Histochemistry 98:39, 1992).

Briefly, slides were brought to room-temperature and covered with a hybridization solution (50% formamide, 1×Denhardt's solution, 1% sarcosyl, 10% dextran sulphate, 0.02M phosphate buffer, 4×SSC, 200 nM DTT, 500 mg/ml salmon sperm DNA) containing 10⁷ epm/ml of labeled probe. Slides were incubated in a humidified chamber at 43° C. for 14–18 hours, then washed 4×15min in 1×SSC at 55° C. In the final rinse, slides were brought to room temperature, washed in dH₂O, dehydrated in ethanol, and air dried.

Autoradiograms were generated by dipping slides in NTB2 nuclear track emulsion and storing in the dark at 4° C. Prior to conventional developing and fixation, sections were allowed to expose for 1–3 weeks, depending on the abundance of transcript. Unstained tissue was viewed under darkfield conditions using a fiber-optic darkfield stage

adapter (MVI), while stained tissue was examined under brightfield conditions. Control experiments using sense probes were conducted to confirm the specificity of hybridization. The antisense oligonucleotide probes are as follows:

mDRAGON-specific for nucleotides 831-879 of SEQ ID NO: 1:

5'-TCG CAC AAA CAC TGT GGT GCC TAT GTA GCG GGC ATG CAT CTC TAC GTA-3'. (SEQ ID NO: 19)

mDL-1-specific for nucleotides 913-960 of SEQ ID NO: 2:

5'-CCC AGC TGT CTG TCG AAT GAT GAT AGT TGT TCC AAT GTA GGC AGC TCG-3' (SEQ ID NO: 20)

mDL-2-specific for nucleotides 1252-1299 of SEQ ID NO: 3:

5'-TTG CCA TCC TCC AAA GCA TAG TAG GCA GCC AGC GTG AAG TTC ACA TCA-3'. (SEQ ID NO: 21)

Synthesis of Dragon Proteins

Nucleic acids that encode Dragon family proteins or fragments thereof may be introduced into various cell types or cell-free systems for expression, thereby allowing purification of these Dragon proteins for biochemical characterization, large-scale production, antibody production, and patient therapy.

Eukaryotic and prokaryotic Dragon expression systems may be generated in which a Dragon family gene sequence is introduced into a plasmid or other vector, which is then used to transform living cells. Constructs in which the Dragon cDNA contains the entire open reading frame inserted in the correct orientation into an expression plasmid may be used for protein expression. Alternatively, portions of the Dragon gene sequences, including wild-type or mutant Dragon sequences, may be inserted. Prokaryotic and eukaryotic expression systems allow various important functional domains of the Dragon proteins to be recovered, if desired, as fusion proteins, and then used for binding, structural, and functional studies and also for the generation of appropriate antibodies.

Typical expression vectors contain promoters that direct the synthesis of large amounts of mRNA corresponding to the inserted Dragon nucleic acid in the plasmid-bearing cells. They may also include a eukaryotic or prokaryotic origin of replication sequence allowing for their autonomous replication within the host organism, sequences that encode genetic traits that allow vector-containing cells to be selected for in the presence of otherwise toxic drugs, and sequences that increase the efficiency with which the synthesized mRNA is translated. Stable long-term vectors may be maintained as freely replicating entities by using regulatory elements of, for example, viruses (e.g., the OriP sequences from the Epstein Barr Virus genome). Cell lines may also be produced that have integrated the vector into the genomic DNA, and in this manner the gene product is produced on a continuous basis.

Expression of foreign sequences in bacteria, such as *Escherichia coli*, requires the insertion of the Dragon nucleic acid sequence into a bacterial expression vector. Such plasmid vectors contain several elements required for the propagation of the plasmid in bacteria, and for expression of the DNA inserted into the plasmid. Propagation of only plasmid-bearing bacteria is achieved by introducing, into the plasmid, selectable marker-encoding sequences that allow plasmid-bearing bacteria to grow in the presence of otherwise toxic drugs. The plasmid also contains a transcriptional promoter capable of producing large amounts of mRNA from the cloned gene. Such promoters may be (but are not necessarily) inducible promoters that initiate transcription upon induction. The plasmid also preferably con-

tains a polylinker to simplify insertion of the gene in the correct orientation within the vector.

Mammalian cells can also be used to express a Dragon protein. Stable or transient cell line clones can be made

15 using Dragon expression vectors to produce Dragon proteins in a soluble (truncated and tagged) or membrane anchored (native) form. Appropriate cell lines include, for example, COS, HEK293T, CHO, or NIH cell lines.

20 Once the appropriate expression vectors containing a Dragon gene, fragment, fusion, or mutant are constructed, they are introduced into an appropriate host cell by transformation techniques, such as, but not limited to, calcium phosphate transfection, DEAE-dextran transfection, electroporation, microinjection, protoplast fusion, or liposome-mediated transfection. The host cells that are transfected with the vectors of this invention may include (but are not limited to) *E. coli* or other bacteria, yeast, fungi, insect cells (using, for example, baculoviral vectors for expression in SF9 insect cells), or cells derived from mice, humans, or other animals. In vitro expression of Dragon proteins, fusions, polypeptide fragments, or mutants encoded by cloned DNA may also be used. Those skilled in the art of molecular biology will understand that a wide variety of expression systems and purification systems may be used to produce recombinant Dragon proteins and fragments thereof. Some of these systems are described, for example, in Ausubel et al. (supra).

40 Once a recombinant protein is expressed, it can be isolated from cell lysates using protein purification techniques such as affinity chromatography. Once isolated, the recombinant protein can, if desired, be purified further by e.g., by high performance liquid chromatography (HPLC; e.g., see Fisher, Laboratory Techniques In Biochemistry And Molecular Biology, Work and Burdon, Eds., Elsevier, 1980).

45 Polypeptides of the invention, particularly short Dragon fragments can also be produced by chemical synthesis (e.g., by the methods described in Solid Phase Peptide Synthesis, 2nd ed., 1984, The Pierce Chemical Co., Rockford, IL).

50 Dragon Fusion Proteins

Also included in the invention are Dragon family proteins fused to heterologous sequences, such as detectable markers (for example, proteins that may be detected directly or indirectly such as green fluorescent protein, hemagglutinin, or alkaline phosphatase), DNA binding domains (for example, GAL4 or LexA), gene activation domains (for example, GAL4 or VP16), purification tags, or secretion signal peptides. These fusion proteins may be produced by any standard method. For production of stable cell lines expressing a Dragon fusion protein, PCR-amplified Dragon nucleic acids may be cloned into the restriction site of a derivative of a mammalian expression vector. For example, KA, which is a derivative of pcDNA3 (Invitrogen, Carlsbad, Calif.) contains a DNA fragment encoding an influenza virus hemagglutinin (HA). Alternatively, vector derivatives encoding other tags, such as c-myc or poly Histidine tags, can be used.

The Dragon expression construct may be co-transfected, with a marker plasmid, into an appropriate mammalian cell line (e.g. COS, HEK293T, or NIH 3T3 cells) using, for example, Lipofectamine™ (Gibco-BRL, Gaithersburg, Md.) according to the manufacturer's instructions, or any other suitable transfection technique known in the art. Suitable transfection markers include, for example, β -galactosidase or green fluorescent protein (GFP) expression plasmids or any plasmid that does not contain the same detectable marker as the Dragon fusion protein. The Dragon-expressing cells can be sorted and further cultured, or the tagged Dragon can be purified.

In one particular example, a DRAGON open reading frame (ORF) was amplified by polymerase chain reaction (PCR) using standard techniques and primers containing restriction sites (e.g. Sal I sites). The top strand primer consisted of the sequence 5'-ATA AGC TTA TGG GCG TGAGAG CAG CAC CTT CC-3' (SEQ ID NO: 22) and the bottom strand primer consisted of the sequence 5'-GAA GTC GAC GAAACA ACT CCT ACA AAAAC-3' (SEQ ID NO: 23). DRAGON cDNA was also amplified without the signal peptide and subcloned into a vector (pSecTagHis) having a strong secretion signal peptide. The same bottom strand primer was used (SEQ ID NO: 23); however, the top strand primer was substituted for one having the sequence 5'-CTC AAG CTT CAG CCT ACT CAA TGC CGA ATC-3' (SEQ ID NO: 24).

In another example, we generated DRAGON-alkaline phosphatase (AP) fusion protein using the mammalian expression vector, pAPtag-5' (Flanagan et al., *Meth. Enzymol.* 327:198-210, 2000). When expressed in mammalian cells (e.g. HEK 293), the DRAGON-AP fusion protein is secreted at high levels into the culture medium and is easily detected by the AP activity assay. The resulting DRAGON-AP fusion protein can be used to screen expression libraries to identify, clone, sequence, and characterize molecules which interact with DRAGON, such as cell surface receptors or endogenous DRAGON ligands. Of course, this method is broadly applicable to all Dragon-family proteins and can be used in conjunction with any number of suitable tags known in the art.

Interaction Trap Assays

Two-hybrid methods, and modifications thereof, may also be used to identify novel proteins that interact with Dragon-family proteins, and hence may be naturally occurring Dragon ligands or receptors. In addition, regulators of Dragon, e.g., proteins that interfere with or enhance the interaction between Dragon and other proteins, may be identified by the use of a three-hybrid system. Such assays are well-known to skilled artisans, and may be found, for example, in Ausubel et al. (*supra*).

Generation of Anti-Dragon Antibodies

In order to prepare polyclonal antibodies, Dragon family proteins, fragments, or fusion proteins containing defined portions of Dragon proteins may be synthesized in bacterial, fungal, or mammalian cells by expression of corresponding DNA sequences in a suitable cloning vehicle. The proteins can be purified, coupled to a carrier protein, mixed with Freund's adjuvant (to enhance stimulation of the antigenic response in an inoculated animal), and injected into rabbits or other laboratory animals. Following booster injections at bi-weekly intervals, the rabbits or other laboratory animals are then bled and the sera isolated. The sera can be used directly or can be purified prior to use by various methods, including affinity chromatography employing reagents such as Protein A-Sepharose, antigen-Sepharose, and anti-mouse-

Ig-Sepharose. The sera can then be used to probe protein extracts from Dragon-expressing tissue electrophoretically fractionated on a polyacrylamide gel to identify Dragon proteins. Alternatively, synthetic peptides can be made that correspond to the antigenic portions of the protein and used to inoculate the animals. As described above, a polyclonal antibody against mDRAGON was created using, as the immunogenic DRAGON fragment, a polypeptide corresponding to residues 388-405 of SEQ ID NO: 5. Suitable immunogens for creating anti-hDRAGON antibodies include, for example, the polypeptide sequences encoded by residues 54-72, 277-294, or 385-408 of SEQ ID NO: 8.

Alternatively, monoclonal antibodies may be prepared using Dragon proteins described above and standard hybridoma technology (see, e.g., Kohler et al., *Nature* 256:495, 1975; Kohler et al., *Eur. J. Immunol.* 6:511, 1976; Kohler et al., *Eur. J. Immunol.* 6:292, 1976; Hammerling et al., *In Monoclonal Antibodies and T Cell Hybridomas*, Elsevier, New York, N.Y., 1981). Once produced, monoclonal antibodies are also tested for specific Dragon protein recognition by Western blot or immunoprecipitation analysis.

Antibodies of the invention may also be produced using Dragon amino acid sequences that do not reside within highly conserved regions, and that appear likely to be antigenic, as analyzed by criteria such as those provided by the Peptide Structure Program (Genetics Computer Group Sequence Analysis Package, Program Manual for the GCG Package, Version 7, 1991) using the algorithm of Jameson and Wolf (*CABIOS* 4:181, 1988).

Use of Dragon Proteins and Nucleic Acids in Diagnosis

Dragon family proteins may be used in diagnosing existing disorders or the propensity for developing disorders of the nervous system (DRAGON and DL-2) or bone, muscle, skin, joint, and cartilage tissue (DL-1), where a decrease or increase in the level of Dragon protein or nucleic acid production, relative to a control, provides an indication of a deleterious condition. Alternatively, a patient sample may be analyzed for one or more alterations in a Dragon nucleic acid sequence, compared to a wild-type Dragon sequence, using a mismatch detection approach. The alteration in the Dragon sequence need not be in a coding region. Alterations in, for example, promoter regions can result in alterations of Dragon protein levels and/or tissue distribution. Wild-type Dragon nucleic acid sequences for use in this assay include SEQ ID NO: 1-4 and 31-32.

Generally, these techniques involve PCR amplification of nucleic acid from the patient sample, followed by identification of the mutation (e.g., mismatch) by either altered hybridization, aberrant electrophoretic gel migration, binding or cleavage mediated by mismatch binding proteins, or direct nucleic acid sequencing. Any of these techniques may be used to facilitate mutant Dragon detection, and each is well known in the art (see, for example, Orita et al., *Proc. Natl. Acad. Sci. USA* 86:2766-2770, 1989; and Sheffield et al., *Proc. Natl. Acad. Sci. USA* 86:232-236, 1989).

Mismatch detection assays may be used to diagnose a Dragon nucleic acid-mediated predisposition to a nervous system, bone, muscle, skin or cartilage condition. For example, a patient heterozygous for a Dragon mutation may show no clinical symptoms and yet possess a higher than normal probability of developing one or more types of these diseases. Given this diagnosis, a patient may take precautions to control their exposure to adverse environmental factors and to carefully monitor their medical condition (for example, through frequent physical examinations). This

type of Dragon diagnostic approach may also be used to detect Dragon nucleic acid mutations in prenatal screens.

Measurement of Dragon RNA is also a useful diagnostic. For example, a decrease in a Dragon mRNA or protein in a subject, relative to a control subject, would suggest a diagnosis of the presence or propensity for acquiring a disorder of the nervous system, or the bone, muscle, skin, or joint tissue. In addition, a decrease in Dragon mRNA or protein, relative to control, may correlate with a poor prognosis for treatment of these conditions using a non-Dragon therapy.

Levels of Dragon protein or nucleic acid expression may be assayed by any standard technique and compared to control samples showing normal Dragon protein or nucleic acid expression. For example, expression in a biological sample (e.g., a biopsy) may be monitored by standard Northern blot analysis, using, for example, probes designed from a Dragon nucleic acid. Measurement of such expression may be aided by PCR (see, e.g., Ausubel et al., supra; PCR Technology: Principles and Applications for DNA Amplification, ed., H. A. Ehrlich, Stockton Press, NY; and Yap and McGee, Nucl. Acids Res. 19:4294, 1991).

In yet another approach, immunoassays may be used to detect or monitor a Dragon protein in a biological sample. Dragon-specific polyclonal or monoclonal antibodies may be used in any standard immunoassay format (e.g., ELISA, Western blot, or RIA assay) to measure Dragon levels; again comparison is to wild-type Dragon levels. Examples of immunoassays are described, e.g., in Ausubel et al. (supra). Immunohistochemical techniques may also be utilized for Dragon detection. For example, a tissue sample may be obtained from a patient, and a section stained for the presence of a Dragon protein using an antibody against that protein and any standard detection system (e.g., one which includes a secondary antibody conjugated to horseradish peroxidase). General guidance regarding such techniques can be found in, e.g., Bancroft and Stevens (Theory and Practice of Histological Techniques, Churchill Livingstone, 1982) and Ausubel et al. (supra).

Identification of Candidate Compounds for Treatment of Dragon-related Conditions

A candidate compound that is beneficial in the treatment, stabilization, or prevention of a Dragon-related condition (e.g. disorders of the nervous system, retina, skin, and bone, muscle, joint, or cartilage tissue) can be identified by the methods of the present invention. A candidate compound can be identified for its ability to affect the biological activity of a Dragon protein or the expression of a Dragon gene or to mimic its action. Compounds that are identified by the methods of the present invention that increase the biological activity or expression levels of a Dragon protein or that compensate for the loss of Dragon protein activity or gene expression, for example, due to loss of the Dragon gene due to a genetic lesion, can be used in the treatment or prevention of a Dragon-related condition. A candidate compound identified by the present invention can mimic the biological activity of a Dragon protein, bind a Dragon protein, modulate (e.g., increase or decrease) transcription of a Dragon gene, or modulate translation of a Dragon mRNA.

Any number of methods are available for carrying out screening assays to identify new candidate compounds that promote the expression of a Dragon gene. In one working example, candidate compounds are added at varying concentrations to the culture medium of cultured cells expressing one of the Dragon nucleic acid sequences of the invention. Gene expression is then measured, for example, by microarray analysis, Northern blot analysis (Ausubel et al.,

supra), or RT-PCR, using any appropriate fragment prepared from the nucleic acid molecule as a hybridization probe. The level of Dragon gene expression in the presence of the candidate compound is compared to the level measured in a control culture medium lacking the candidate compound. A compound which promotes an increase in the expression of a Dragon gene is considered useful in the invention and may be used as a therapeutic to treat a human patient.

In another working example, the effect of candidate compounds may be measured at the level of Dragon protein production using the same general approach and standard immunological techniques, such as Western blotting or immunoprecipitation with an antibody specific for a Dragon protein. For example, immunoassays may be used to detect or monitor the expression of at least one of the polypeptides of the invention in an organism. Polyclonal or monoclonal antibodies that are capable of binding to a Dragon protein may be used in any standard immunoassay format (e.g., ELISA, Western blot, or RIA assay) to measure the level of the protein. In some embodiments, a compound that promotes an increase in Dragon expression or biological activity is considered particularly useful.

Expression of a reporter gene that is operably linked to a Dragon promoter can also be used to identify a candidate compound for treating or preventing a Dragon-related condition. Assays employing the detection of reporter gene products are extremely sensitive and readily amenable to automation, hence making them ideal for the design of high-throughput screens. Assays for reporter genes may employ, for example, calorimetric, chemiluminescent, or fluorometric detection of reporter gene products. Many varieties of plasmid and viral vectors containing reporter gene cassettes are easily obtained. Such vectors contain cassettes encoding reporter genes such as lacZ/ β -galactosidase, green fluorescent protein, and luciferase, among others. A genomic DNA fragment carrying a Dragon-specific transcriptional control region (e.g., a promoter and/or enhancer) is first cloned using standard approaches (such as those described by Ausubel et al. (supra)). The DNA carrying the Dragon transcriptional control region is then inserted, by DNA subcloning, into a reporter vector, thereby placing a vector-encoded reporter gene under the control of the Dragon transcriptional control region. The activity of the Dragon transcriptional control region operably linked to the reporter gene can then be directly observed and quantified as a function of reporter gene activity in a reporter gene assay.

In one embodiment, for example, the Dragon transcriptional control region could be cloned upstream from a luciferase reporter gene within a reporter vector. This could be introduced into the test cells, along with an internal control reporter vector (e.g., a lacZ gene under the transcriptional regulation of the β -actin promoter). After the cells are exposed to the test compounds, reporter gene activity is measured and Dragon reporter gene activity is normalized to internal control reporter gene activity.

In addition, candidate compounds may be identified using any of the Dragon fusion proteins described above (e.g., as compounds that bind to those fusion proteins), or by any of the two-hybrid or three-hybrid assays described above.

A candidate compound identified by the methods of the present invention can be from natural as well as synthetic sources. Those skilled in the field of drug discovery and development will understand that the precise source of test extracts or compounds is not critical to the methods of the invention. Examples of such extracts or compounds include, but are not limited to, plant-, fungal-, prokaryotic-, or animal-based extracts, fermentation broths, and synthetic

compounds, as well as modification of existing compounds. Numerous methods are also available for generating random or directed synthesis (e.g., semi-synthesis or total synthesis) of any number of chemical compounds, including, but not limited to, saccharide-, lipid-, peptide-, and nucleic acid-based compounds. Synthetic compound libraries are commercially available from Brandon Associates (Merrimack, N.H.) and Aldrich Chemical (Milwaukee, Wis.). Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant, and animal extracts are commercially available from a number of sources, including Biotics (Sussex, UK), Xenova (Slough, UK), Harbor Branch Oceanographic Institute (Ft. Pierce, Fla.), and PharmaMar, U.S.A. (Cambridge, Mass.). In addition, natural and synthetically produced libraries are produced, if desired, according to methods known in the art, e.g., by standard extraction and fractionation methods. Furthermore, if desired, any library or compound is readily modified using standard chemical, physical, or biochemical methods.

Use of Transgenic Animals to Identify a Candidate Compound

The present invention also provides methods for using transgenic and knockout animals that develop a Dragon-related condition and accurately recapitulate many of the features of the Dragon-related condition associated with loss or mutation of a Dragon gene. Desirably, the Dragon gene is used to produce the transgenic animal or the Dragon gene is the target of the knockout. However, other genes involved in or related to Dragon expression or activity, may also be used to produce transgenic animals so that the effect on a Dragon-related condition may be studied in this context.

A transgenic animal expressing a mutant Dragon gene can be used to identify candidate compounds that are useful for the treatment or prevention of a Dragon-related condition. Transgenic animals expressing a conditional mutant Dragon gene (e.g., using a tetracycline regulatable system) can also be generated by methods well known to those skilled in the art; such methods are described in, for example, WO 94/29442, WO 96/40892, WO 96/01313, and Yamamoto et al. (Cell 101:57–66, 2000). In addition, the knockout animal may be a conditional knockout using, for example, the FLP/FRT system described in, for example, U.S. Pat. No. 5,527,695, and in Lyznik et al. (Nucleic Acid Research 24:3784–3789, 1996) or the Cre-lox recombination system described, for example, in Kilby et al. (Trends in Genetics 9:413–421, 1993).

Transgenic animals may be made using standard techniques, such as those described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, N.Y., 1989). Any tissue specific promoter may direct the expression of any Dragon protein used in the invention, such as neuron-specific promoters, muscle-specific promoters, skin-specific promoters, retina-specific promoters, and bone-specific promoters.

The disclosed transgenic and knock-out animals may be used as research tools to determine genetic and physiological features of a Dragon-related condition, and for identifying compounds that can affect such conditions. Knockout animals also include animals where the normal Dragon gene(s) has been inactivated or removed and replaced with a polymorphic allele of this gene. These animals can serve as a model system for the risk of developing, treating, stabilizing, or preventing a Dragon-related condition that is associated with a Dragon gene polymorphism or mutation.

In general, a transgenic or knockout animal can be used to identify a candidate compound useful for treating or

preventing a Dragon-related condition by contacting the transgenic or knockout animal with the candidate compound and comparing the presence, absence, or level of expression of genes, either at the RNA level or at the protein level, in tissue from a transgenic or knockout animal as described above, and tissue from a matching non-transgenic or knockout animal. Standard techniques for detecting RNA expression, e.g., by Northern blotting, or protein expression, e.g., by Western blotting, are well known in the art. The response to or progression of disease in a transgenic or knockout animal, as compared with non-transgenic or knockout animals can be used to identify compounds that may be effective therapeutics against a Dragon-related condition, such as nervous system disorders or disorders of muscle, skin, bone, or cartilage tissue. Transgenic and knockout animals can also be used to predict whether compounds identified as therapeutics will affect disease progression.

Any transgenic animal, or cells derived from these animals, may be constructed and used for compound screening. Preferable animal models include, without limitation, mice, rats, rabbits, and flies.

Regulation of Stem Cell Fate Using Dragon Family Proteins

Differentiation of stem cells, particularly ES cells, can be accomplished by exposing the cells to supraphysiological concentrations of Dragon proteins. Specifically, DRAGON or DL-2 can induce a stem cell to adopt a neuronal phenotype, whereas DL-1 promotes myogenic phenotypes. Stem cell differentiation may be accomplished using any appropriate technique. For example, transgenic stem cells overexpressing a Dragon protein can be created. Preferably, the Dragon gene is operably linked to an inducible promoter in order to control the timing and level of Dragon protein expression. Alternatively, stem cell differentiation can be done by the treatment with an exogenous Dragon protein. Typically, a recombinant Dragon protein is produced using a non-stem cell line (e.g., CHO cells), the Dragon protein is isolated, and the stem cells are treated in vitro with the protein.

Dragon-induced stem cell differentiation into neuronal phenotypes can be facilitated by blocking competing differentiation pathways (e.g., pathways that lead to differentiation into cell types of mesodermal or endodermal origin). Examples of these competing pathways include, but are not limited to, signaling pathways for TGF- β superfamily members (Nodal, Activin, and bone morphogenic proteins (BMPs) 2, 4, and 7), which have been shown to be important for endoderm and mesoderm differentiation (*Nature Reviews Neurosci.* 3: 271–280). By inhibiting any one of these TGF- β family members that lead to endoderm or mesoderm differentiation, differentiation into a cell of neuroectodermal origin is favored.

It will be apparent to one of skill in the art that the timing and extent of TGF- β pathway inhibition and overexpression or application of a Dragon protein will vary depending on the methods and dosages used. For example, inhibition of a of TGF- β pathway by gene knockout technology persists throughout the lifetime of an ES cell, whereas inhibition of the same pathway via antisense oligonucleotides is generally transient such that antisense oligonucleotides need to be reapplied. In the latter case, one of skill in the art would be able to readily determine when and how much of the antisense oligonucleotide to reapply to promote neuronal differentiation.

Stem cells that have been induced to differentiate along a neuronal (using DRAGON or DL-2) or myogenic (using DL-1) lineage can be transplanted into a patient in need of

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cell replacement therapy. For example, patients diagnosed as having neurodegenerative diseases (e.g., Alzheimer's disease, Parkinson's disease, and Huntington's disease) can be treated by transplanting, into affected brain regions, stem cells that have been induced to differentiate along a neuronal lineage by exposure to DRAGON or DL-2. Stem cells treated with DL-1 to induce myogenic differentiation can be transplanted into patients diagnosed as having a muscle wasting disease such as muscular dystrophy, myotonia congenital, or myotonic dystrophy.

Administration of a Dragon Protein or a Candidate Compound for the Treatment or Prevention of a Dragon-related Condition

The present invention also includes the administration of a Dragon family protein for the treatment or prevention of a Dragon-related condition. The administration of a biologically active Dragon protein that, regardless of its method of manufacture, retains full biological activity, can be envisioned as restoring Dragon biological activity in a patient lacking endogenous activity of a Dragon protein due to a loss or reduction of expression or biological activity, e.g., by mutation or loss of a Dragon gene or cells that normally express a Dragon.

Peptide agents of the invention, such as a Dragon protein, can be administered to a subject, e.g., a human, directly or in combination with any pharmaceutically acceptable carrier or salt known in the art. Pharmaceutically acceptable salts may include non-toxic acid addition salts or metal complexes that are commonly used in the pharmaceutical industry. Examples of acid addition salts include organic acids such as acetic, lactic, pamoic, maleic, citric, malic, ascorbic, succinic, benzoic, palmitic, suberic, salicylic, tartaric, methanesulfonic, toluenesulfonic, or trifluoroacetic acids or the like; polymeric acids such as tannic acid, carboxymethyl cellulose, or the like; and inorganic acids such as hydrochloric acid, hydrobromic acid, sulfuric acid phosphoric acid, or the like. Metal complexes include zinc, iron, and the like. One exemplary pharmaceutically acceptable carrier is physiological saline. Other physiologically acceptable carriers and their formulations are known to one skilled in the art and described, for example, in Remington's Pharmaceutical Sciences, (19th edition), ed. A. Gennaro, 1995, Mack Publishing Company, Easton, Pa.

Pharmaceutical formulations of a therapeutically effective amount of a peptide agent or candidate compound of the invention, or pharmaceutically acceptable salt thereof, can be administered orally, parenterally (e.g. intramuscular, intraperitoneal, intravenous, or subcutaneous injection), or by intrathecal or intracerebroventricular injection in an admixture with a pharmaceutically acceptable carrier adapted for the route of administration.

Methods well known in the art for making formulations are found, for example, in Remington's Pharmaceutical Sciences (19th edition), ed. A. Gennaro, 1995, Mack Publishing Company, Easton, Pa. Compositions intended for oral use may be prepared in solid or liquid forms according to any method known to the art for the manufacture of pharmaceutical compositions. The compositions may optionally contain sweetening, flavoring, coloring, perfuming, and/or preserving agents in order to provide a more palatable preparation. Solid dosage forms for oral administration include capsules, tablets, pills, powders, and granules. In such solid forms, the active compound is admixed with at least one inert pharmaceutically acceptable carrier or excipient. These may include, for example, inert diluents, such as calcium carbonate, sodium carbonate, lactose,

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sucrose, starch, calcium phosphate, sodium phosphate, or kaolin. Binding agents, buffering agents, and/or lubricating agents (e.g., magnesium stearate) may also be used. Tablets and pills can additionally be prepared with enteric coatings.

Liquid dosage forms for oral administration include pharmaceutically acceptable emulsions, solutions, suspensions, syrups, and soft gelatin capsules. These forms contain inert diluents commonly used in the art, such as water or an oil medium. Besides such inert diluents, compositions can also include adjuvants, such as wetting agents, emulsifying agents, and suspending agents.

Formulations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, or emulsions. Examples of suitable vehicles include propylene glycol, polyethylene glycol, vegetable oils, gelatin, hydrogenated naphthalenes, and injectable organic esters, such as ethyl oleate. Such formulations may also contain adjuvants, such as preserving, wetting, emulsifying, and dispersing agents. Biocompatible, biodegradable lactide polymer, lactide/glycolide copolymer, or polyoxyethylene-polyoxypropylene copolymers may be used to control the release of the compounds. Other potentially useful parenteral delivery systems for the proteins of the invention include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes.

Liquid formulations can be sterilized by, for example, filtration through a bacteria-retaining filter, by incorporating sterilizing agents into the compositions, or by irradiating or heating the compositions. Alternatively, they can also be manufactured in the form of sterile, solid compositions which can be dissolved in sterile water or some other sterile injectable medium immediately before use.

The amount of active ingredient in the compositions of the invention can be varied. One skilled in the art will appreciate that the exact individual dosages may be adjusted somewhat depending upon a variety of factors, including the protein being administered, the time of administration, the route of administration, the nature of the formulation, the rate of excretion, the nature of the subject's conditions, and the age, weight, health, and gender of the patient. Generally, dosage levels of between 0.1 $\mu\text{g/kg}$ to 100 mg/kg of body weight are administered daily as a single dose or divided into multiple doses. Desirably, the general dosage range is between 250 $\mu\text{g/kg}$ to 5.0 mg/kg of body weight per day. Wide variations in the needed dosage are to be expected in view of the differing efficiencies of the various routes of administration. For instance, oral administration generally would be expected to require higher dosage levels than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, which are well known in the art. In general, the precise therapeutically effective dosage will be determined by the attending physician in consideration of the above identified factors.

The protein or candidate compound of the invention can be administered in a sustained release composition, such as those described in, for example, U.S. Pat. No. 5,672,659 and U.S. Pat. No. 5,595,760. The use of immediate or sustained release compositions depends on the type of condition being treated. If the condition consists of an acute or subacute disorder, a treatment with an immediate release form will be preferred over a prolonged release composition. Alternatively, for preventative or long-term treatments, a sustained released composition will generally be preferred.

The protein or candidate compound of the present invention can be prepared in any suitable manner. The protein or candidate compound can be isolated from naturally occur-

ring sources, recombinantly produced, or produced synthetically, or produced by a combination of these methods. The synthesis of short peptides is well known in the art. See e.g. Stewart et al., Solid Phase Peptide Synthesis (Pierce Chemical Co., 2d ed., 1984).

Gene Therapy

Another example of how Dragon family polynucleotides of the invention can be effectively used in treatment is gene therapy. See, generally, for example, U.S. Pat. No. 5,399,346. The general principle is to introduce the polynucleotide, for example, a Dragon gene, into a target cell in a patient, and allow it to supplement the activity of the defective endogenous Dragon protein. Alternatively, a Dragon gene can be inserted into an embryonic or adult stem or progenitor cell to promote cell survival or induce differentiation into a particular cell fate.

Entry into the cell is facilitated by suitable techniques known in the art such as providing the polynucleotide in the form of a suitable vector, or encapsulation of the polynucleotide in a liposome.

A desired mode of gene therapy is to provide the polynucleotide in such a way that it will replicate inside the cell,

enhancing and prolonging the desired effect. Thus, the polynucleotide is operably linked to a suitable promoter, such as the natural promoter of the corresponding gene, a heterologous promoter that is intrinsically active in neuronal, bone, muscle, skin, joint, or cartilage cells, or a heterologous promoter that can be induced by a suitable agent.

OTHER EMBODIMENTS

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims. Other embodiments are within the claims.

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<211> LENGTH: 436
<212> TYPE: PRT
<213> ORGANISM: Mus musculus

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Gly Asp Cys Gln Gln Pro Thr Gln Cys Arg Ile Gln Lys Cys Thr Thr
50 55 60
Asp Phe Val Ala Leu Thr Ala His Leu Asn Ser Ala Ala Asp Gly Phe
65 70 75 80
Asp Ser Glu Phe Cys Lys Ala Leu Arg Ala Tyr Ala Gly Cys Thr Gln
85 90 95
Arg Thr Ser Lys Ala Cys Arg Gly Asn Leu Val Tyr His Ser Ala Val
100 105 110
Leu Gly Ile Ser Asp Leu Met Ser Gln Arg Asn Cys Ser Lys Asp Gly
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His Ser His Gly Gly Val Arg Glu His Gly Gly Gly Asp Gln Arg Pro

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		195					200					205								
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	210					215					220									
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225					230					235					240					
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	370					375				380										
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Gly	Cys	Arg	Asp	Leu	Pro	Val	Gly	Leu	Gly	Leu	Thr	Cys	Leu	Ile	Leu					
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			20					25					30							
Gln	Cys	Lys	Ile	Leu	Arg	Cys	Asn	Ala	Glu	Tyr	Val	Ser	Ser	Thr	Leu					
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Arg	Leu	Arg	Gly	Gly	Gly	Ser	Pro	Asp	Thr	Pro	Arg	Gly	Gly	Gly	Arg					
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				245					250					255		
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			340					345					350			
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		355					360					365				
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		20					25						30			
Thr	Leu	Ala	Phe	Leu	Leu	Cys	Ser	Phe	Pro	Ala	Ala	Ile	Ser	Pro	Cys	
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Lys	Ile	Leu	Lys	Cys	Asn	Ser	Glu	Phe	Trp	Ser	Ala	Thr	Ser	Ser	Gly	
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Asp	Leu	Ala	Tyr	His	Ser	Ala	Val	His	Gly	Ile	Glu	Asp	Leu	Met	Ser
			100					105					110		
Gln	His	Asn	Cys	Ser	Lys	Asp	Gly	Pro	Thr	Ser	Gln	Pro	Arg	Val	Arg
		115					120					125			
Thr	Leu	Pro	Pro	Ala	Gly	Asp	Ser	Gln	Glu	Arg	Ser	Asp	Ser	Pro	Glu
	130					135					140				
Ile	Cys	His	Tyr	Glu	Lys	Ser	Phe	His	Lys	His	Ser	Ala	Ala	Pro	Asn
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Tyr	Thr	His	Cys	Gly	Leu	Phe	Gly	Asp	Pro	His	Leu	Arg	Thr	Phe	Thr
				165					170					175	
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			180					185					190		
Asn	Asn	Tyr	Leu	Asn	Val	Gln	Val	Thr	Asn	Thr	Pro	Val	Leu	Pro	Gly
		195					200					205			
Ser	Ala	Ala	Thr	Ala	Thr	Ser	Lys	Thr	Leu	Ala	Thr	Val	Leu	Gly	Pro
	210					215					220				
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225					230					235					240
Lys	Val	Tyr	Gln	Ala	Glu	Met	Asp	Glu	Leu	Pro	Ser	Ala	Phe	Ala	Asp
				245					250					255	
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			260					265					270		
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Gln	Gly	Leu	Tyr	Leu	Cys	Leu	Arg	Gly	Cys	Pro	Leu	Asn	Gln	Gln	Ile
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Ala	Val	Ala	Lys	Cys	Lys	Glu	Lys	Leu	Pro	Val	Glu	Asp	Leu	Tyr	Tyr
	370					375					380				
Gln	Ala	Cys	Val	Phe	Asp	Leu	Leu	Thr	Thr	Gly	Asp	Val	Asn	Phe	Thr
385					390					395					400
Leu	Ala	Ala	Tyr	Tyr	Ala	Leu	Glu	Asp	Gly	Lys	Met	Leu	His	Ser	Asn
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Lys	Asp	Lys	Leu	His	Leu	Phe	Glu	Arg	Thr	Arg	Glu	Leu	Pro	Gly	Ala
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		435					440					445			
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<212> TYPE: PRT																
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			20					25					30			
Leu	Leu	Leu	Leu	Leu	Phe	Ser	Leu	Gly	Leu	Leu	His	Ala	Gly	Asp	Cys	
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Ser	Leu	Thr	Ser	His	Leu	Asn	Ser	Ala	Val	Asp	Gly	Phe	Asp	Ser	Glu	
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Phe	Cys	Lys	Ala	Leu	Arg	Ala	Tyr	Ala	Gly	Cys	Thr	Gln	Arg	Thr	Ser	
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Lys	Ala	Cys	Arg	Gly	Asn	Leu	Val	Tyr	His	Ser	Ala	Val	Leu	Gly	Ile	
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Ser	Thr	Asn	Pro	Glu	Val	Thr	His	Asp	Pro	Cys	Asn	Tyr	His	Ser	His	
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Ala	Gly	Ala	Arg	Glu	His	Arg	Arg	Gly	Asp	Gln	Asn	Pro	Pro	Ser	Tyr	
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		180						185					190			
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	210					215					220					
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225					230					235				240		
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Ala	Arg	Tyr	Ile	Gly	Thr	Thr	Val	Phe	Val	Arg	Gln	Val	Gly	Arg	Tyr	
	275						280					285				
Leu	Thr	Leu	Ala	Ile	Arg	Met	Pro	Glu	Asp	Leu	Ala	Met	Ser	Tyr	Glu	
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Glu	Ser	Gln	Asp	Leu	Gln	Leu	Cys	Val	Asn	Gly	Cys	Pro	Leu	Ser	Glu	
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Arg	Ile	Asp	Asp	Gly	Gln	Gly	Gln	Val	Ser	Ala	Ile	Leu	Gly	His	Ser	
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Thr	Ala	Asn	Thr	Gln	Cys	His	Glu	Lys	Met	Pro	Val	Lys	Asp	Ile	Tyr	
	355						360					365				
Phe	Gln	Ser	Cys	Val	Phe	Asp	Leu	Leu	Thr	Thr	Gly	Asp	Ala	Asn	Phe	
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Thr	Ala	Ala	Ala	His	Ser	Ala	Leu	Glu	Asp	Val	Glu	Ala	Leu	His	Pro	
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Arg	Lys	Glu	Arg	Trp	His	Ile	Phe	Pro	Ser	Ser	Gly	Asn	Gly	Thr	Pro
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Gly	Gly	Gly	Gly	Gly	Arg	Gly	Gly	Gly	Val	Gly	Ser	Gly	Gly	Leu	Cys
65					70				75					80	
Arg	Ala	Leu	Arg	Ser	Tyr	Ala	Leu	Cys	Thr	Arg	Arg	Thr	Ala	Arg	Thr
				85					90					95	
Cys	Arg	Gly	Asp	Leu	Ala	Phe	His	Ser	Ala	Val	His	Gly	Ile	Glu	Asp
			100					105					110		
Leu	Met	Ile	Gln	His	Asn	Cys	Ser	Arg	Gln	Gly	Pro	Thr	Ala	Pro	Pro
			115				120					125			
Pro	Pro	Arg	Gly	Pro	Ala	Leu	Pro	Gly	Ala	Gly	Ser	Gly	Leu	Pro	Ala
			130				135				140				
Pro	Asp	Pro	Cys	Asp	Tyr	Glu	Gly	Arg	Phe	Ser	Arg	Leu	His	Gly	Arg
145					150					155					160
Pro	Pro	Gly	Phe	Leu	His	Cys	Ala	Ser	Phe	Gly	Asp	Pro	His	Val	Arg
				165					170					175	
Ser	Phe	His	His	His	Phe	His	Thr	Cys	Arg	Val	Gln	Gly	Ala	Trp	Pro
			180					185					190		
Leu	Leu	Asp	Asn	Asp	Phe	Leu	Phe	Val	Gln	Ala	Thr	Ser	Ser	Pro	Met
			195				200					205			
Ala	Leu	Gly	Ala	Asn	Ala	Thr	Ala	Thr	Arg	Lys	Leu	Thr	Ile	Ile	Phe
			210				215					220			
Lys	Asn	Met	Gln	Glu	Cys	Ile	Asp	Gln	Lys	Val	Tyr	Gln	Ala	Glu	Val
225					230					235					240
Asp	Asn	Leu	Pro	Val	Ala	Phe	Glu	Asp	Gly	Ser	Ile	Asn	Gly	Gly	Asp
				245					250					255	
Arg	Pro	Gly	Gly	Ser	Ser	Leu	Ser	Ile	Gln	Thr	Ala	Asn	Pro	Gly	Asn
			260					265					270		
His	Val	Glu	Ile	Gln	Ala	Ala	Tyr	Ile	Gly	Thr	Thr	Ile	Ile	Ile	Arg
			275				280					285			
Gln	Thr	Ala	Gly	Gln	Leu	Ser	Phe	Ser	Ile	Lys	Val	Ala	Glu	Asp	Val
			290				295				300				
Ala	Met	Ala	Phe	Ser	Ala	Glu	Gln	Asp	Leu	Gln	Leu	Cys	Val	Gly	Gly
305					310					315				320	
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325																330																335															
Ala	Ile	Thr	Ile	Asp	Thr	Ala	Arg	Arg	Leu	Cys	Lys	Glu	Gly	Leu	Pro																																
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Val	Glu	Asp	Ala	Tyr	Phe	His	Ser	Cys	Val	Phe	Asp	Val	Leu	Ile	Ser																																
			355				360						365																																		
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			370				375						380																																		
Arg	Ala	Phe	Leu	Pro	Asp	Leu	Glu	Lys	Leu	His	Leu	Phe	Pro	Ser	Asp																																
385						390						395																																			
Ala	Gly	Val	Pro	Leu	Ser	Ser	Ala	Thr	Leu	Leu	Ala	Pro	Leu	Leu	Ser																																
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Gly	Leu	Phe	Val	Leu	Trp	Leu	Cys	Ile	Gln																																						
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Met	Gly	Met	Gly	Arg	Gly	Ala	Gly	Arg	Ser	Ala	Leu	Gly	Phe	Trp	Pro																																
			20					25					30																																		
Thr	Leu	Ala	Phe	Leu	Leu	Cys	Ser	Phe	Pro	Ala	Ala	Thr	Ser	Pro	Cys																																
			35					40					45																																		
Lys	Ile	Leu	Lys	Cys	Asn	Ser	Glu	Phe	Trp	Ser	Ala	Thr	Ser	Gly	Ser																																
50					55						60																																				
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65						70						75			80																																
Ser	Tyr	Ala	Leu	Cys	Thr	Arg	Arg	Thr	Ala	Arg	Thr	Cys	Arg	Gly	Asp																																
			85					90					95																																		
Leu	Ala	Tyr	His	Ser	Ala	Val	His	Gly	Ile	Glu	Asp	Leu	Met	Ser	Gln																																
			100					105					110																																		
His	Asn	Cys	Ser	Lys	Asp	Gly	Pro	Thr	Ser	Gln	Pro	Arg	Leu	Arg	Thr																																
			115					120					125																																		
Leu	Pro	Pro	Ala	Gly	Asp	Ser	Gln	Glu	Arg	Ser	Asp	Ser	Pro	Glu	Ile																																
130						135						140																																			
Cys	His	Tyr	Glu	Lys	Ser	Phe	His	Lys	His	Ser	Ala	Thr	Pro	Asn	Tyr																																
145						150						155			160																																
Thr	His	Cys	Gly	Leu	Phe	Gly	Asp	Pro	His	Leu	Arg	Thr	Phe	Thr	Asp																																
			165					170					175																																		
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			180					185					190																																		
Asn	Tyr	Leu	Asn	Val	Gln	Val	Thr	Asn	Thr	Pro	Val	Leu	Pro	Gly	Ser																																
			195					200					205																																		
Ala	Ala	Thr	Ala	Thr	Ser	Lys	Leu	Thr	Ile	Ile	Phe	Lys	Asn	Phe	Gln																																
210						215						220																																			
Glu	Cys	Val	Asp	Gln	Lys	Val	Tyr	Gln	Ala	Glu	Met	Asp	Glu	Leu	Pro																																
225						230						235			240																																
Ala	Ala	Phe	Val	Asp	Gly	Ser	Lys	Asn	Gly	Gly	Asp	Lys	His	Gly	Ala																																
			245					250					255																																		
Asn	Ser	Leu	Lys	Ile	Thr	Glu	Lys	Val	Ser	Gly	Gln	His	Val	Glu	Ile																																
			260					265					270																																		

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Gln	Ala	Lys	Tyr	Ile	Gly	Thr	Thr	Ile	Val	Val	Arg	Gln	Val	Gly	Arg
		275					280					285			
Tyr	Leu	Thr	Phe	Ala	Val	Arg	Met	Pro	Glu	Glu	Val	Val	Asn	Ala	Val
	290					295					300				
Glu	Asp	Trp	Asp	Ser	Gln	Gly	Leu	Tyr	Leu	Cys	Leu	Arg	Gly	Cys	Pro
305					310					315					320
Leu	Asn	Gln	Gln	Ile	Asp	Phe	Gln	Ala	Phe	His	Thr	Asn	Ala	Glu	Gly
				325					330					335	
Thr	Gly	Ala	Arg	Arg	Leu	Ala	Ala	Ala	Ser	Pro	Ala	Pro	Thr	Ala	Pro
			340					345					350		
Glu	Thr	Phe	Pro	Tyr	Glu	Thr	Ala	Val	Ala	Lys	Cys	Lys	Glu	Lys	Leu
	355						360					365			
Pro	Val	Glu	Asp	Leu	Tyr	Tyr	Gln	Ala	Cys	Val	Phe	Asp	Leu	Leu	Thr
	370					375					380				
Thr	Gly	Asp	Val	Asn	Phe	Thr	Leu	Ala	Ala	Tyr	Tyr	Ala	Leu	Glu	Asp
385					390					395					400
Val	Lys	Met	Leu	His	Ser	Asn	Lys	Asp	Lys	Leu	His	Leu	Tyr	Glu	Arg
				405					410					415	
Thr	Arg	Asp	Leu	Pro	Gly	Arg	Ala	Ala	Ala	Gly	Leu	Pro	Leu	Ala	Pro
			420					425					430		
Arg	Pro	Leu	Leu	Gly	Ala	Leu	Val	Pro	Leu	Leu	Ala	Leu	Leu	Pro	Val
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Phe	Cys														
	450														
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<220> FEATURE:															
<223> OTHER INFORMATION: Synthetic															
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Arg Lys															
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<223> OTHER INFORMATION: Synthetic															
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1					5				10					15	
Leu	Gly	Ala	Ser	Gly	Gly	Gly	Gly	Gly	Ala	Arg	Ala	Glu	Val	Leu	Phe
			20					25					30		
Arg	Cys	Pro	Pro	Cys	Thr	Pro	Glu	Arg	Leu	Ala	Ala	Cys	Gly	Pro	Pro
		35					40					45			
Pro	Val	Ala	Pro	Pro	Ala	Ala	Val	Val	Met	Gly	Glu	Gly	Thr	Cys	Glu
	50					55					60				
Asp	Arg	Arg	Asp	Ala	Glu	Tyr	Gly	Ala	Ser	Pro	Glu	Gln	Val	Ala	Asp
65					70				75						80
Asn	Gly	Asp	Asp	His	Ser	Glu	Gly	Gly	Leu	Val	Glu	Asn			
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<210> SEQ ID NO 13
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic

<400> SEQUENCE: 13

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1 5 10 15

Leu Leu Pro Leu Leu Leu Pro Ala Gly Cys Arg Ala Leu Glu Glu
20 25 30

Thr Leu Met Asp Thr Lys Trp Val Thr Ser Glu Leu Ala Trp Thr Ser
35 40 45

His Pro Glu Ser Gly Trp Glu Glu Val Ser Gly Tyr Asp Glu Ala Met
50 55 60

Asn Pro Ile Arg Thr Tyr Gln Val Cys Asn Val Arg Glu Ser
65 70 75

<210> SEQ ID NO 14
<211> LENGTH: 148
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic

<400> SEQUENCE: 14

Pro Gly Ala Arg Gly Arg Arg Arg Arg Arg Arg Pro Met Ser Pro Pro
1 5 10 15

Pro Pro Pro Pro Pro Val Arg Ala Leu Pro Leu Leu Leu Leu Leu Ala
20 25 30

Gly Pro Gly Ala Ala Ala Pro Pro Cys Leu Asp Gly Ser Pro Cys Ala
35 40 45

Asn Gly Gly Arg Cys Thr Gln Leu Pro Ser Arg Glu Ala Ala Cys Leu
50 55 60

Cys Pro Pro Gly Trp Val Gly Glu Arg Cys Gln Leu Glu Asp Pro Cys
65 70 75 80

His Ser Gly Pro Cys Ala Gly Arg Gly Val Cys Gln Ser Ser Val Val
85 90 95

Ala Gly Thr Ala Arg Phe Ser Cys Arg Cys Pro Arg Gly Phe Arg Gly
100 105 110

Pro Asp Cys Ser Leu Pro Asp Pro Cys Leu Ser Ser Pro Cys Ala His
115 120 125

Gly Ala Arg Cys Ser Val Gly Pro Asp Gly Arg Phe Leu Cys Ser Cys
130 135 140

Pro Pro Gly Tyr
145

<210> SEQ ID NO 15
<211> LENGTH: 166
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic

<400> SEQUENCE: 15

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1 5 10 15

Leu Lys Arg Asp Val Glu Phe Leu Ala Gln Leu Lys Ile Met Asp Tyr

[illegible]

<400> SEQUENCE: 16

[illegible]

<400> SEQUENCE: 17

Glu	Pro	Cys	His	Lys	Lys	Val	Cys	Ala	His	Gly	Met	Cys	Gln	Pro	Ser
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Ser	Gln	Ser	Gly	Phe	Thr	Cys	Glu	Cys	Glu	Glu	Gly	Trp	Met	Gly	Pro
			20					25					30		
Leu	Cys	Asp	Gln	Arg	Thr	Asn	Asp	Pro	Cys	Leu	Gly	Asn	Lys	Cys	Val

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35					40					45										
His Gly Thr Cys Leu Pro Ile Asn Ala Phe Ser Tyr Ser Cys Lys Cys																				
50						55						60								
Leu Glu Gly His Gly Gly Val Leu Cys Asp Glu Glu Glu Asp Leu Phe																				
65						70						75								
Asn Pro Cys Gln Met Ile Lys Cys Lys His Gly Lys Cys Arg Leu Ser																				
					85						90									
Gly Val Gly Gln Pro Tyr Cys Glu Cys Asn Ser Gly Phe Thr Gly Asp																				
					100						105									
															110					
<210> SEQ ID NO 18																				
<211> LENGTH: 420																				
<212> TYPE: PRT																				
<213> ORGANISM: Canhorbitidtis elegans																				
<400> SEQUENCE: 18																				
Met Arg Arg His Trp Lys Glu Phe Glu Cys Glu Lys Trp Glu Ser Cys																				
1						5						10								
Asn Asp Asn Ser His Val Lys Arg Lys His Val Asn Thr Gly His Ile																				
					20						25									
Cys Gly Gly Lys Phe Glu Leu Ser Glu Lys Asn Leu Ala Ala Lys Phe																				
					35						40									
Lys Tyr Ser Gly Asp Thr Val Trp Arg Gly Arg Pro Asn Phe Leu Lys																				
					50						55									
Ser Leu Cys Tyr Phe Asn Pro Pro Pro Ser Asn Arg Lys Leu Lys Tyr																				
65						70						75								
Cys Ser Leu Phe Gly Asp Pro His Leu Ile Met Phe Asn Gly Ser Val																				
					85						90									
Gln Thr Cys Ser Glu Glu Gly Ala Arg Pro Leu Val Asp Asn Arg Tyr																				
					100						105									
Phe Leu Val Gln Val Thr Asn Arg Asn Val Arg Gly Glu Ala Leu Thr																				
					115						120									
Thr Thr Val Thr Lys Val Thr Val Leu Val Arg Lys His Asn Cys Thr																				
					130						135									
Ala Ser Leu Arg Tyr Glu Ala Ser Ser Asp Glu Glu Gly Leu Pro Arg																				
					145						150									
Gly Phe Val Asp Gly Thr Thr Phe Gln Met Thr Ser Lys His Ser Val																				
					165						170									
Glu Val Leu Trp Gln Asp Asp Asn Tyr Val Glu Ile Ala Leu His Phe																				
					180						185									
Ile His Ser Ser Ile His Ile Arg Arg Gln Gly Pro Tyr Leu Ser Val																				
					195						200									
Ser Val Arg Ala Pro Thr Ile Val Leu Glu Thr Gly Gly Asp Val Ala																				
					210						215									
Arg Glu Leu Cys Trp Ser Gly Cys Arg Lys Ser Ser Arg Ile Pro Ala																				
					225						230									
Glu Leu Ala Val Glu Met Thr Lys Lys Phe Ala Glu Cys Tyr Arg Arg																				
					245						250									
Arg Val His Val Pro Lys Lys Val Ala Glu Glu Thr Thr Phe Leu Ser																				
					260						265									
Glu Gln Lys Val Leu Pro Ile Tyr Asp Arg Cys Lys Asp Ile Gly Asn																				
					275						280									
Ile Gly Val Phe Phe Asp Ala Ser Ala Arg Lys Ile Leu Asn Phe Arg																				
					290						295									

[illegible]

<400> SEQUENCE: 19

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<400> SEQUENCE: 20

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<400> SEQUENCE: 21

ttgccatcct ccaaagcata gtaggcagcc agcgtgaagt tcacatca 48

<400> SEQUENCE: 22

ataagcttat gggcgtgaga gcagcacctt cc 32

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<210> SEQ ID NO 23
<211> LENGTH: 29
<212> TYPE: DNA
<213> ORGANISM: Mus musculus
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Primer
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<400> SEQUENCE: 23	
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<212> TYPE: DNA	
<213> ORGANISM: Mus musculus	
<220> FEATURE:	
<223> OTHER INFORMATION: Synthetic Primer	
<400> SEQUENCE: 24	
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ctacatctac tagtgctgtg caccctctcc tccctcactc ccataggtga gagtcagggt	120
cagactcctc agtgccgcat ccagaagtgc actaccgact tcgtttctct gacgtcccat	180
ctgaacccat cactggatgg ctttgatacg gagttctgca aggcgctgcg agcctattcg	240
gcctgtacgc agcgtacagc caagagctgc agggggaacc tggctctcca ctccgccatg	300
ctgggcatca ctgaccttat gagccagagg aactgctcca aagacgggcc cacgtcctcc	360
acccatcccc tcatccctat cgagccttgc aactatcaca gccggcatca ccaccacgtg	420
tcgcggttcg ggacgggggt gccgaacac cctcgtctga tgtacctgtt ctgtggcctg	480
ttcggggacc ctcatcttag gacttttaaa gaccagtttc aaacgtgtaa agttgaagg	540
gcttggcctc tcattgataa caactacctg tcagtgcagg tctaataatg tccagtgggt	600
tatggatcca gtgccacagc taccaataag atcacaataa tcttcaaacc ataccaagaa	660
tgcacagacc agaaggtcta ccaggccgtg acagacgacc ttccagccgc cttcgtagac	720
ggcaccatca gcggagggtga cagtgcagacc cgcagcatct ggatcctgga gaaatctccc	780
ggtcggcatg tagaaatcca cgctgcgtac atcgggggtca ccatcatcat acgccagcag	840
ggccggttacc tgacactagc tgtgcgaatg cctgaggaac tggccatggc ctttgatgaa	900
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ggacacctcc agctgcccgt gcttggcctc cagcaggctg gctttcagca gcagcagcag	1020
cccagggtgg aagcccagag aggcgtcttc actcttgaaa gtgcctccag gaggtgcagg	1080
gaccaactgg aggtgaagga catctatttc cactcctgtg tgtttgacct gctcactaca	1140
ggagatgcca acttcaccac tgccgcctac aatgccctga aagacatgga gacactgcat	1200
cccaaaaagg agcgtggca gattttcccc aactcggctt ccaggctgag tcctttttca	1260
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<211> LENGTH: 1233	
<212> TYPE: DNA	
<213> ORGANISM: Danio rerio	
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ggcaataaag	aggggtgtaaa	tacgggatac	tgcagcgcgc	tgcgatccta	cgccctctgc	240
acccagcggg	cggcccgcgc	ctgcagggga	gaccttgctg	atcactctgc	tgtacagggc	300
atcgaggacc	tgctcataca	ataccgctgc	ccaaaagcag	gccccacggc	tcagcctcag	360
cctcggtccc	tgccccaaag	ccctctctcc	ggggatggat	gcttctacga	gaagggtttt	420
atacagaggg	agggccgggc	cccagaatat	ctacactgtg	gggtgttttg	agaccgcgat	480
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gataaccagt	atctgtacat	ccaggccacc	agcagtccca	ccagggagag	ctctgacacc	600
accataactc	cagaggtcac	ggtgatcttc	cagaactggc	gtgagtgcgc	tgaacagcag	660
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ggggaccgca	gagggcatca	gagtctgcgt	attcactcac	aagaccagag	ccggcatgct	780
gaaatctggg	ctactcatat	tgggaccatg	ataatagtgc	ggcaggttgg	ccagtcgctg	840
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<213> ORGANISM: Danio rerio						
<400> SEQUENCE: 27						
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ttctgggcct	ccacctccaa	ttctggccca	gaagaggagt	tttgccactgc	tctgcgagca	180
tacaacagtt	gtgtacgtcg	caccgctcgt	acatgccggg	gtgacctggc	ctaccattca	240
gccaacatg	gtatagagga	tcttatgagc	caacacaact	gctccaaaga	agggccact	300
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catattccct	ctgacgagcc	agaggatatg	cattatgagc	ggagtctacc	acgtaatgct	420
gcacctocca	attacacca	ttgtggcttt	tttggggatc	cacatcttcg	cacattcaac	480
gatgactttc	aaacctgtaa	agttgaggga	gcctggccac	ttatccacaa	taagtacctg	540
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ctaacaatca	tcttcaacag	ctttcaagaa	tgcgtagacc	agaagacgta	ccgtgcagag	660
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tatattggca	caacaattgt	agttcggaaa	gttggccact	accttacctt	tgctgtgcgg	840
atgccagagg	aggtggtaaa	ttcagtggaa	gaccaggaca	atcaggatct	ttacctctgc	900
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<211> LENGTH: 436
<212> TYPE: PRT
<213> ORGANISM: Danio rerio

<400> SEQUENCE: 28

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Lys Cys Thr Thr Asp Phe Val Ser Leu Thr Ser His Leu Asn Pro Ser
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Ala Cys Thr Gln Arg Thr Ala Lys Ser Cys Arg Gly Asn Leu Val Phe
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His Ser Ala Met Leu Gly Ile Thr Asp Leu Met Ser Gln Arg Asn Cys
100 105 110
Ser Lys Asp Gly Pro Thr Ser Ser Thr His Pro Val Ile Pro Ile Glu
115 120 125
Pro Cys Asn Tyr His Ser Arg His His His His Val Ser Arg Phe Gly
130 135 140
Thr Gly Val Pro Glu His Pro Arg Leu Met Tyr Leu Phe Cys Gly Leu
145 150 155 160
Phe Gly Asp Pro His Leu Arg Thr Phe Lys Asp Gln Phe Gln Thr Cys
165 170 175
Lys Val Glu Gly Ala Trp Pro Leu Ile Asp Asn Asn Tyr Leu Ser Val
180 185 190
Gln Val Thr Asn Val Pro Val Val Tyr Gly Ser Ser Ala Thr Ala Thr
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Asn Lys Ile Thr Ile Ile Phe Lys Pro Tyr Gln Glu Cys Thr Asp Gln
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Lys Val Tyr Gln Ala Val Thr Asp Asp Leu Pro Ala Ala Phe Val Asp
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Gly Thr Ile Ser Gly Gly Asp Ser Glu Thr Arg Ser Ile Trp Ile Leu
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Glu Lys Ser Pro Gly Arg His Val Glu Ile His Ala Ala Tyr Ile Gly
260 265 270
Val Thr Ile Ile Ile Arg Gln Gln Gly Arg Tyr Leu Thr Leu Ala Val
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Phe	Asp	Pro	Ala	Tyr	Thr	His	Cys	Ser	Ser	Leu	Phe	Pro	Gly	Arg	Asp	
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385					390					395					400
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<213> ORGANISM: Homo sapiens

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- What is claimed is:
1. An isolated DRG-11-Responsive Axonal Guidance and Outgrowth of Neurite (DRAGON) protein comprising the amino acid sequence of SEQ ID NO:8, wherein said protein promotes the adhesion of cultured dorsal root ganglion (DRG) neurons.
2. The DRAGON protein of claim 1, wherein said protein comprises the amino acid sequence of SEQ ID NO:8.
3. An isolated DRAGON protein comprising an amino acid sequence at least 95% identical to SEQ ID NO:8 and lacking the C-terminal GPI anchoring domain of said DRAGON protein, wherein said protein promotes the adhesion of cultured dorsal root ganglion (DRG) neurons.
4. An isolated DRAGON protein comprising the amino acid sequence of SEQ ID NO:8 with the exception that the C-terminal GPI anchoring domain is deleted.
5. An isolated DRAGON protein comprising an amino acid sequence at least 95% identical to SEQ ID NO:5, wherein said protein promotes the adhesion of cultured dorsal root ganglion (DRG) neurons.
6. The DRAGON protein of claim 5, wherein said protein comprises the amino acid sequence of SEQ ID NO:5.
7. The DRAGON protein of claim 6, wherein said protein consists of the amino acid sequence of SEQ ID NO:5.
8. The DRAGON protein of claim 5 lacking the C-terminal GPI anchoring domain of said DRAGON protein.
9. An isolated DRAGON protein comprising the amino acid sequence of SEQ ID NO:5 with the exception that the C-terminal GPI anchoring domain is deleted.
- * * * * *